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## (54) Title: MORPHOGENIC PROTEIN-SPECIFIC CELL SURFACE RECEPTORS AND USES THEREFOR

## (57) Abstract

Disclosed are: (1) nucleic acid sequences, amino acid sequences, homologies, structural features and various other data characterizing a morphogen cell surface receptors particularly OP-1-binding cell surface receptors; (2) methods for producing receptor proteins, including fragments thereof, using recombinant DNA technology; (3) methods for identifying novel morphogen receptors and their encoding DNAs; (4) methods and compositions for identifying compounds capable of modulating endogenous morphogen receptor levels; and (5) methods and compositions for identifying morphogen receptor binding analogs useful in the design of morphogen agonists and antagonists for therapeutic, diagnostic and experimental uses.

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# "MORPHOGENIC PROTEIN-SPECIFIC CELL SURFACE RECEPTORS AND USES THEREFOR"

### Field of the Invention

This invention relates generally to the field of tissue morphogenesis and more particularly to morphogenic protein-specific cell surface receptors.

#### 5 Background of the Invention.

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Cell differentiation is the central characteristic of tissue morphogenesis which initiates during embryogenesis, and continues to various degrees throughout the life of an organism in adult tissue repair and regeneration mechanisms. The degree of morphogenesis in adult tissue varies among different tissues and is related, among other things, to the degree of cell turnover in a given tissue.

The cellular and molecular events which govern the stimulus for differentiation of cells is an area of intensive research. In the medical and veterinary fields, it is anticipated that the discovery of the factor or factors which control cell differentiation and tissue morphogenesis will advance significantly medicine's ability to repair and regenerate diseased or damaged mammalian tissues and organs. Particularly useful areas for human and veterinary therapeutics include reconstructive surgery and in the treatment of tissue degenerative diseases including arthritis, emphysema, osteoporosis, cardiomyopathy, cirrhosis, degenerative nerve diseases, inflammatory diseases, and cancer, and in the regeneration of tissues, organs and limbs. (In this and related applications, the terms "morphogenetic" and "morphogenic" are used interchangeably.)

A number of different factors have been isolated in recent years which appear to play a role in cell differentiation. Recently, a distinct subfamily of the "superfamily" of structurally related proteins referred to in the art as the "transforming growth factor-b (TGF-B) superfamily of proteins have been identified as true tissue morphogens.

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The members of this distinct "subfamily" of true tissue morphogenic proteins share substantial amino acid sequence homology within their morphogenetically active C-terminal domains (at least 50% identity in the Cterminal 102 amino acid sequence), including a conserved six or seven cysteine skeleton, and share the in vivo activity of inducing tissue-specific morphogenesis in a variety of organs and tissues. The proteins apparently contact and interact with progenitor cells e.g., by binding suitable cell surface molecules, predisposing or otherwise stimulating the cells to proliferate and differentiate in a morphogenetically permissive environment. These morphogenic proteins are capable of inducing the developmental cascade of cellular and molecular events that culminate in the formation of new organspecific tissue, including any vascularization, connective tissue formation, and nerve innervation as required by the naturally occurring tissue. The proteins have been shown to induce morphogenesis of both bone cartilage and bone, as well as periodontal tissues, dentin, liver, and neural tissue, including retinal tissue.

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The true tissue morphogenic proteins identified to date include proteins originally identified as bone inductive proteins. These include OP-1, (osteogenic protein-1, also referred to in related applications as \*OP1\*), its Drosophila homolog, 60A, with which it shares 69% identity in the C-terminal "seven cysteine" domain, and the related proteins OP-2 (also referred to in related applications as \*OP2\*) and OP-3, both of which share approximately 70-75% identity with OP-1 in the C-terminal seven cysteine domain, as well as BMP5, BMP6 and its murine homolog, Vgr-1, all of which share greater than 85% identity with OP-1 in the C-terminal seven cysteine domain, and the BMP6 Xenopus homolog, Vgl, which shares approximately 57% identity with OP-1 in the C-terminal seven cysteine domain. Other bone inductive proteins include the CBMP2 proteins (also referred to in the art as BMP2 and BMP4) and their Drosophila homolog, DPP. Another tissue morphogenic protein is GDF-1 (from mouse). See, for example, PCT documents US92/01968 and US92/07358, the disclosures of which are incorporated herein by reference.

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As stated above, these true tissue morphogenic proteins are recognized in the art as a distinct subfamily of proteins different from other members of the TGF-ß superfamily in that they share a high degree of sequence identity in the C-terminal domain and in that the true tissue morphogenic proteins are able to induce, on their own, the full cascade of events that result in formation of functional tissue rather than merely inducing formation of fibrotic (scar) tissue. Specifically, members of the family of morphogenic proteins are capable of all of the following in a morphogenetically permissive environment: stimulating cell proliferation and cell differentiation, and supporting the growth and maintenance of differentiated cells. The morphogenic proteins apparently may act as endocrine, paracrine or autocrine factors.

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The morphogenic proteins are capable of significant species

"crosstalk." That is, xenogenic (foreign species) homologs of these proteins
can substitute for one another in functional activity. For example, DPP and
60A; two Drosophila proteins, can substitute for their mammalian homologs,
BMP2/4 and OP-1, respectively, and induce endochondral bone formation at a
non-bony site in a standard rat bone formation assay. Similarly, BMP2 has
been shown to rescue a dpp mutation in Drosophila. In their native form,
however, the proteins appear to be tissue-specific, each protein typically
being expressed in or provided to one or only a few tissues or, alternatively,
expressed only at particular times during development. For example, GDF-1
appears to be expressed primarily in neural tissue, while OP-2 appears to be
expressed at relatively high levels in early (e.g., 8-day) mouse embryos. The
endogenous morphogens may be synthesized by the cells on which they act, by
neighboring cells, or by cells of a distant tissue, the secreted protein being
transported to the cells to be acted on.

A particularly potent tissue morphogenic protein is OP-1. This protein, and its xenogenic homologs, are expressed in a number of tissues, primarily in tissues of urogenital origin, as well as in bone, mammary and salivary gland tissue, reproductive tissues, and gastrointestinal tract

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tissue. It is also expressed in different tissues during embryogenesis, its presence coincident with the onset of morphogenesis of that tissue.

The morphogenic protein signal transduction across a cell membrane appears to occur as a result of specific binding interaction with one or more cell surface receptors. Recent studies on cell surface receptor binding of various members of the TGF-ß protein superfamily suggests that the ligands can mediate their activity by interaction with two different receptors, referred to as Type I and Type II receptors to form a hetero-complex. A cell surface bound beta-glycan also may enhance the binding interaction. The Type I and Type II receptors are both serine /threonine kinases, and share similar structures: an intracellular domain that consists essentially of the kinase, a short, extended hydrophobic sequence sufficient to span the membrane one time, and an extracellular domain characterized by a high concentration of conserved cysteines.

These include "TGF-&R II", a TGF-& Type II receptor (Lin et al. (1992) Cell 68:775-785); and numerous activin-binding receptors. See, for example, Mathews et al. (1991) Cell 65:973-982 and international patent application WO 92/20793, published November 26, 1992, disclosing the "ActR II" sequence; Attisano et al., (1992) Cell 68:97-108, disclosing the "ActR-IIB" sequence; and Legerski et al. (1992) Biochem Biophys. Res. Commun 183:672-679. A different Type II receptor shown to have affinity for activin is Atr-II (Childs et al.(1993) PNAS 90:9475-9479.) Two Type II receptors have been identified in C. elegans, the daf-1 gene, (Georgi et al. (1990) Cell 61:635-645), having no known ligand to date, and daf-4, which has been shown to bind BMP4, but not activin or TGF-& (Estevez, et al. (1993) Nature 365:644-649.)

Ten Dijke et al. disclose the cloning of six different Type I cell surface receptors from murine and human cDNA libraries. ((1993) Oncogene 8:2879-2887, and Science (1994) 264:101-104. These receptors, referenced as ALK-1 to ALK-6 ("activin receptor-like kinases"), share significant sequence identities (60-79%) and several have been identified as TGF-ß binding (ALK-5) or activin binding (ALK-2, ALK-4) receptors. Xie et al. also report a

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Drosophila Type I receptor encoded by the sax gene (<u>Science</u> (1994) <u>263</u>:1756-1759). The authors suggest that the protein binds DPP.

To date, the Type I receptors with which the morphogenic proteins described herein interact on the cell surface have not yet been identified, and no Type II receptor has been described as having binding affinity for OP-1 and its related sequences. Identification of these cell surface molecules, with which the morphogens interact and through which they may mediate their biological effect, is anticipated to enhance elucidation of the molecular mechanism of tissue morphogenesis and to enable development of morphogen receptor binding "analogs", e.g., compounds (which may or may not be amino acid-based macromolecules) capable of mimicing the binding affinity of a morphogen for its receptor sufficiently to act either as a receptor binding agonist or antagonist. These "analogs" have particular utility in therapeutic, diagnostic and experimental research applications.

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It is an object of this invention to provide nucleic acid molecules and amino acid sequences encoding morphogenic protein binding cell surface receptors, particularly OP-1-specific binding receptor sequences. Another object is to provide methods for identifying genes in a variety of species and/or tissues, and in a variety of nucleic acid libraries encoding morphogenic protein binding receptors, particularly receptors that bind OP-1. Still another object is to provide means for designing biosynthetic receptorbinding ligand analogs, particularly OP-1 analogs, and/or for identifying natural-occurring ligand analogs, including agonists and antagonists, using the receptor molecules described herein, and analogs thereof. Another object is to provide antagonists, including soluble receptor constructs comprising the extracellular ligand-binding domain, which can modulate the availability of OP1 for receptor binding in vivo. Another obect is to provide means and compositions for competing with activin-receptor and BMP2/4-receptor interactions. Yet another object is to provide means and compositions for ligand affinity purification and for diagnostic detection and quantification of ligands in a body fluid using OP1-specific cell surface receptors and ligand-binding fragments thereof. Still another object is to provides means

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and compositions for modulating the endogenous expression or concentration of these receptor molecules. Yet another object is to provide ligand-receptor complexes and analog sequences thereof, as well as antibodies capable of identifying and distinguishing the complex from its component proteins. Still another object is to provide means and compositions for modulating a morphogenesis in a mammal. These and other objects and features of the invention will be apparent from the description, drawings and claims which follow.

## Summary of the Invention

Type I and Type II cell surface receptor molecules capable of specific binding affinity with true tissue morphogenic proteins, particularly OP-1-related proteins, now have been identified. Accordingly, the invention provides ligand-receptor complexes comprising at least the ligand binding domain of these receptors and OP-1 or an OP-1 receptor-binding analog as the ligand; means for identifying and/or designing useful OP-1 receptor-binding analogs and OP-1-binding- receptor analogs; and means for modulating the tissue morphogenesis capability of a cell.

The morphogen cell surface receptors useful in this invention are referred to in the art as Type I or Type II serine/threonine kinase receptors. They share a conserved structure, including an extracellular, ligand-binding domain generally composed of about 100-130 amino acids (Type I receptors; up to about 196 amino acids for Type II receptors), a transmembrane domain sufficient to span a cellular membrane one time, and an intracellular (cytoplasmic) domain having serine/threonine kinase activity. The intact receptor is a single polypeptide chain of about 500-550 amino acids and having an apparent molecular weight of about 50-55 kDa.

Of particular utility in the methods and compositions of the invention are the Type I cell surface receptors referenced herein and in the literature as, ALK-2, ALK-3 and ALK-6, whose nucleic acids and encoded amino acid sequences are represented by the sequences in Seq. ID Nos. 3, 5 and 7 respectively, and which, as demonstrated herein below, have specific binding affinity for OP1 and OP1-related analogs. Accordingly, in one embodiment, the

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receptor sequences contemplated herein include OP-1 binding analogs of the ALK-2, ALK-3 and ALK-6 proteins described herein.

As used herein, ligand-receptor binding specificity is understood to mean a specific, saturable noncovalent interaction between the ligand and the receptor, and which is subject to competitive inhibition by a suitable competitor molecule. Preferred binding affinities (defined as the amount of ligand required to fill one-half (50%) of available receptor binding sites) are described herein by dissociation constant (Kd). In one embodiment, preferred binding affinities of the ligand-receptor complexes described herein have a Kd of less than  $10^{-7}$ M, preferably less than  $5\times10^{-7}$ M, more preferably less than  $10^{-8}$ M. In another preferred embodiment, the receptor molecules have little or no substantial binding affinity for TGF-8.

As used herein, an "OP1-specific receptor analog" is understood to mean a sequence variant of the ALK-2, ALK-3 or ALK-6 sequences which shares at least 40%, preferably at least 45%, and most preferably at least 50%, amino acid identity in the extracellular ligand binding domain with the sequence defined by residues 23-122 of Seq. ID No. 7 (ALK-6), and which has substantially the same binding affinity for OP1 as ALK-2, ALK-3 or ALK-6. ALK-6 and ALK-3 share 46% amino acid sequence identity in their ligand binding domains.

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Accordingly, in one preferred embodiment, the OP1-specific receptor analogs share at least 46% amino acid sequence identity with the extracellular, ligand binding domains of ALK-6 or ALK-3.

As will be appreciated by those having ordinary skill in the art, OP1specific receptor analogs also can have binding affinity for other, related
morphogenic proteins. As used herein, an OP1-specific receptor analog is
understood to have substantially the same binding affinity for OP-1 as ALK-2,
ALK-3 or ALK-6 if it can be competed successfully for OP-1 binding in a
standard competition assay with a known OP-1 binding receptor, e.g., with ALK2, ALK-3 or ALK-6. In one preferred embodiment, OP1-specific receptor analogs
have a binding affinity for OP-1 defined by a dissociation constant of less
than about 10<sup>-7</sup> M, preferably less than about 5 x 10<sup>-7</sup>M or 10<sup>-8</sup> M. It is

anticipated however, that analogs having lower binding affinities, e.g., on the order of  $10^{-6} M$  also will be useful. For example, such analogs may be provided to an animal to modulate availability of serum-soluble OP1 for receptor binding in vivo. Similarly, where tight binding interaction is desired, for example as part of a cancer therapy wherein the analog acts as a ligand-receptor antagonist, preferred binding affinities may be on the order of  $5 \times 10^{-8} M$ .

In another embodiment, the OP-1 binding receptor analogs contemplated by the invention include proteins encoded by nucleic acids which hybridize with the DNA sequence encoding the extracellular, ligand binding domain of ALK-2, ALK-3 or ALK-6 under stringent hybridization conditions, and which have substantially the same OP-1 binding affinity as ALK-2, ALK-3 or ALK-6. As used herein, stringent hybridization conditions are as defined in the art, (see, for example, Molecular Cloning: A Laboratory Manual, Maniatis et al., eds. 2d.ed., Cold Spring Harbor Press, Cold Spring Harbor, 1989.) An exemplary set of conditions is defined as: hybridization in 40% formamide, 5 X SSPE, 5 X Denhardt's Solution, and 0.1% SDS at 37°C overnight, and washing in 0.1 X SSPE, 0.1% SDS at 50°C.

In still another embodiment, the OP-1 binding receptor analogs contemplated by the invention include part or all of a serine/threonine kinase receptor encoded by a nucleic acid that can be amplified with one or more primers derived from ALK-1 (Seq. ID No. 1), ALK-2, ALK-3 or ALK-6 sequence in a standard PCR (polymerase chain reaction) amplification scheme. In particular, a primer or, most preferably, a pair of primers represented by any of the sequences of SEQ ID Nos. 12-15 are envisioned to be particularly useful. Use of primer pairs (e.g., SEQ. ID No. 12/15; 13/15; 14/15) are described in WO94/11502 (PCT/GB93/02367).

Useful OP1-specific receptor analogs include xenogenic (foreign species) homologs of the murine and human ALK sequences described herein, including those obtained from other mammalian species, as well as other, eukaryotic, non-mammalian xenogenic homologs. Also contemplated are biosynthetic constructs and naturally-occurring sequence variants of ALK-2,

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ALK-3 and ALK-6, provided these molecules, in all cases, share the appropriate identity in the ligand binding domain, and bind OP-1 specifically as defined herein. In one embodiment, sequence variants include receptor analogs which have substantially the same binding affinity for OP1 as ALK-2, ALK-3 or ALK-6 and which are recognized by an antibody having binding specificity for ALK-2, ALK-3 or ALK-6.

In another embodiment the receptors and OP-1 binding receptor analogs contemplated herein provide the means by which a morphogen, e.g., OP-1, can mediate a cellular response. In one embodiment these receptors include ALK-2, ALK-3, or ALK-6, or sequence variants or OP-1 binding analogs thereof. In another embodiment, ALK-1, including sequence variants thereof is contemplated to participate in an OP-1 mediated cellular response.

OP1-specific receptor analogs may be used as OP1 antagonists. For example, a soluble form of a receptor, e.g., consisting essentially of only the extracellular ligand-binding domain, may be provided systemically to a mammal to bind to soluble ligand, effectively competing with ligand binding to a cell surface receptor, thereby modulating (reducing) the availability of free ligand in vivo for cell surface binding.

20 ligands in the invention include OP-1 and OP-1 receptor-binding analogs. As used herein, an "OP-1 analog" or "OP-1 receptor-binding analog" is understood to include all molecules able to functionally substitute for OP-1 in Type I receptor binding, e.g., are able to successfully compete with OP-1 for receptor binding in a standard competition assay. In one embodiment, useful OP-1 receptor-binding analogs include molecules whose binding affinity is defined by a dissociation constant of less than about 5x 10<sup>-6</sup>M, preferably less than about 10<sup>-7</sup>M or 5x 10<sup>-7</sup>M. As for the OP-specific receptor analogs above, both stronger and weaker binding affinities are contemplated to be useful in particular applications. In one preferred embodiment, these receptors.

The OP-1 analogs contemplated herein, all of which mimic the binding activity of OP-1 or an OP-1-related protein sufficiently to act as a substitute for OP-1 in receptor binding, can act as OP-1 agonists, capable of mimicking OP-1 both in receptor binding and in inducing a transmembrane effect e.g., inducing threonine or serine-specific phosphorylation following binding. Alternatively, the OP-1 analog can act as an OP-1 antagonist, capable of mimicking OP-1 in receptor binding only, but unable to induce a transmembrane effect, thereby blocking the natural ligand from interacting with its receptor, for example. Useful applications for antagonists include their use as therapeutics to modulate uncontrolled differentiated tissue growth, such as occurs in malignant transformations such as in osteosarcomas or Paget's disease.

OP-1 analogs contemplated by the invention can be amino acid-based, e.g., sequence variants of OP-1, or antibody-derived sequences capable of functionally mimicking OP-1 binding to an OP-1-specific receptor. Examples of such antibodies may include anti-idiotypic antibodies. In a specific embodiment, the anti-idiotypic antibody mimics OP1 both in receptor binding and in ability to induce a transmembrane effect. Alternatively, the OP-1 analogs can be composed in part or in whole of other chemical structures, e.g., the analogs can be comprised in part or in whole of nonproteinaceous molecules. In addition, the OP-1 analogs contemplated can be naturally sourced or synthetically produced.

As used herein, OP-1 related sequences include sequences sharing at least 60%, preferably greater than 65% or even 70% identity with the C-terminal 102 amino acid sequence of OP-1 as defined in Seq ID NO.7, and which are able to substitute for OP-1 in ligand binding to ALK-2, ALK-3 or ALK-6, (e.g., able to compete successfully with OP-1 for binding to one or more of these receptors.) OP-1 related sequences contemplated by the invention include xenogenic homologs (e.g., the Drosophila homolog 60A), and the related sequences referenced herein and in the literature as OP-2, OP-3, BMP5, BMP6 (and its xenogenic homolog Vgr-1.) OP-1 related sequences also include sequence variants encoded by a nucleic acid which hybridizes with a DNA

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sequence comprising the C-terminal 102 amino acids of Seq. ID No. 9 under stringent hybridization conditions and which can substitute for OP1 in an OP1-receptor binding assay. In another embodiment, an OP1 sequence variant includes a protein which can substitute for OP1 in a ligand-receptor binding assay and which is recognized by an antibody having binding specificity for OP1.

As used herein, "amino acid sequence homology" is understood to mean amino acid sequence similarity, and homologous sequences sharing identical or similar amino acids, where similar amino acids are conserved amino acids as defined by Dayoff et al., Atlas of Protein Sequence and Structure; vol.5, Suppl.3, pp.345-362 (M.O. Dayoff, ed., Nat'l BioMed. Research Fdn., Washington D.C. 1978.) Thus, a candidate sequence sharing 60% amino acid homology with a reference sequence requires that, following alignment of the candidate sequence with the reference sequence, 60% of the amino acids in the candidate sequence are identical to the corresponding amino acid in the reference sequence, or constitute a conserved amino acid change thereto. "Amino acid sequence identity" is understood to require identical amino acids between two aligned sequences. Thus, a candidate sequence sharing 60% amino acid identity with a reference sequence requires that, following alignment of the candidate sequence with the reference sequence, 60% of the amino acids in the candidate sequence are identical to the corresponding amino acid in the reference sequence.

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As used herein, all receptor homologies and identities calculated use ALK-6 as the reference sequence, with the extracellular domain reference sequence constituting residues 23-122 of Seq. ID No.7; and the intracellular serine/threonine kinase domain reference sequence constituting residues 206-495 of Seq. ID No.7. Similarly, all OP-1 related protein homologies and identities use OP-1 as the reference sequence, with the C-terminal 102 amino acids described in Seq. ID No. constituting the seven cysteine domain.

Also as used herein, sequences are aligned for homology and identity calculations as follows: Sequences are aligned by eye to maximize sequence identity. Where receptor amino acid extracellular domain sequences are

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compared, the alignment first maximizes alignment of the cysteines present in the two sequences, then modifies the alignment as necessary to maximize amino acid identity and similarity between the two sequences. Where amino acid intracellular domain sequences are compared, sequences are aligned to maximize alignment of conserved amino acids in the kinase domain, where conserved amino acids are those identified by boxes in Fig.3. The alignment then is modified as necessary to maximize amino acid identity and similarity. In all cases, internal gaps and amino acid insertions in the candidate sequence as aligned are ignored when making the homology/identity calculation. Exemplary alignments are illustrated in Figs. 2 and 3 where the amino acid sequences for the extracellular and intracellular domains, respectively are presented in single letter format. In the figures "gaps" created by sequence alignment are indicated by dashes.

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In one aspect, the invention contemplates isolated ligand-receptor complexes comprising OP-1 or an OP-1 analog as the ligand in specific binding interaction with an OP-1 binding Type I receptor or receptor analog, as defined herein. In another aspect, the invention contemplates the ligandreceptor complex comprises part or all of an OP-1 binding Type II receptor. Type II receptors contemplated to be useful include Type II receptors defined in the literature (referenced hereinabove) as having binding specificity for activin or a bone morphogenic protein such as BMP-4. Such Type II receptors include daf4, ActRII and AtrII. In still another aspect, the ligand-receptor complex comprises both a Type I and a Type II receptor and OP1, or an OP1 analog as the ligand. In all complexes, the bound receptor can comprise just the extracellular, ligand binding domain, or can also include part or all of the transmembrane sequence, and/or the intracellular kinase domain. Similarly, the OP-1 ligand may comprise just the receptor binding sequence, longer sequences, including the mature dimeric species or any soluble form of the protein or protein analog.

The OP-1 and OP-1 analogs described herein can interact specifically with Type I and Type II receptors also known to interact with other morphogenic proteins (e.g., EMP2/BMP4) and activin. Thus invention also

contemplates the use of OP-1 and OP-1 receptor-binding analogs as competitors of specific BMP-receptor and activin-receptor interactions. As will be appreciated by those having ordinary skill in the art, these binding competitors may act as either agonists or antagonists (e.g., to inhibit an activin or BMP-mediated cellular response).

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In another aspect, the invention contemplates binding partners having specific binding affinity for an epitope on the ligand-receptor complex. In a preferred embodiment, the binding partner can discriminate between the complex and the uncomplexed ligand or receptor. In another embodiment, the binding partner has little or no substantial binding affinity for the uncomplexed ligand or receptor. In another preferred embodiment, the binding partner is a binding protein, more preferably an antibody. These antibodies may be monoclonal or polyclonal, may be intact molecules or fragments thereof (e.g., Fab, Fab', (Fab)'2), or may be biosynthetic derivatives, including, but not limited to, for example, monoclonal fragments, such as single chain F, fragments, referred to in the literature as sFus, BABs and SCAs, and chimeric monoclonals, in which portions of the monoclonals are humanized (excluding those portions involved in antigen recognition (e.g., complementarity determining regions, "CDRs".) See, for example, U.S. Pat. Nos. 5,091,513 and 5,132,405, the disclosures of which are incorporated herein by reference. Biosynthetic chimeras, fragments and other antibody derivatives may be synthesized using standard recombinant DNA methodology and/or automated chemical nucleic acid synthesis methodology well described in the art and as described below.

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In still another aspect, the invention provides molecules useful in the design and/or identification of receptor-binding morphogenic protein analogs as described below, as well as kits and methods, e.g., screening assays, for identifying these analogs. The molecules useful in these assays can include part or all of the receptor sequence of SEQ ID NO. 3, 5 or 7, including amino acid sequence variants and OP-1 binding analogs and amino acid sequence variants thereof.

As described above, sequence variants are contemplated to have substantially the same binding affinity for OP-1 as the receptors represented by the sequences in SEQ. ID Nos. 3-7. OP-1 binding receptor analogs include other, known or novel Type I or Type II serine/threonine kinase receptors having binding affinity and specificity for OP-1 as defined herein and which (1) share at least 40% amino acid identity with residues 23-122 of Seq. ID No. 7, (2) are encoded by a nucleic acid that hybridizes under stringent conditions with a nucleic acid comprising the sequence defined by nucleotides 256-552 of Seq ID No. 7; or (3) are encoded by a nucleic acid obtainable by amplification with one or more primer sequences defined by Seq. ID Nos. 12-15. Currently preferred for the assays of the invention are receptor sequences comprising at least the sequence which defines the extracellular, ligand binding domains of these proteins. The kits and assays may include just Type I receptors or both Type I and Type II receptors. Similarly, the kits and screening assays can be used in the design and/or identification of OP1specific receptor analogs. The OP-1 receptor-binding analogs and OP-1-binding receptor analogs thus identified then can be produced in reasonable quantities using standard recombinant expression or chemical synthesis technology well known and characterized in the art. Alternatively, promising candidates can be modified using standard biological or chemical methodologies to, for example, enhance the binding affinity of the candidate analog as described in Example 10, below, and the preferred candidate derivative then produced in quantity.

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In still another aspect, the receptor and/or OP1-specific receptor analogs can be used in standard methodologies for affinity purifying and/or quantifying OP1 and OP1 analogs. For example, the receptor's ligand binding domain first may be immobilized on a surface of a well or a chromatographic column; ligand in a sample fluid then may be provided to the receptor under conditions to allow specific binding; non-specific binding molecules then removed, e.g., by washing, and the bound ligand then selectively isolated and/or quantitated. Similarly, OP1 and OP1 analogs can be used for affinity purifying and/or quantifying OP1-specific receptors and receptor analogs. In

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one embodiment, the method is useful in kits and assays for diagnostic purposes which detect the presence and/or concentration of OP1 protein or related morphogen in a body fluid sample including, without limitation, serum, peritoneal fluid, spinal fluid, and breast exudate. The kits and assays also can be used for detecting and/or quantitating OP-1-specific receptors in a sample.

In still another aspect the invention comprises OP1-specific receptors and OP-1-binding receptor analogs useful in screening assays to identify organs, tissues and cell lines which express OP1-specific receptors. These cells then can be used in screening assays to identify ligands that modulate endogenous morphogen receptor expression levels, including the density of receptors expressed on a cell surface. Useful assay methodologies may be modeled on those described in PCT US92/07359, and as described below.

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The invention thus relates to compositions and methods for the use of morphogen-specific receptor sequences in diagnostic, therapeutic and experimental procedures. Active receptors useful in the compositions and methods of this invention can include truncated or full length forms, as well as forms having varying glycosylation patterns. Active receptors useful in the invention also include chimeric constructs as described below. Active OP1-specific receptors/analogs can be expressed from intact or truncated genomic or cDNA, or from synthetic DNAs in procaryotic or eucaryotic host cells, and purified, cleaved, refolded and oxidized as necessary to form active molecules. Useful host cells include prokaryotes, including E. coli and B. subtilis, and eukaryotic cells, including mammalian cells, such as fibroblast 3T3 cells, CHO, COS, melanoma or BSC cells, Hela and other human cells, the insect/baculovirus system, as well as yeast and other microbial host cell systems.

Thus, in view of this disclosure, skilled genetic engineers now can, for example, identify and produce OP1-specific cell surface receptors or analogs thereof; create and perform assays for screening candidate OP1 receptor-binding analogs and evaluate promising candidates and their progency in therapeutic regimes and preclinical studies; modulate the availability of

endogenous morphogen for cell surface interactions; modulate endogenous morphogen-specific cell surface receptor levels; elucidate the signal transduction pathway induced by morphogen-cell surface receptor binding; and modulate tissue morphogenesis in vivo.

## 5 Brief Description of the Drawings

Figure 1 is a schematic representation of the encoded ALK-2, ALK-3, ALK-6 amino acid sequences, showing the signal sequence 10, the transmembrane domain 12, the extracellular ligand binding domain 14, and the intracellular serine/threonine kinase domain 16;

Figure 2 is a homology alignment of the extracellular domains of ALK-2, ALK-3, and ALK-6, aligned to maximize amino acid identity, wherein conserved amino acids are identified by boxes and conserved cysteines are indicated by asterisks: and

Figure 3 is a homology alignment of the intracellular domain of ALK-2, ALK-3 and ALK-6, aligned to maximize amino acid identity, wherein conserved amino acids are boxed and the serine/threonine kinase domain is indicated by arrows.

#### Detailed Description

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Disclosed herein are Type I and Type II receptors having binding specificity for true tissue morphogenic proteins, particularly OP1 and OP1-related proteins. It further has been determined that OP1 binds to a broader range of receptors than other known tissue morphogens or TGF-ß family members. The Type I receptors disclosed herein, can be used together with OP1 and OP1 analogs for therapeutic, diagnostic and experimental uses as described herein below. Moreover, soluble forms of the OP1-binding receptor proteins, e.g., forms consisting essentially of the extracellular domain or a fragment thereof sufficient to bind OP1 with specificity, can be used as a soluble therapeutic morphogen antagonist, as described below.

Following this disclosure, related OP1-specific receptors are available, as are high and medium flux screening assays for identifying OP1 analogs and OP1-specific receptor analogs. These analogs can be naturally

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occurring molecules, or they can be designed and biosynthetically created using a rational drug design and an established structure/function analysis. The analogs can be amino acid-based or can be composed in part or whole of non-proteinaceous synthetic organic molecules. Useful analogs also can include antibodies, preferably monoclonal antibodies (including fragments thereof, e.g., Fab, Fab', and (Fab)'2), or synthetic derivatives thereof, such as monoclonal single chain  $F_{\mathbf{v}}$  fragments known in the art as  $sF_{\mathbf{v}}s$ , BABs, and SCAs (see below), and bispecific antibodies or derivatives thereof. these antibodies mimic the binding activity of OP-1 to a cell surface receptor without inducing the biological response OP-1 does upon binding, the antibody can compete for OP-1 binding and act as an antagonist. These antibodies or derivatives thereof also can mimic OP-1 both in receptor binding and signal transduction, in which case the antibody acts as an OP-1 agonist. The antibodies and derivatives also can be used for inducing the morphogenic cellular response by crosslinking receptors to morphogenic proteins, particularly OP1 and OP1-related proteins to form either homo- or heterocomplexes of the Type I and Type II receptors.

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The OP1-binding receptor sequences described herein (ALK2, ALK3, ALK6) also can be used to create chimeric sequences, wherein, for example, part or all of either the extracellular domain or the intracellular domain is a non-ALK sequence or is created from two or more ALK sequences. These chimeric receptors can be synthesized using standard recombinant DNA methodology and/or automated chemical nucleic acid synthesis methodology well described in the art and as disclosed below. Chimerics can be used, for example, in OP1 analog assays, wherein the OP1-binding extracellular domain is coupled to a non-ALK intracellular domain that is well characterized and/or readily detectable as a second messenger response system, as described below. Chimerics also can be used, for example, in high flux OP1 analogs screens and as part of purification protocols, wherein a soluble ligand binding domain of an OP1-specific receptor is immobilized onto a support e.g., by covalent or non-covalent interactions, with a chromatographic matrix or the well surface of a 96-well plate. When immobilized onto a chromatographic matrix surface,

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the receptor fragment can be used in a protocol to isolate OP1 or OP1 analogs. When immobilized on a well surface the receptor fragment is particularly useful in a screening assay to identify receptor-binding OP1 analogs in a standard competition assay.

The true tissue morphogenic proteins contemplated to be useful in the methods and compositions of the invention include forms having varying glycosylation patterns and varying N-termini. The proteins can be naturally occurring or biosynthetically derived, and can be produced by expression of recombinant DNA in prokaryotic or eukaryotic host cells. The proteins are active as a single species (e.g., as homodimers), or combined as a mixed species. Useful sequences and eucaryotic and procaryotic expression systems are well described in the art. See, for example, US Patent Nos. 5,061,911 and 5,266,683 for useful expression systems.

Particularly contemplated herein are OP1 and OP1-related sequences. Useful OP1 sequences are recited in US Pat Nos. 5,011,691; 5,018,753 and 5,266,683; in Ozkaynak et al. (1990) EMBO J 9:2085-2093; and Sampath et al. (1993) PNAS 90: 6004-6008. OP-1 related sequences include xenogenic homologs, e.g.; 60A, from Drosophila, Wharton et al. (1991) PNAS 88:9214-9218; and proteins sharing greater than 60% identity with OP1 in the C-terminal seven cysteine domain, preferably at least 65% identity. Examples of OF-1 related sequences include BMP5, BMP6 (and its species homolog Vgr-1, Lyons et al. (1989) PNAS 86:4554-4558), Celeste, et al. (1990) PNAS 87:9843-9847 and PCT international application WO93/00432; OP-2 (Ozkaynak et al. (1992) J.Biol. Chem. 267:13198-13205) and OP-3 (PCT international application WO94/06447). 25 As will be appreciated by those having ordinary skill in the art, chimeric constructs readily can be created using standard molecular biology and mutagenesis techniques combining various portions of different morphogenic protein sequences to create a novel sequence, and these forms of the protein also are contemplated herein.

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A particularly preferred embodiment of the proteins contemplated by the invention includes proteins whose amino acid sequence in the cysteine-rich Cterminal domain has greater than 60% identity, and preferably greater than 65%

identity with the amino acid sequence of OPS (OP-1 sequence defining the C-terminal conserved six cysteines, e.g., residues 335-431 of Seq. ID No. 9).

In another preferred aspect, the invention contemplates osteogenic proteins comprising species of polypeptide chains having the generic amino acid sequence herein referred to as "OPX" which accommodates the homologies between the various identified species of the osteogenic OP1 and OP2 proteins, and which is described by the amino acid sequence presented below and in Sequence ID No. 11.

Cys Xaa Xaa His Glu Leu Tyr Val Ser Phe 10 Xaa Asp Leu Gly Trp Xaa Asp Trp Xaa Ile Ala Pro Xaa Gly Tyr Xaa Ala Tyr Tyr Cys 15 Glu Gly Glu Cys Xaa Phe Pro Leu Xaa Ser Xaa Met Asn Ala Thr Asn His Ala Ile Xaa Gln Xaa Leu Val His Xaa Xaa Xaa Pro Xaa 20 Xaa Val Pro Lys Xaa Cys Cys Ala Pro Thr Xaa Leu Xaa Ala Xaa Ser Val Leu Tyr Xaa 75 25 Asp Xaa Ser Xaa Asn Val Ile Leu Xaa Lys Xaa Arg Asn Met Val Val Xaa Ala Cys Gly Cys His, 30

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and wherein Xaa at res. 2 = (Lys or Arg); Xaa at res. 3 = (Lys or Arg); Xaa at res. 11 = (Arg or Gln); Xaa at res. 16 = (Gln or Leu); Xaa at res. 19 = (Ile or Val); Xaa at res. 23 = (Glu or Gln); Xaa at res. 26 = (Ala or Ser); Xaa at res. 35 = (Ala or Ser); Xaa at res. 39 = (Asn or Asp); Xaa at res. 41 = (Tyr or Cys); Xaa at res. 50 = (Val or Leu); Xaa at res. 52 = (Ser or Thr); Xaa at res. 56 = (Phe or Leu); Xaa at res. 57 = (Ile or Met); Xaa at res. 58 = (Asn or Lys); Xaa at res. 60 = (Glu, Asp or Asn); Xaa at res. 61 = (Thr, Ala or Val); Xaa at res. 65 = (Pro or Ala); Xaa at res. 71 = (Gln or Lys); Xaa at res. 73 = (Asn or Ser); Xaa at res. 75 = (Ile or Thr); Xaa at res. 80 = (Phe or Tyr); Xaa at res. 82 = (Asp or Ser); Xaa at res. 84 = (Ser or Asn); Xaa at res. 89 = (Lys or Arg); Xaa at res. 91 = (Tyr or His); and Xaa at res. 97 = (Arg or Lys).

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In still another preferred aspect, the invention contemplates osteogenic proteins encoded by nucleic acids which hybridize to DNA or RNA sequences encoding the C-terminal seven cysteine domain of OP1 or OP2 under stringent hybridization conditions.

- A brief description of the various terms of OP-1 useful in the invention is described below:
  - Refers generically to the family of osteogenically active OP1 proteins produced by expression of part or all of the hOP1 gene. Also referred to in related applications as "OPI" and "OP-1".
- OP1-PP -Amino acid sequence of human OP1 protein (prepro form), Seq. ID No. 9, residues 1-431. Also referred to in related applications as "OP1-PP" and "OPP".
- OP1-18Ser Amino acid sequence of mature human OP1 protein, Seq. ID No. 9, residues 293-431. N-terminal amino acid is serine. Originally identified as 15 migrating at 18 kDa on SDS-PAGE in COS cells. Depending on protein glycosylation pattern in different host cells, also migrates at 23kDa, 19kDa and 17kDa on SDS-PAGE. Also referred to in related applications as "OP1-18."
  - OP1-16Ser; OP1-16Ala; OP1-16 Met; OP1-16 leu; OP1-16Val N-terminally truncated mature human OP1 protein species defined, respectively, by residues 300-431; 316-431; 315-431; 313-431 and 318-431.
  - OPS Amino acid sequence defining the C-terminal six cysteine domain, residues 335-431 of Seq. ID No. 9.

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- OP7 Amino acid sequence defining the C-terminal seven cysteine domain, residues 330-431 of Seg. ID No. 9.
- 25 Soluble form OP1 mature dimeric OP1 species having one or, preferably two copies of pro domain, e.g., at least residues 158-292 of Seq. ID No. 9, preferably residues 48-292 or 30-292, non-covalently complexed with the dimer.

The cloning procedure for obtaining OP1-binding ALK nucleic acid sequences, means for expressing receptor sequences, as well as other material aspects concerning the nature and utility of these sequences, including how to make and how to use the subject matter claimed, will be further understood from the following, which constitutes the best mode currently contemplated for practicing the invention.

#### Example 1. IDENTIFICATION OF ALK-1, ALK-2, ALK-3 and ALK-6

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The cloning and characterization of ALK-1, -2, -3, and -6 receptors are described in detail in ten Dijke et al. (1993) Oncogene 8:2879-2887; and (1994) Science 264:101-104. The general structures of these proteins is described in Fig. 1, and the sequence alignments between the ALK genes are shown in Figs 2 and 3. These molecules have similar domain structures: an N-terminal predicted hydrophobic signal sequence (von Heijne (1986) Nucl. Acids Res. 14: 4683-4690) is followed by a relatively small extracellular cysteinerich ligand binding domain, a single hydrophobic transmembrane region (Kyte & Doolittle (1982) J. Mol. Biol. 157, 105-132) and a C-terminal intracellular portion, which consists almost entirely of a kinase domain (Figure 3).

The extracellular domains of these receptors, defined essentially by residues 22-118 (SEQ. ID No. 1 ) for ALK-1; residues 16-123 (SEQ ID No. 3) for ALK-2; residues 24-152 (SEQ. ID No. 5) for ALK-3; and residues 23-122 (SEQ ID No. 7) for ALK-6, have cysteine-rich regions, but sequence similarity varies among the proteins. For example, ALK-3 and ALK-6 share a high degree of sequence similarity in their extracellular domains (46% identity) whereas ALK-2 shows less similarity with ALK 3 or ALK6 (see Fig. 2.)

The positions of many of the cysteine residues in these receptors can be aligned, indicating that the extracellular domains likely adopt a similar structural configuration.

The intracellular domains of these receptors are characterized by a serine/threonine kinase, defined essentially by residues 204-494 (SEQ. ID. No. 1) for ALK-1; residues 210-510 (SEQ ID No. 3) for ALK-2; residues 236-527 (SEQ ID No. 5) for ALK-3; and residues 206-497 (SEQ ID No. 7) for ALK-6. The catalytic domains of kinases can be divided into 12 subdomains with stretches

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of conserved amino-acid residues. The key motifs are found in serine/threonine kinase receptors indicating that they are functional kinases. The consensus sequence for the binding of ATP (Gly-X-Gly-X-X-Gly in subdomain I followed by a Lys residue further downstream in subdomain II) is found in all the ALKS. Moreover, ALK-1, ALK-2, ALK-3 and ALK-6 have the sequence motifs or similar motifs HRDLKSKN (Subdomain VIB) and GTKRYMAPE (Subdomain VIII), that are found in most of the serine/threonine kinase receptors and can be used to distinguish them from tyrosine kinase receptors. Two short inserts in the kinase domain (between subdomain VIA and VIB and between X and XI are unique to members of this serine/threonine kinase receptor family. In the intracellular domain, these regions, together with the juxtamembrane part and C-terminal tail, are the most divergent between family members.

Type II serine/threonine kinase receptors known in the art are described and referenced herein above. <a href="mailto:Example 2">Example 2</a>. RECEPTOR EXPRESSION A. General Considerations

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Receptor DNA, or a synthetic form thereof, can be inserted, using conventional techniques well described in the art (see, for example, Maniatis (1989) Molecular Cloning A Laboratory Manual), into any of a variety of expression vectors and transfected into an appropriate host cell to produce recombinant protein polypeptide chains, including both full length and truncated forms thereof. Shortened sequences, for example, can be used for the production of soluble receptor fragments.

Useful host cells include <u>E</u>. <u>coli</u>, <u>Saccharomyces cerevisiae</u>, <u>Pichia pastoris</u>, the insect/baculovirus cell system, myeloma cells, and various other mammalian cells. The full length forms of the proteins of this invention preferably are expressed in mammalian cells, as disclosed herein. Soluble forms may be expressed from both mammalian or bacterial cell systems. The vector additionally may include various sequences to promote correct expression of the recombinant protein, including transcription promoter and termination sequences, enhancer sequences, preferred ribosome binding site sequences, preferred mRNA leader sequences, preferred protein processing sequences, preferred signal sequences for protein secretion, and the like.

The DNA sequence encoding the gene of interest also may be manipulated to remove potentially inhibiting sequences or to minimize unwanted secondary structure formation. The recombinant morphogen receptor also may be expressed as a fusion protein. After translation, the protein may be purified from the cells themselves or recovered from the culture medium. The DNA also may include sequences which aid in expression and/or purification of the recombinant protein. One useful sequence for example, is a hexa-His (His<sub>6</sub>) sequence, which adds a histidine tail to allow affinity purification of the protein on an IMAC Cu2+ column (see below.)

For example, the DNA encoding the extracellular domain may be inserted into a suitable expression vector for transformation into a prokaryote host such as <u>E. coli</u> or <u>B. subtilis</u>, to produce a soluble, morphogen binding fragment. The DNA may expressed directly or may be expressed as part of a fusion protein having a readily cleavable fusion junction. An exemplary protocol for prokaryote expression using MR-1 DNA is provided below. Recombinant protein is expressed in inclusion bodies and may be purified therefrom using the technology disclosed in U.S. Patent No. 5,013,653, for example.

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The DNA also may be expressed in a suitable mammalian host. Useful hosts include fibroblast 3T3 cells, (e.g., NIH 3T3, from CRL 1658) COS (simian kidney ATCC, CRL-1650) or CH0 (Chinese hamster ovary) cells (e.g., CHO-DXB11, from Lawrence Chasin, <a href="Proc. Nat'l. Acad. Sci.">Proc. Nat'l. Acad. Sci.</a> (1980) 77(7):4216-4222), minklung epithelial cells (MV1Lu), human foreskin fibroblast cells, human glioblastoma cells, and teratocarcinoma cells. Other useful eukaryotic cell systems include yeast cells, the insect/baculovirus system or myeloma cells.

To express an OP1-specific cell surface receptor, the DNA is subcloned into an insertion site of a suitable, commercially available vector along with suitable promoter/enhancer sequences and 3' termination sequences. Useful promoter/enhancer sequence combinations include the CMV promoter (human cytomegalovirus (MIE) promoter) present, for example, on pCDM8, as well as the mammary tumor virus promoter (MMTV) boosted by the Rous sarcoma virus LTR enhancer sequence (e.g., from Clontech, Inc., Palo Alto). A useful induceable

promoter includes, for example, A Zn<sup>2\*</sup> induceable promoter, such as the Zn<sup>2\*</sup> metallothionein promoter (Wrana et al. (1992) <u>Cell 71:1003-1014.</u>) <u>Other induceable promoters are well known in the art and can be used with similar success. Expression also may be further enhanced using transactivating enhancer sequences. The plasmid also preferably contains an amplifiable marker, such as DHFR under suitable promoter control, e.g., SV40 early promoter (ATCC #37148). Transfection, cell culturing, gene amplification and protein expression conditions are standard conditions, well known in the art, such as are described, for example in Ausubel et al., ed., <u>Current Protocols in Molecular Biology</u>, John Wiley & Sons, NY (1989). Briefly, transfected cells are cultured in medium containing 5-10% dialyzed fetal calf serum (FCS), and stably transfected high expression cell lines obtained by amplification and subcloning and evaluated by standard Western and Northern blot analysis. Southern blots also can be used to assess the state of integrated receptor sequences and the extent of their copy number amplification.</u>

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The expressed protein then is purified using standard procedures. A currently preferred methodology uses an affinity column, such as a ligand affinity column or an antibody affinity column, the bound material then washed, and receptor molecules selectively eluted in a gradient of increasing ionic strength, changes in pH or addition of mild denaturants. Alternatively, where a useful anchor sequence has been added to the DNA, such as a (His)<sub>6</sub> sequence, the column may be a standard affinity column such as Cu<sup>2+</sup> IMAC column. Here, for example, the cell culture media containing the recombinant protein is passed over a Cu<sup>2+</sup> IMAC column (for example, prepared with 25 mM imidazole). The bound protein then is washed with a compatible solution and eluted with EDTA. The anchor sequence can be removed by a standard chemical or enzymatic procedure.

Mammalian cell expression is preferred where morphogen receptor expression on a cell surface is desired. For example, cell surface expression may be desired to test morphogen or morphogen analog binding specificity for a cell surface receptor under <u>in vivo</u> conditions. Cell surface expression also

may be most efficacious for medium flux cellular screen assays as described below.

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## B.1 Exemplary Mammalian Cell Culture

The receptors tested in Examples 8 and 9 described below were expressed in (1) COS-1 cells; (2) mink lung epithelial cells (Mv1Lu); (3) AG1518 human foreskin fibroblasts; (4) MG-63 human osteosarcoma cells; (5) PC12 rat pheochromocytoma cells (all obtained from American Type Culture Collection, Rockville, MD); (6) U-1240 MG human glioblastoma cells (Bengt Westermark, et al. (1988) Cancer Research 48:3910-3918); (7) Tera-2 teratocarcinoma cells (clone 13, Thompson et al. (1984) J Cell Sci 72:37-64); (8) MC3T3-El cells · 10 (Sudo et al. (1983) J. Cell Biol. 96:191-198, and (9) ROS 17/2.8 rat osteosarcoma cells (Majeska et al. (1985) Endocrinology 116:170-179. The ROS cells were cultured in Ham's F12 medium containing 14mM HEPES buffer, 2.5 mM L-glutamine, 1.1 mM CaCl<sub>2</sub>, 5% fetal bovine serum and antibiotics; MC3T3-E1 cells were cultured in a-MEM with 10% fetal bovine serum and antibiotics, and Tera-2 cells were cultured in 5% CO2 atmosphere at 37oC in a-MEM containing 10% fetal bovine serum, 100 units/ml of penicillin and 50 mg/ml of streptomycin, using tissue culture dishes pretreated with 0.1% swine skin gelatin (Sigma) in phosphate-buffered saline. Unless otherwise specified, cells were cultured in DMEM containing 10% fetal bovine serum and antibiotics. B.2 Transfection of cDNA

The receptors tested in Example 8.1 (ALK 1-6, daf-4) were transfected as follows. Transient expression plasmids of ALK-1 to -6 and daf-4 were generated by subcloning into an expression vector (pSV7d, Truett et al. (1985) DNA 4:333-349) or into the pcDNA I expression vector (Invitrogen, San Diego). For transient transfection, COS-1 cells were transfected with 10 mg each of plasmids by a calcium phosphate precipitation method using a mammalian transfection kit (Stratagene, La Jolla), following the manufacturer's protocol. One day after transfection, the cells were used for the affinity labeling and cross-linking experiments.

Example 3. ANTIBODY PRODUCTION

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#### A. General Considerations

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Antibodies capable of specifically binding the receptor molecules, ligand molecules, or the ligand-receptor complex itself, useful as analogs and useful in immunoassays and in the immunopurification of morphogen receptors described may be obtained as described below.

Where antibodies specific to the OPI-specific receptors are desired, but which do not interfere with ligand binding, the antigenic sequence preferably comprises the juxtamembrane sequence. Where antibodies capable of competing for ligand binding are desired, the ligand binding domain may be used as the antigen source. Where antibodies to the complex are desired, the complex itself preferably is used as the antigenic sequence and candidate antibodies then tested for cross reactivity with uncomplexed ligand and receptors versus the ligand-receptor complex. Finally, bispecific antibodies may used to complex ligand to a cell surface receptor (Type I or Type II) 15 and/or to target an agent or ligand to cells or tissue expressing a Type I or Type II morphogen-specific receptor. Preferred bispecific antibody derived molecules are single chain binding sites described in US Pat No. 5,091,513 and 5,132,405, the disclosures of which are incorporated hereinabove by reference.

Antibodies useful as OP1 receptor-binding analogs may be obtained using the receptor ligand binding domain as the immunogen source and testing receptor-binding analogs for their ability to compete with OP1 in a competition binding assay. Similarly, where antibodies useful as OP1-specific receptor analogs are desired, OP1 is the immunogen source and the antibody candidate tested in a competition assay with receptor protein.

Polyclonal antibodies specific for a morphogen receptor of interest may be prepared generally as described below. Each rabbit is given a primary immunization (e.g., 500mg) of antigen in 0.1% SDS mixed with 500 ml Complete Freund's Adjuvant. The antigen is injected intradermally at multiple sites on the back and flanks of the animal. The rabbit is boosted after a month with 500mg of antigen in the same manner using incomplete Freund's Adjuvant. Test bleeds are taken from the ear vein seven days later. Two additional boosts and test bleeds are performed at monthly intervals until antibody against the

antigenic sequence is detected in the serum using a standard Western blot.

Then, the rabbit is boosted monthly with 100 mg/ml of antigen and bled (15 ml per bleed) at days seven and ten after boosting.

Similarly, monoclonal antibodies specific for a given morphogen receptor molecule of interest may be prepared as described below: A mouse is given two injections of the antigenic sequence. The protein preferably is recombinantly produced. Where it is desired that the antibody recognize an epitope on the morphogen binding surface of a receptor an antigenic fragment derived from the extracellular domain preferably is provided. The first injection contains 100mg of antigen in complete Freund's adjuvant and is given subcutaneously. The second injection contains 50 mg of antigen in incomplete adjuvant and is given intraperitoneally. The mouse then receives a total of 230 mg of antigen in four intraperitoneal injections at various times over an eight month period. One week prior to fusion, the mouse is boosted intraperitoneally with antigen (e.g., 100 mg) and may be additionally boosted with an antigen-specific peptide conjugated to bovine serum albumin with a suitable crosslinking agent. This boost can be repeated five days (IP), four days (IP), three days (IP) and one day (IV) prior to fusion. The mouse spleen cells then are fused to commercially available myeloma cells at a ratio of 1:1 using PEG 1500 (Boehringer Mannheim, Germany), and the fused cells plated and screened for ALK-specific antibodies, e.g., using ALK-2, ALK-3 or ALK-6 as antigen. The cell fusion and monoclonal screening steps readily are performed according to standard procedures well described in standard texts widely available in the art. (See, for example, <u>Guide to Protein Purification</u> Murray P. Deutscher, ed., Academic Press, San Diego, 1990.

#### B. Exemplary ALK-Specific Antisera

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Antibodies used in the assay of Example 8 were obtained as follows.

Rabbit antisera against ALK-1 to -6 were made against synthetic peptides
corresponding to the divergent, intracellular juxtamembrane parts. (ALK-1:
residues 119-141; ALK-2: residues 151-172; ALK-3 residues 181-202; ALK-6:
residues 151-168.) Peptides were synthesized with an Applied Biosystems 430 A
Peptide Synthesizer using t-butoxycarbonyl chemistry, and purified by reverse

phase HPLC. The synthetic peptides were coupled to keyhole limpet hemocyanin (Calbiochem-Behring) using glutaraldehyde, as decribed by Gullick et al. (1985) EMBO J 4: 2869-2877. The coupled peptides then were mixed with Freund's adjuvant and used to immunize rabbits using standard methodologies.

## 5 Example 4. OP1-RECEPTOR BINDING ASSAYS

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Ligand binding specificity is determined by evaluating the ability of a receptor molecule to bind a specific ligand, and the ability of that ligand to compete against itself and other molecules which bind the receptor. Useful ligands will have a binding affinity for a soluble morphogen receptor extracellular domain such that dissociation constant (Kd) is less than about  $10^{-6}$ M, preferably less than  $5 \times 10^{-7}$ M. Where stronger binding interaction is desired, preferred affinities are defined by a Kd of  $10^{-8}$ - $10^{-9}$ M. OP1-related proteins are expected to be able to bind with specificity to multiple different receptor molecules, although likely with differing affinities.

Ligand binding specificity can be assayed as follows, essentially following standard protocols well described in the art and disclosed, for example, in Legerski et al. (1992) <u>Biochem. Biophys. Res. Comm. 183</u>:672-679 and Frakar et al., (1978) <u>Biochem. Biophys. Res. Comm. 80</u>:849-857. In the ligand binding assays, a ligand having a known, quantifiable affinity for a morphogen receptor molecule of interest is labelled, typically by radioiodination (125I), e.g., by chromogenic or fluorogenic labeling, or by metabolic labelling, e.g., 35s, and aliquots of cells expressing the receptor on their surface are incubated with the labelled ligand, in the presence of various concentrations of unlabelled potential competitor ligand. In the assays described in Examples 8 and 9, below, this competitor typically is the candidate morphogen analog or an aliquot from a broth or extract anticipated to contain a candidate morphogen analog.

Alternatively, a crosslinking agent may be used to covalently link the ligand to the bound receptor, and the crosslinked complex then immunoprecipitated with an antibody specific to the ligand, receptor, or complex. (See, Example 8.)

A standard, exemplary protocol for determining binding affinity is provided below. Briefly, cells expressing a receptor on their cell surface are plated into 35 mM dishes and incubated for 48 hours in DMEM (Dulbecco's modified Eagle medium) plus 10% fetal calf serum. Purified morphogen, here, e.g., OP-1, or an OP1-analog is iodinated with  $Na^{125}I$  by chloramine T oxidation, preferably having a specific activity of about 50-100 mCi/mg, essentially following the protocol of Frolik et al. (1984) J. Biol. Chem. 595:10995-11000. Labelled morphogen then is purified using standard procedures, e.g., chromatographically. Plated cells then are washed twice with physiologically buffered saline in the presence of 0.1% BSA, and incubated at 22oC in the presence of BSA, buffer and labelled morphogen (1 ng) and various concentrations (e.g., 0-10mg/ml) of unlabelled competitor, e.g., unlabelled morphogen or candidate ligand analogs. Following binding, cells are washed three times with cold buffer, solubilized in 0.5 ml of 0.5 N NaOH, removed from the dish, and radioactivity determined by gamma or scintillation counter. Data then are expressed as percent inhibition, where 100% inhibition of specific binding is the difference between binding in the absence of competitor and binding in the presence of a 100-fold molar excess of unlabelled morphogen. Binding parameters preferably are determined using a computer program such as LIGAND (Munsum et al. (1980) Anal. Biochem. 107:220-259.)

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Where the receptor cell surface binding domain is to be provided as a soluble protein, the assay can be performed in solution, most readily as an immunoprecipitation assay. In currently preferred assays the morphogen molecule is labelled and incubated with unlabelled receptor and candidate morphogen analogs. Receptor-specific antibody then is provided to the solution to precipitate the receptor-morphogen complex and the amount of labelled morphogen in the precipitated complex determined using standard detection means.

Where the receptor or ligand is to be used in an affinity isolation protocol, the molecule preferably is immobilized on a surface, preferably a matrix surface over which sample fluid will flow, allowing the ligand of

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interest to bind, at letting nonbinding components pass through as effluent. The complex then can be removed intact or the ligand selectively removed with a desired eluant.

#### 4.1 Screening Assay Considerations

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In the analog screening assays described in Example 9 below, the preferred protocol for assaying ligand-receptor binding is a standard competition or radioimmunoassay (RIA). Here the OP1 is labelled and the relative binding affinity of a candidate OP1 analog ligand in a sample is measured by quantitating the ability of the candidate (unlabelled ligand analog) to inhibit binding of the labelled ligand (competitor morphogen) by the receptor. In performing the assay, fixed concentrations of receptor and labelled morphogen are incubated in the absence and presence of unknown samples containing candidate ligands. Sensitivity can be increased by preincubating the receptor with candidate ligand before adding the labelled morphogen. After the labelled competitor has been added, sufficient time is allowed for adequate competitor binding, and then free and bound labelled morphogen are separated, and one or the other is measured. Useful morphogen labels include radioactive labels, chromogenic or fluorogenic labels, and conjugated enzymes having high turnover numbers, such as horseradish peroxidase, alkaline phosphatase, or b-galactosidase, used in combination with chemiluminescent or fluorogenic substrates. In the same manner, OP1-specific receptor analogs can be assayed for their affinity for OP1 in competition assays with labelled OP1 specific receptors.

Assays for evaluating a candidate OP1 receptor-binding analog's ability to mimic OP-1 in signal-transduction across a membrane are exemplified in detail in Example 9.2, below. Briefly, the assay involves use of a cell (1) known to express an OP-1-specific receptor; or (2) which can be modified to express an OP-1-specific receptor, and/or (3) which can induce an OP-1-mediated cellular response. In the assay, the ability of a candidate analog to induce an OP-1-specific cellular response is monitored. Numerous OP-1 responsive cells and OP-1-mediated inducible cellular and biochemical markers are known and described in the art. Alternatively, and as exemplified below,

an OP-1 inducible reporter gene system can be constructed and used to advantage in the assay.

#### 4.2 Diagnostic Assay Considerations

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The ability to detect morphogens in solution provides a valuable tool for diagnostic assays, allowing one to monitor the level of morphogen free in the body, e.g., in serum and other body fluids. For example, OP-1 has been detected in a number of different body fluids, including serum and spinal fluid, including cerebro-spinal fluid, saliva, milk and other breast exudates. (See, for example, PCT US92/07432, PCT US93/07231, W094/06449).

As one example, OP-1 is an intimate participant in normal bone growth and resorption. Thus, soluble OP-1 is expected to be detected at higher concentrations in individuals experiencing high bone formation, such as children, and at substantially lower levels in individuals with abnormally low rates of bone formation, such as patients with osteoporosis, aplastic bone disease, or osteopenia. Monitoring the level of OP-1 in serum thus provides a means for evaluating the status of bone tissue and bone homeostasis in an individual, as well as a means for monitoring the efficacy of a treatment to regenerate damaged or lost bone tissue. Alternatively, the level of OP-1 in bone tissue can be assessed in a bone tissue biopsy.

Similarly, OP-1 and other morphogens have been identified in brain tissue. In particular, OP-1 is expressed and/or localized in developing and adult rat brain and spinal cord tissue, in the hippocampus, substantia nigra and the adendema glial cells, as well as associated with astrocytes and the extracellular matrix surrounding neuronal cell bodies. (See, PCTUS93/07331). Thus, monitoring the level of OP-1 in spinal fluid or associated with a nerve tissue biopsy can provide means for evaluating the status of nerve tissue in an individual, as well as means for monitoring the efficacy of a nerve regeration or repair therapy.

For serum assays, the serum preferably first is partially purified to remove some of the excess, contaminating serum proteins, such as serum albumin. Preferably the serum is extracted by precipitation in ammonium sulfate (e.g., 45%) such that the complex is precipitated. Further

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purification can be achieved using purification strategies that take advantage of the differential solubility of soluble morphogen complex or mature morphogens relative to that of the other proteins present in serum. Further purification also can be achieved by chromatographic techniques well known in the art. The sample fluid then can be assayed for OP1 using the OP1-specific receptor(s) and binding assays as described herein.

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For a tissue biopsy, cells can be collected and stained with a labelled OP-1-specific antibody or receptor molecule. Alternatively, the OP-1 protein selectively can be extracted and quantitated as described above.

Morphogens useful in the binding/screening assays contemplated herein include the soluble forms of the protein, e.g., the mature dimeric species complexed with one or two copies of the pro domain, the mature dimeric species alone, and truncated forms comprising essentially just the C-terminal active domain.

#### 15 Example 5. TRANSMEMBRANE SIGNAL INDUCTION ASSAYS/OP1 MIMETICS

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The kinase activity of the intracellular domains of the OP1-specific receptors can be tested in an autophophorylation assay as described by Mathews et al. (PCT/US92/03825, published November 26, 1992) . Briefly, the DNA fragment encoding at least the intracellular kinase domain of an OP1-specific receptor is subcloned into pGEX-2T (Smith et al.(1988) Gene 67:31-40) to create a fusion protein between the putative kinase domain and glutathione Stransferase (GST). The plasmid is introduced into E, coli and the expressed fusion protein purified using glutathione affinity chromatography. About 100-200 ng of fusion protein or purified GST then are incubated with 25 mCi (gp32P) ATP in 50mM tris, 10mM MgCl2 buffer for 30 minutes at 37° C. Products then are analyzed by gel electrophoresis and autoradiography. The fusion protein, but not GST alone, becomes phophorylated, indicating that the kinase domain is functional. Phosphoamino acid analysis then can be performed to determine the predominant amino acid being phosphorylated. Similar assays can be performed using similar fusion constructs expressed in mammalian cells.

Various signaling transduction assays are provided in Example 9, below. An assay also can be developed for testing kinase activity transduction upon

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ligand binding using a ligand-induced kinase activity assay known in the art. Here, the ability of OP-1 analog to induce phosphorylation upon binding to the receptor is tested.

See, for example, various assays for measuring ligand-induced kinase activity described by Accili et al. (1991) J. Biol. Chem. 266:434-439 and Nakamura et al. (1992) J.Biol. Chem. 267: 18924-18928,. For example, ligandinduced kinase activity (e.g., receptor autophosphorylation) can be measured in vitro by incubating purified receptor in the presence and absence of ligand (here, OP1 or OP1 analog, e.g.,  $10^{-7}$ M) under conditions sufficient to allow binding of the ligand to the receptor, followed by exposure to 32P-ATP (e.g., 100mCi in the presence of 10mM Tris-HCl (pH 7.6), 10mM MgCl2, 10mM MnCl2, 1mM dithiothreitol, 0.15M NaCl2, 0.1% Triton X-100 and 3% glycerol) and the amount of phosphorylation measured, e.g., by SDS polyacrylamide gel electrophoresis and autoradiography following immmunoprecipitation with antiphosphoserine, antiphosphothreonine or antiphosphotyrosine antibody (e.g., commercially available or made using standard antibody methodologies.) While a low level of autophosphorylation may be detected in the absence of ligand, incubation with ligand is anticipated to significantly increase (e.g., 5-20 fold increase) the amount of phosphorylation detected.

In another assay for detecting ligand-induced receptor autophosphorylation, involving intact cells, receptor DNA is transfected into a suitable host cell, e.g., a fibroblast, which then is grown under standard conditions to create a confluent monolayer of cells expressing the receptor on their cell surface. On the day of the experiment, cells are incubated with or without ligand (e.g., OP1 or OP1 analog, e.g.,  $10^{-7}$ M) at  $37^{\circ}$ C, and then quickly washed with a "stopping solution" containing ATP (e.g., 0.1M NaF, 4 mM EDTA, 10mM ATP, 10 mM sodium orthovanadate, 4mM sodium pyrophosphate). The cells then are frozen in a dry ice/ethanol bath, solubilized and the receptors immunoprecipitated, e.g., with an antireceptor antibody, as described herein. The immune complexes then are segregated, washed, separated by gel electrophoresis using standard procedures and transferred to a membrane for

Western blot analysis using standard procedures. Phosphorylation of the receptor then can be visualized by immunodetection with a suitable antibody (e.g., antiphosphoserine, antiphosphothreonine or antiphosphotyrosine), as described above. The bound antibody (e.g., bound antiphosphoserine, antiphosphothreonine or antiphosphotyrosine) then can be detected with <sup>125</sup>I labelled protein A, followed by autoradiography. The amount of phosphorylated receptor detected is anticipated to be significantly greater (5-20 fold increase) in receptors incubated with ligand than receptors exposed to ATP in the absence of ligand.

Ligand-induced receptor phosphorylation of exogenous substrates similarly can be assayed essentially using the methodology described herein. Here, a suitable substrate (e.g., a synthetic polypeptide containing serine, threonine or tyrosine amino acids) is provided to the receptor following ligand exposure and prior to incubation with ATP. The substrate subsequently can be segregated by immunoprecipitation with an antibody specific for the substrate, and phosphorylation detected as described above. As for autophosphorylation, the amount of phosphorylated substrate detected following ligand incubation is anticipated to be greater than that detected for substrates exposed to receptors in the absence of ligand.

Alternatively, a reporter gene construct can be created to assay transmembrane signal induction. Here, the expression control elements for an OP-1 inducible protein marker is fused to the open reading frame sequence for any reporter gene, and induction of the reporter gene expression then assayed. Useful reporter genes include the luciferase gene or GAL4 as well as other, easily characterizable markers.

#### Example 6. CHIMERIC RECEPTOR MOLECULES

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Chimeric receptor molecules, e.g., comprising an ALK or ALK analog extracellular and transmembrane region and, for example, part or all of an intracellular domain from another, different receptor or an intracellular domain from a different cell surface molecule, may be constructed using standard recombinant DNA technology and/or an automated DNA synthesizer to construct the desired sequence. As will be appreciated by persons skilled in

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the art, useful junctions include sequences within the transmembrane region and/or sequences at the junction of either the intracellular or the extracellular domains. Also envisioned are chimers where the extracellular domain or the intracellular domains themselves are chimeric sequences.

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Chimeric sequences are envisioned to be particularly useful in screening assays to determine candidate binding ligands (e.g., OP1 analogs, see below), where the non-receptor intracellular domain provides a suitable second messenger response system that is easy to detect. Potentially useful other second messenger response systems include those which, when activated, induce phosphoinositide hydrolysis, adenylate cyclase, guanylate cyclase or ion channels.

Chimeric receptor molecules have particular utility in gene therapy protocols. For example, a population of cells expressing a chimeric morphogen receptor molecule on their surface and competent for expressing a desired phenotype can be implanted in a mammal at a particular tissue locus. By careful choice of the ligand binding domain used on these receptors a physician can administer to the individual a morphogen agonist capable of:

(1) binding to the chimeric receptor alone and (2) stimulating the proliferation and/or differentiation of the implanted cells without affecting endogenous cell populations.

Example 7. CONSIDERATIONS FOR IDENTIFYING OTHER OP1 SPECIFIC RECEPTORS IN NUCLEIC ACID LIBRARIES

Identification of ALK Type I receptors that can bind OP-1 allows one to identify other morphogen receptor sequences in different species as well as in different tissues. The OP1-binding ALK sequences themselves can be used as a probe or the sequence may be modified to account for other potential codon usage (e.g., human codon bias.) Currently preferred probe sequences are those which encode the receptor's extracellular domain.

Probes based on the nucleic acid sequence of Seq. ID Nos.1, 3, 5 or 7 can be synthesized on commercially available DNA synthesizers, e.g. Applied Biosystems model 381A, using standard techniques, e.g. Gait, Oligonucleotide Synthesis: A Practical Approach, (IRL Press, Washington D.C., 1984). It is

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preferable that the probes are at least 8-50 bases long, more preferably 18-30 bases long. Probes can be labeled in a variety of ways standard in the art, e.g. using radioactive, enzymatic or colormetric labels as described, for example, by Berent et al, (May/June 1985) Biotechniques: 208-220; and Jablonski et al. (1986) Nucleic Acids Research 14: 6115-6128.

Preferably, low stringency conditions are employed when screening a library for morphogen receptor sequences using a probe derived from OP1binding receptor. Preferred ALK-specific probes are those corresponding to bases encoding the extracellular domain ("ECD"), or encoding a unique 10 (nonhomologous) sequence within the cytoplasmic domain. Useful probes may be designed from bases encoding the juxtamembrane region, for example. probe may be further modified to use a preferred species codon bias. Alternatively, probes derived from the serine/threonine kinase domain can be used to identify new members of the receptor kinase family which can be screened for OP1 binding using the methods described in Example 8.

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For example, for a probe of about 20-40 bases a typical prehybridization, hybridization, and wash protocol is as follows: (1) prehybridization: incubate nitrocellulose filters containing the denatured target DNA for 3-4 hours at 55oC in 5% Denhardt's solution, 6% SSC (20% SSC consists of 175g NaCl, 88.2 g sodium citrate in 800 ml  $\rm H_2O$  adjusted to pH. 7.0 with 10N NaOH), 0.1% SDS, and 100 mg/ml denatured salmon sperm DNA, (2) hybridization: incubate filters in prehybridization solution plus probe at 42oC for 14-48 hours, (3) wash; three 15 minutes washes in 6X SSC and 0.1% SDS at room temperature, followed by a final 1-1.5 minute wash in 6x SSC and 0.1% SDS at 55oC. Other equivalent procedures, e.g. employing organic solvents such as formamide, are well known in the art.

Alternatively, morphogen receptor-specific DNA can be amplified using a standard PCR (polymerase chain reaction) methodology such as the one disclosed herein, to amplify approximately 500 base pair fragments. As for the 30 hybridization screening probes described above, the primer sequences preferably are derived from conserved sequences in the serine/threonine kinase

domain. The primers disclosed herein, in Seq. ID Nos. 12-15 are envisioned to be particularly useful, particularly in combination..

Examples of useful PCR amplifications, including the use of the primers recited herein, are disclosed in ten Dijke, et al. (1993) Oncogene 8:2879-2887 and (1994) Science 264:101-104,, and which also describe the isolation protocols for ALK-1, ALK-2, ALK-3 and ALK-6.

## 7.1 TISSUE DISTRIBUTION OF MORPHOGEN RECEPTORS

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Determining the tissue distribution of OP1-specific receptors can be used to identify tissue and cell sources which express these receptors, to identify new, related OP1-specific receptor molecules, as well as to identify target tissues for OP1-receptor interactions under naturally occurring conditions. The OP-1 specific receptor molecules (or their mRNA transcripts) readily are identified in different tissues using standard methodologies and minor modifications thereof in tissues where expression may be low. For example, protein distribution can be determined using standard Western blot analysis or immunohistological detection techniques, and antibodies specific to the morphogen receptor molecules of interest. Similarly, the distribution of morphogen receptor transcripts can be determined using standard Northern hybridization protocols and transcript-specific probes or by in situ hybridization.

Any probe capable of hybridizing specifically to a transcript, and distinguishing the transcript of interest from other related transcripts can be used. Because the morphogen receptors described herein likely share high sequence homology in their intracellular domains, the tissue distribution of a specific morphogen receptor transcript may best be determined using a probe specific for the extracellular domain of the molecule. For example, a particularly useful ALK-specific probe sequence is one derived from a unique portion of the 5' coding sequence, the sequence corresponding to the juxtamembrane region, or the 5' or 3' noncoding sequences. The chosen fragment then is labelled using standard means well known and described in the art and herein.

Using these receptor-specific probes, which can be synthetically engineered or obtained from cloned sequences, receptor transcripts can be identified and localized in various tissues of various organisms, using standard methodologies well known to those having ordinary skill in the art. A detailed description of a suitable hybridization protocol is described in Ozkaynak, et al., (1991) Biochem. Biophys. Res. Commn. 179:116-123, and Ozkaynak, et al. (1992) J Biol. Chem. 267:25220-25227. Briefly, total RNA is prepared from various tissues (e.g., murine embryo and developing and adult liver, kidney, testis, heart, brain, thymus, stomach) by a standard methodologies such as by the method of Chomczynski et al. ((1987) Anal. 10 Biochem 162:156-159) and described below. Poly (A)+ RNA is prepared by using oligo (dT)-cellulose chromatography (e.g., Type 7, from Pharmacia LKB Biotechnology, Inc.). Poly (A) + RNA (generally 15 mg) from each tissue is fractionated on a 1% agarose/formaldehyde gel and transferred onto a Nytran membrane (Schleicher & Schuell). Following the transfer, the membrane is baked at 80oC and the RNA is cross-linked under UV light (generally 30 seconds at 1  $mW/cm^2$ ). Prior to hybridization, the appropriate probe is denatured by heating. The hybridization is carried out in a lucite cylinder rotating in a roller bottle apparatus at approximately 1 rev/min for approximately 15 hours at 37oC using a hybridization mix of 40% formamide, 5 x Denhardts, 5 x SSPE, and 0.1% SDS. Following hybridization, the non-specific counts are washed off the filters in 0.1 x SSPE, 0.1% SDS at 50oC.

## Example 8. Demonstration That ALK-2, ALK-3 and ALK-6

### are OP1-Binding Receptors

25 The tissue morphogenic proteins OP1 and BMP4 were tested for specific binding interaction with the ALK receptors in receptor-transfected cells (where the receptor is over-expressed), and in nontransfected cells. It previously was known that ALK-5 interacted specifically with TGFB1 and ALK-2 and ALK-4 interacted specifically with activin. In the experiment, complexes were crosslinked and immuno-precipitated with an ALK-specific antibody as described below. To date, no binding with ALK-1 under the conditions of this protocol have been detected.

Binding and affinity cross-linking using disuccinimidyl suberate (Pierce Chemical Co.) were performed using standard methods (e.g., Franzen et al. (1993) <u>Cell 75</u>:681-692 and Ichijo et al. (1990) <u>Exp. Cell Res.</u> <u>187</u>:10995-11000.) A typical protocol is described below. Modifications from this protocol for individual experiments were standard changes anticipated to produce the same result as for the recited procedure. Briefly, cells in multi-well plates were washed with binding buffer (e.g., phosphate buffered saline containing 0.9 mM CaCl2, 0.49mM MgCl2 and 1 mg/ml bovine serum albumin (BSA)), incubated on ice in the same buffer with labelled ligand, in the presence and absence of excess unlabelled ligand for sufficient time for the reaction to equilibrate (e.g., 3 hours.) Cells were washed and the crosslinking was done in the binding buffer without BSA together with 0.28 mM disuccinimidyl suberate for 15 min on ice. Cells were harvested by addition of 1 ml of detachment buffer (10 mM Tris-HCl, pH 7.4, 1 mM EDTA, 10% glycerol, 0.3 mM PMSF.) Cells then were pelleted by centrifugation, then resuspended in 50 ml of solubilization buffer (125 mM NaCl, 10mM Tris-HCl, pH 7.4, 1mM EDTA, 1% Triton X-100, 0.3 mM PMSF, 1% Trasylol) and incubated for 40 minutes on ice. Cells were centrifuged again and supernatants subjected to analysis by standard SDS-gel electrophoresis using 4%-15% polyacrylamide gels, followed by autoradiography.

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Cell lysates obtained following affinity cross-linking via the general protocol described above were immunoprecipitated using antisera against ALKs (e.g., raised against the ALK juxtamembrane region), or directly analyzed by SDS-gel electrophoresis using gradient gels consisting of 5-12% or 5-10% polyacrylamide. The gels were fixed and dried, and then subjected to autoradiography or analysis using phosphorImager (Molecular Dynamics).

# 8.1 Binding of OP-1 and/or BMP-4 to ALKs in Transfected Cells.

COS-1 cells transfected with ALK cDNA were tested for the binding of <sup>125</sup>I-OP-1 and <sup>125</sup>I-BMP-4, in the presence or absence of co-transfected Type II receptor DNA: daf-4 cDNA or ActRII (Estevez et al. (1993) <u>Nature 365</u>:644-649 and Attisano et al. (1992) <u>Cell 68</u>:97-108, disclosing the DNA sequence for these Type II receptors and the disclosure of which is incorporated herein by

reference.) Since the cross-linked complexes were sometimes difficult to visualize because of high background, samples were immunoprecipitated by antisera against each ALK. The results are presented in Table I below. In the Table, "N/T" means "not tested". Binding was specific as determined by standard competition assays. The values represented by "+/-", "+", "++", "++", and "-" are all qualitative descriptors of the relative amount of radioactivity measured when the crosslinked molecules were gel electrophoresed and subjected to autoradiography. More radioactivity measured indicates a stronger binding interaction detected. In the Table the strength of binding interaction is as follows: +++ > ++ > ++ > +- > -.

TABLE I

15		125	OP1			125 <sub>I BM</sub>	P4	
13		+daf4	-daf4	_ActRII		+daf4	-daf4	+ActRII
	ALK1	_	_	-	ALK1	-	_	N/T
	ALK2	++	+/-	++	ALK2	_ '	-	N/T
	ALK3	++	_	+/-	ALK3	+++	++	N/T
	ALK4	_	-	-	ALK4	-	_	N/T
	ALK5	-	-	-	ALK5	-	-	N/T
	ALK6	+++	++ '	+	ALK6	+++	+++	N/T

In the absence of daf-4, OP-1 bound to ALK-6, whereas BMP-4 bound to ALK-3 and ALK-6. Weaker binding of OP-1 to ALK-2 was also observed. Other

20 ALKs did not bind OP-1 or EMP-4 in the absence of Daf-4. When ALK cDNAs were co-transfected with the daf-4 cDNA, increased binding of OP-1 to ALK-2 and ALK-6 was seen. In addition, ALK-3 also was found to bind OP-1 in the co-transfected cells. Similarly, increased binding of BMP-4 to ALK-3 and ALK-6 could be observed. Co-transfection of two different types of ALKs did not further increase the binding of OP-1 or BMP-4. In cells co-transfected with the DNA for ActRII and ALK-2, ALK-3 or ALK-6, OP1-receptor binding was enhanced.

The sizes of the cross-linked complexes were slightly higher for ALK-3 than for ALK-2 and ALK-6, consistent with its slightly larger size. Complexes

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of about 95 kDa as well as multiple components of 140-250 kDa were also coimmunoprecipitated with certain of the ALKs.

In standard competition assays performed with the Type I receptors in the presence and absence of the Type II receptors, the binding of OP-1 and EMP-4 could be competed with excess amounts of unlabeled OP-1, verifying the binding specificity of these interactions where they occurred.

These results demonstrate that ALK-2, ALK-3 and ALK-6 can serve as

Type I receptors for OP-1. Notably, ligand binding apparently can be enhanced in the presence of Type II receptors. Moreover, OP-1 is able to interact with both a "bone morphogen" Type II receptor (daf 4) and an "activin" Type II receptor (ActRII), whereas, for example, activin only interacts with the ActRII Type II receptor. The data indicate that OP1 has a broader spectrum of receptor (Type I and Type II) binding affinities than do other tissue morphogenic proteins, or other members of the TGF-ß family. It is anticipated that OP1 will have specific binding interactions with other "activin-binding" or "bone morphogen-binding" Type II receptors.

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The ability of OP-1 to bind to Type I, Type II receptors having binding specificity for activin or BMP4 but not TGF-B, indicates that OP-1 and OP-1 analogs will be useful as competitors of activin or BMP4 binding to cell surface receptors. In particular, OP-1 and OP-1 analogs will be useful for competing with activin-ALK-2 binding and/or activin-ALK-2/ActRII (or other Type II) receptor binding; and for competing with BMP4 (or BMP2)-ALK-6 binding, and/or BMP2/4-ALK-6/daf 4 (or other Type II) binding. The OP-1 competitors may act as antagonists (e.g., binding competitors unable to induce the signal transduction cascade upon binding) or as agonists (e.g., able both to bind and induce the signal transduction cascade).

8.2 Identification of OP1-Specific Receptors in Nontransfected Cells

ALK-5 has been shown to bind TGF-61, and ALKs 2, 4 bind activin A with
high affinities in nontransfected cells (ten Dijke, Oncogene, (1993\_; Science,

(1994) referenced herein above.) In the present experiment, the binding
affinity of OP1 and/or BMP4 to receptors in nontransfected cells was

demonstrated as follows. The results corroborate the transfected cell data,

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verifying that OP1 interacts specifically with ALK-2, ALK-3 and ALK-6, but not ALK-4 or ALK-5.

MC3T3-E1 osteoblasts are well characterized cells known to respond to OP-1 and BMP-4 in the induction of alkaline phosphatase activity (Paralkar (1991) PNAS 8: 3397-3401.) These cells were affinity labeled using \$125\_{I-OP-1}\$ as described herein above, and cross-linked complexes of about 75 kDa were seen, which were immunoprecipitated only with the ALK-2 antiserum. Tera-2 teratocarcinoma cells and Mv1Lu cells responded to OP-1 as measured by production of plasminogen activator inhibitor-1 (PAI-1). Similar to MC3T3-E1 cells, cross-linked complexes using \$125\_{I-OP-1}\$ in Tera-2 teratocarcinoma were immunoprecipitated only by the ALK-2 antiserum.

On the other hand, cross-linked complexes using \$125\$I-OP-1 to Mv1Lu cells were immunoprecipitated by ALK-2 as well as ALK-3 and ALK-6 antisera. Mv1Lu cells are known to express ALK-4 and ALK-5 (Ebner (1993) Science \$260:1344-1348)\$, but cross-linked complexes with \$125\$I-OP-1 were not precipitated by antisera against these receptors. Similarly, cross-linked complexes in U-1240 MG glioblastoma cells were immunoprecipitated by ALK-2 and ALK-6 antisera, and weakly by ALK-3 antiserum. In contrast, cross-linking of \$125\$I-OP-1 to AG1518 human foreskin fibroblasts yielded weak immunoprecipitated components only by ALK-3 antiserum. Type II receptor-like components of about 95 kDa as well as high molecular weight complexes of 140-250 kDa co-immunoprecipitated with certain ALKs in the Tera-2 cells, Mv1Lu cells and U-1240 MG cells.

Receptors for BMP-4 have also been investigated using nontransfected cells. Cross-linked complexes using <sup>125</sup>I-BMP-4 to MC3T3-E1 cells and AG1518 human foreskin fibroblasts were immunoprecipitated only by ALK-3. On the other hand, cross-linking of <sup>125</sup>I-BMP-4 to Tera-2 cells did not yield any immunoprecipitated components by antisera against ALKs.

125<sub>I-OP-1</sub> and/or <sup>125</sup><sub>I-BMP-4</sub> also were demonstrated by affinity cross-30 linking to interact specifically with receptors in other BMP-responsive cells, e.g., MG-63 osteosarcoma cells and PC12 pheochromocytoma cells. A summary of

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the binding of ALKs to OP-1 or BMP-4 in different cell types is shown in Table II, below. In the Table, "N/T" means not tested, and receptors presented in brackets indicate comparatively lower quantities of radioactive complexes detected.

#### TABLE II

Cell lines	Binding of OP-1	Binding of BMP-4
Mouse osteoblasts	ALK-2	ALK-3
(MC3T3-E1	•	
Mink lung epithelial cells (Mv1Lu)	ALK-2, -3, -6	N/T
cerrs (MVILu)	ALK2. [-3]6	N/T
Human glioblastoma		
	ALK2	
Human teratocarcinoma (Tera-2)		
	[ALK3]	ALK3
Human foreskin		•
fibroblasts (AG1518)		
	ALK2, [-3]	N/T.
Rat osteosarcoma (ROS17/2.8)		

Example 9. OP1, OP1-SPECIFIC RECEPTOR ANALOG SCREENING ASSAYS

The present invention is useful to determine whether a ligand, such as a known or putative drug, is capable of binding to and/or activating an OP1-specific cell surface receptor as described herein. Ligands capable of specific binding interaction with a given OP1-specific receptor (e.g., ALK-2, ALK-3, ALK-6) are referred to herein as OP1 analogs and can be used for therapeutic and diagnostic applications. Some analogs will have the ability to stimulate morphogenetic activity in the cell, mimicking both the receptor binding and signal transducing activity of OP1. These are referred to OP1 agonists or mimetics. Others will have strong binding affinity but will not stimulate morphogenesis, these are OP1 antagonists. The analogs can be amino acid-based, or they can be composed of non-proteinaceous chemical structures.

The methods and kits described below similarly can be used to identify
OP1-specific receptor analogs, capable of mimicking the binding affinity of
ALK-2,ALK-3 or ALK-6 for OP1. The analogs can be provided to a mammal to

interact with serum-soluble OP1, effectively sequestering the protein and modulating its availability for cell surface interaction.

Transfection of an isolated clone encoding a morphogen receptor into the cell systems described above provides an assay system for the ability of ligands to bind to and/or to activate the receptor encoded by the isolated DNA molecule. Transfection systems, such as those described above, are useful as living cell cultures for competitive binding assays between known or candidate drugs and ligands which bind to the receptor and compete with the binding of known morphogens, which are labeled by radioactive, enzymatic, spectroscopic or other reagents. Membrane preparations containing the receptor and isolated from transfected cells are also useful in these competitive binding assays. Alternatively, and currently preferred, purified receptor molecules or their ligand binding extracellular domains can be plated onto a microtiter well surface, in a modification of a sandwich assay, e.g., as a competition assay, such as an RIA, described above. Finally, as described above, solution assays, and using only the receptor extracellular domain, also may be used to advantage in these assays. Functional assays of second messenger systems or their sequelae in transfection systems act as assays for binding affinity and efficacy in the activation of receptor function or efficacy in the antagonism of receptor function. Such a transfection system constitutes a \*drug discovery system", useful for the identification of natural or synthetic compounds with potential for drug development that can be further modified or used directly as therapeutic compounds to activate or inhibit the natural functions of the receptor encoded by the isolated DNA molecule.

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Once such candidate drugs (e.g., OP-1 or receptor-binding analogs thereof) are identified, they can be produced in reasonable, useful quantities using standard methodologies known in the art. Amino acid-based molecules can be encoded by synthetic nucleic acid molecules, and expressed in a recombinant expression system as described herein above or in the art. Alternatively, such molecules can be chemically synthesized, e.g., by means of an automated peptide synthesizer, for example. Non-amino acid-based molecules can be produced by standard organic chemical synthesis procedures. Where the

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candidate molecule is of undetermined structure, or composition, its composition readily can be determined by, for example, mass spectroscopy. Two approaches to identifying analogs typically are practiced in the art: high flux screens and rational design of ligand mimetics. High flux screens typically screen naturally sourced materials or chemical banks for their ability to bind a protein of interest, here, e.g., the receptor. Typically, compounds are obtained from a range of sources, e.g., chemical banks, microbial broths, plant and animal extracts, and the like. In a high flux screen typically, purified receptor, preferably the soluble, ligand binding extracellular domain, is plated onto a microtiter well surface and a standard volume of a sample solution to be tested then is added. Also added is a standard volume having a known quantity of a purified ligand known to bind the receptor with specificity. Preferably the ligand is labelled with a substance that is readily detectable by automated means (e.g., radiolabel, chromophoric, 15 fluorometric, enzymatic or spectroscopic label.) The wells then are washed and the amount of label remaining after washing or the amount of label remaining associated with the receptor then is detected. Positive scores are identified by the ability of the test substance to prevent interaction of the labelled ligand with the receptor. The screening assays can be performed without undue experimentation, using standard molecular and cell biology tools in common use in the art. For example, screening assays can be performed in standard 96-well plates. Fifteen such plates reasonably can be set up at a time to perform multiple screening assays in parallel. Thus, with only 10-11 reiterations of the screening assay 15,625 (56) compounds can be screened for their binding affinity. Even allowing for a maximum incubation time of 2 25 hours, all 15,625 compounds reasonably can be assayed in a matter of days.

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High flux screens exploit both the high degree of specificity of the labelled ligand for its receptor, as well as high throughput capacity of computer driven robotics and computer handling of data. Candidate analogs identified in this manner, then can be analyzed structurally and this information used to design and to synthesize analogs having enhanced potency, increased duration of action, increased selectivity and reduced side effects.

Candidates also can be used in a rational design program as described below. Finally, candidate analogs also can be tested to determine morphogenetic effect, if any, as described below.

The second approach to the identification of analogs uses a rational design approach to create molecules capable of mimicking the binding effect of OP1 with an OP1-specific receptor. Here the relevant structure for receptor binding is analyzed to identify critical sequences and structures necessary for binding activity and this information can be used to design and synthesize minimal size morphogen analogs. As for candidate compounds in the high flux assay, design candidates can be tested for receptor binding activity as described above. As described above, a candidate sequence can be further modified by, for example standard biological or chemical mutagenesis techniques to create a candidate derivative having, for example, enhanced binding affinity or another preferred characteristic.

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Antibodies capable of interacting specifically with the receptor and competing with OP1 binding also can be used as an analog. Antibodies can be generated as described above.

OP1 analogs may be evaluated for their ability to mimic OP1 or to inhibit OP1 binding (e.g., agonists or antagonists) by monitoring the effect of the analogs on cells bearing an OP1-specific receptor (e.g., ALK-2, ALK-3 or ALK-6.) OP1 agonists are anticipated to have utility in any application where tissue morphogenesis is desired, such as in the regeneration of damaged tissue resulting from mechanical or chemical trauma, degenerative diseases, tissue destruction resulting from chronic inflammation, cirrhosis,

inflammatory diseases, cancer and the like, and in the regeneration of tissues, organs and limbs. OPI antagonists are envisioned to have utility in applications where tissue morphogenesis is to be limited as, for example, in the treatment of malignant transformations including, but not limited to, osteosarcomas and Paget's disease.

30 Several exemplary systems for assaying the ability of a candidate analog transduce an OP-1-specific signal across the cellular membrane are described below.

#### 9.1 Induction of Osteoblast Differentiation Markers

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For example, OP1 is known to preferentially induce differentiation of progenitor cells, including embryonic mesenchymal cells and primary osteoblasts (see, for example, PCT US92/07432) As one example, OP1 analogs can be tested for their ability to induce differentiation of primary osteoblasts, by measuring the ability of these analogs to induce production of alkaline phosphatase, PTH-mediated cAMP and osteocalcin, all of which are induced when primary osteoblasts are exposed to OP-1, 60A or DPP.

Briefly, the assays may be performed as follows. In this and all examples involving osteoblast cultures, rat osteoblast-enriched primary cultures preferably are used. Although these cultures are heterogeneous in that the individual cells are at different stages of differentiation, these cultures are believed to more accurately reflect the metabolism and function of osteoblasts in vivo than osteoblast cultures obtained from established cell lines. Unless otherwise indicated, all chemicals referenced are standard, commercially available reagents, readily available from a number of sources, including Sigma Chemical, Co., St. Louis; Calbiochem, Corp., San Diego and Aldrich Chemical Co., Milwaukee.

Rat osteoblast-enriched primary cultures are prepared by sequential

collagenase digestion of newborn suture-free rat calvaria (e.g., from 1-2 dayold animals, Long-Evans strain, Charles River Laboratories, Wilmington, MA),
following standard procedures, such as are described, for example, in Wong et
al., (1975) PNAS 72:3167-3171. Rat osteoblast single cell suspensions then
are plated onto a multi-well plate (e.g., a 24 well plate) at a concentration
of 50,000 osteoblasts per well in alpha MEM (modified Eagle's medium, Gibco,
Inc., Long Island) containing 10% FBS (fetal bovine serum), L-glutamine and
penicillin/streptomycin. The cells are incubated for 24 hours at 37°C, at
which time the growth medium is replaced with alpha MEM containing 1% FBS and
the cells incubated for an additional 24 hours so that the cells are in serumdeprived growth medium at the time of the experiment.

Alkaline Phosphatase Induction of Osteoblasts

The cultured cells in serum-free medium are incubated with OP1, OP1 analog or a negative control, using a range of concentrations. For example, 0.1, 1.0, 10.0, 40.0 or 80.0 ng OP-1/ml medium typically are used. 72 hours after the incubation period the cell layer is extracted with 0.5 ml of 1% Triton X-100. The resultant cell extract then, is centrifuged, and 100 ml of the extract is added to 90 ml of paranitrosophenylphospate (PNPP)/glycerine mixture and incubated for 30 minutes in a 37°C water bath and the reaction stopped with 100 ml NaOH. The samples then are run through a plate reader (e.g., Dynatech MR700 plate reader, and absorbance measured at 400 nm, using p-nitrophenol as a standard) to determine the presence and amount of alkaline phosphate activity. Protein concentrations are determined by the Biorad method. Alkaline phosphatase activity is calculated in units/mg protein, where 1 unit=1 nmol p-nitrophenol liberated/30 minutes at 37°C. OP-1 induces a five -fold increase in the specific activity of alkaline phosphate by this method. Agonists are expected to have similar induction effects. Antagonists should inhibit or otherwise interfere with OP1 binding, and diminished alkaline phophatase induction should result when the assay is performed with an antagonist in the presence of a limiting amount of OP1.

#### Induction of PTH-Mediated cAMP.

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The effect of a morphogen analog on parathyroid hormone-mediated cAMP production in rat osteoblasts in vitro may be demonstrated as follows.

Rat osteoblasts are prepared and cultured in a multiwell plate as described above. The cultured cells then are divided into three groups: (1) wells which receive, for example, 1.0, 10.0 and 40.0 ng OP-1/ml medium); (2) wells which receive the candidate analog at various concentration ranges; and (3) a control group which receives no additional factors. The plate is then incubated for another 72 hours. At the end of the 72 hours the cells are treated with medium containing 0.5% bovine serum albumin (BSA) and 1mM 3-isobutyl-1-methylxanthine for 20 minutes followed by the addition into half of the wells of human recombinant parathyroid hormone (hPTH, Sigma, St. Louis) at a concentration of 200 ng/ml for 10 minutes. The cell layer then is extracted from each well with 0.5 ml of 1% Triton X-100. The cAMP levels then are

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determined using a radioimmunoassay kit (e.g., Amersham, Arlington Heights, Illinois). OP-1 doubles cAMP production in the presence of PTH. Agonists are expected to have similar induction effects. Antagonists are expected to inhibit or otherwise interfere with OP1 binding, and diminished cAMP production should result when the assay is performed with an antagonist in the presence of limiting the amount of OP1.

#### Induction of Osteocalcin Production

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in vitro.

Osteocalcin is a bone-specific protein synthesized by osteoblasts which plays an integral role in the rate of bone mineralization in vivo.

Circulating levels of osteocalcin in serum are used as a marker for osteoblast activity and bone formation in vivo. Induction of osteocalcin synthesis in osteoblast-enriched cultures can be used to demonstrate morphogenic efficacy

Rat osteoblasts are prepared and cultured in a multi-well plate as above. In this experiment the medium is supplemented with 10% FBS, and on day 2, cells are fed with fresh medium supplemented with fresh 10 mM bglycerophosphate (Sigma, Inc.). Beginning on day 5 and twice weekly thereafter, cells are fed with a complete mineralization medium containing all of the above components plus fresh L(+)-ascorbate, at a final concentration of 50mg/ml medium. OP1 or OP1 analog then is added to the wells directly, e.g., in 50% acetonitrile (or 50% ethanol) containing 0.1% trifluoroacetic acid (TFA), at no more than 5ml OP1/ml medium. Control wells receive solvent vehicle only. The cells then are re-fed and the conditioned medium sample diluted 1:1 in standard radioimmunoassay buffer containing standard protease inhibitors and stored at -20°C until assayed for osteocalcin. Osteocalcin synthesis is measured by standard radioimmunoassay using a commercially available osteocalcin-specific antibody and can be confirmed by Northern blot analysis to calculate the amount of osteocalcin mRNA produced in the presence and absence of OP-1 or OP1 analog. OP-1 induces a dose-dependent increase in osteocalcin production (5-fold increase using 25 mg of OP-1 protein/ml), and a 20-fold increase in osteocalcin mRNA. Agonists are expected to have similar induction effects; antagonists are expected to inhibit or otherwise interfere

with OP1 binding, thereby substantially interfering with osteocalcin induction in the presence of a limiting amount of OP1.

Mineralization is determined on long term cultures (13 day) using a modified von Kossa staining technique on fixed cell layers: cells are fixed in fresh 4% paraformaldehyde at 230 C for 10 min, following rinsing cold 0.9% NaCl. Fixed cells then are stained for endogenous alkaline phosphatase at pH 9.5 for 10 min, using a commercially available kit (Sigma, Inc.) Purple stained cells then are dehydrated with methanol and air dried. After 30 min incubation in 3% AgNO<sub>3</sub> in the dark, H<sub>2</sub>O-rinsed samples are exposed for 30 sec to 254 nm UV light to develop the black silver-stained phosphate nodules. Individual mineralized foci (at least 20 mm in size) are counted under a dissecting microscope and expressed as nodules/culture. OP-1 induces a 20-fold increase in initial mineralization rate. Agonists are expected to have similar induction effects; antagonists are expected to inhibit or otherwise interfere with OP1 binding, thereby inhibiting mineralization induction in the presence of a limiting amount of OP1:

#### 9.2 Induction of a Constructed Reporter Gene

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Alternatively, a reporter gene construct can be used to determine the ability of candidate molecule to induce signal transduction across a membrane following receptor binding. For example, PAI-1 protein, (Plasminogen Activator Inhibitor-1) expression can be induced by OP-1 in Mv1Lu- cells (see above). Also, as demontrated above, these cells express ALK-2, -3 and -6 surface receptors. In addition, preliminary studies indicate that ALK-1, when overexpressed in a chemically mutagenized derivative of these cells, also apparently mediates PAI-1 induction in the presence of OP1.

Accordingly, PAI-1 promoter elements can be fused to a reporter gene and induction of the reporter gene monitored following incubation of the transfected cell with a candidate analog. As one example, the luciferase reporter gene can be used, in, for example, the construct p3TP-Lux described by Wrana et al. (1992) Cell 71: 1003-1014 and Attisano et al. (1993) Cell 74: 671-680. This reporter gene construct includes a region of the human PAI-1

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gene promoter in combination with three sets of tetradecanoyl phorbol - acetate responsive elements upstream of the lucifrase open reading frame.

In a typical assay, transfected cells starved in DMEM containing 0.1% fetal bovine serum and antibiotics (e.g., 100 units/ml penicillin and 50  $\mu$ g/ml streptomycin) for 6 hrs., and then exposed to ligand for 24 hr. Luciferase activity in the cell lysate then is measured using a luminometer in the luciferase assay system, according to the manufacturer's protocol(Promega). In MvlLu mutant cells, "R mutant" cells co-transfected with ALK-2 and Act RII, OP1 mediated induction of luciferase activity.

10 9.3 Inhibition of Epithelial cell proliferation

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OP1 is known to inhibit epithelial cells. Thus, the ability of a candidate analog to inhibit cell proliferation, as measured by <sup>3</sup> H-thymidine uptake by an epithelial cell can be used in an assay to evaluate signal transduction activity of the candidate. Analogs competent to inhibit epithelial cell growth are contemplated to have particular utility in therapeutic applications where limitation of a proliferating cell population is desired. Such applications include chemotherapies and radiation therapies where limiting the growth of a normally proliferating population of cells can protect these cells from the cytotoxic effects of these cancer therapies. (see e.g., W094/06420). In addition, psoriasis and other tissue disorders resulting from uncontrolled cell proliferation, including benign and malignant neoplasties, can be modulated by use of an OP1 analog.

As an example, mink lung epithelial cell growth is inhibited by OP-1. (see, PCT US93/08885; WO94/06420.) As described above, derivatives of these cells [e.g., "R-4 mutants", clone 4-2, Laiho et al. (1990) <u>J. Biol. Chem. 265</u>: 18518-18524] can be transfected with DNA encoding OP1-specific receptors and induced to express these receptors. The transfected cells, then can be assayed for a candidate analog's ability to block cell growth. As one example, when R-4 cells are transfected with ALK-3 under a Zn<sup>2+</sup>-inducible promoter, and induced to express the receptor following induction with ZnCl<sub>2</sub>, cell growth can be inhibited in the presence of OP1 in a dose dependent

manner. Preliminary experiments with ALK-1 indicates that this receptor also can mediate this OP-1-specific effect.

In a typical assay, cells are seeded in 24-well cell culture plates at a density of 10<sup>4</sup> cells per well in DMEM with 10 % FBS, and incubated overnight. The medium is replaced with DMEM containing 0.2 % FBS and 100uM ZnLC2, and the cells are incubated for 5 h, after which the medium is replaced with fresh DMEM containing 0.2 % FBS, 100uM ZnCL2 and various concentrations of OP-1 or an analog candidate. After 16 h of incubation, 0.25 ci of [3H)thymidine (Amersham) are added and the cells incubated for an additional 2 h. Thereafter, the cells are fixed in 10% trichloroacetic acid for more than 15 min on ice, and solubilized with 1 M NaOH. The cell extracts are neutralized with 1 M HCl and 3H radioactivity determined in a liquid scintillation counter.

Example 10. SCREENING ASSAY FOR COMPOUNDS WHICH ALTER ENDOGENOUS OP1 RECEPTOR EXPRESSION LEVELS

Candidate compound(s) which can be administered to affect the level of a given endogenous OP1 receptor can be found using the following screening assay, in which the level of OP1 receptor production by a cell type which produces measurable levels of the receptor is determined by incubating the cell in culture with and without the candidate compound, in order to assess the effects of the compound on the cell. This also can be accomplished by detection of the OP1 receptor either at the protein level by Western blot or immunolocalization, or at the RNA level by Northern blot or in situ hydridization. The protocol is based on a procedure for identifying compounds which alter endogenous levels of OP1 expression, a detailed description also may be found in PCT US 92/07359, incorporated herein by reference.

Cell cultures of, for example, bone, brain, intestine, lung, heart, eye, breast, gonads, kidney, adrenals, urinary bladder, brain, or other organs, may be prepared as described widely in the literature. For example, kidneys may be explanted from neonatal or new born or young or adult rodents (mouse or rat) and used in organ culture as whole or sliced (1-4 mm) tissues.

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Primary tissue cultures and established cell lines, also derived from kidney, adrenals, urinary, bladder, brain, mammary, or other tissues may be established in multiwell plates (6 well or 24 well) according to conventional cell culture techniques, and are cultured in the absence or presence of serum for a period of time (1-7 days). Cells can be cultured, for example, in Dulbecco's Modified Eagle medium (Gibco, Long Island, NY) containing serum (e.g., fetal calf serum at 1%-10%, Gibco) or in serum-deprived medium, as desired, or in defined medium (e.g., containing insulin, transferrin, glucose, albumin, or other growth factors).

Cell samples for testing the level of OP1 receptor production are collected periodically and evaluated for receptor production by immunoblot analysis (Sambrook et al., eds., 1989, Molecular Cloning, Cold Spring Harbor Press, Cold Spring Harbor, NY), or, alternatively, a portion of the cell culture itself can be collected periodically and used to prepare polyA+ RNA for mRNA analysis by Northern blot analysis. To monitor de novo receptor synthesis, some cultures are labeled according to conventional procedures with an 35S-methionine/35S-cysteine mixture for 6-24 hours and then evaluated to quantitate receptor synthesis by conventional immunoassay methods.

Alternatively, anti-receptor antibodies may be labelled and incubated with the cells or cell lysates, and the bound complexes detected and quantitated by conventional means, such as those described hereinabove. Northern blots may be performed using a portion of the OP1 receptor coding sequence to create hybridization probes, and following the RNA hybridization protocol described herein.

25 Example 11. GENERAL FORMULATION/ ADMINISTRATION CONSIDERATIONS

The analogs and constructs described herein can be provided to an individual as part of a therapy to enhance, inhibit, or othewise modulate the in vivo binding interaction between OP1 and one or more OP1-specific cell surface receptors. The molecules then comprise part of a pharmaceutical composition as described herein below and can be administered by any suitable means, preferably directly or systemically, e.g., parenterally or orally. Where the therapeutic molecule is to be provided directly (e.g., locally, as

by injection, to a desired tissue site), or parenterally, such as by intravenous, subcutaneous, intramuscular, intraorbital, ophthalmic, intraventricular, intracranial, intracapsular, intraspinal, intracisternal, intraperitoneal, buccal, rectal, vaginal, intranasal or by aerosol administration, the therapeutic preferably comprises part of an aqueous solution. The solution preferably is physiologically acceptable so that in addition to delivery of the desired morphogen to the patient, the solution does not otherwise adversely affect the patient's electrolyte and volume balance. The aqueous medium for the therapeutic molecule thus may comprise normal physiologic saline (0.9% NaCl, 0.15M), pH 7-7.4 or other pharmaceutically acceptable salts thereof.

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Useful solutions for oral or parenteral administration can be prepared by any of the methods well known in the pharmaceutical art, described, for example, in Remington's Pharmaceutical Sciences, (Gennaro, A., ed.), Mack Pub., 1990. Formulations may include, for example, polyalkylene glycols such as polyethylene glycol, oils of vegetable origin, hydrogenated naphthalenes, and the like. Formulations for direct administration, in particular, can include glycerol and other compositions of high viscosity. Biocompatible, preferably bioresorbable polymers, including, for example, hyaluronic acid, collagen, tricalcium phosphate, polybutyrate, polylactide, polyglycolide and lactide/glycolide copolymers, may be useful excipients to control the release of the morphogen in vivo.

Other potentially useful parenteral delivery systems for these therapeutic molecules include ethylene-vinyl acetate copolymer particles, osmotic pumps, implantable infusion systems, and liposomes. Formulations for inhalation administration may contain as excipients, for example, lactose, or can be aqueous solutions containing, for example, polyoxyethylene-9-lauryl ether, glycocholate and deoxycholate, or oily solutions for administration in the form of nasal drops, or as a gel to be applied intranasally.

Alternatively, the morphogens described herein may be administered orally.

The therapeutic molecules also can be associated with means for targeting the therapeutic to a desired tissue. For example, tetracycline and diphosphonates (bisphosphonates) are known to bind to bone mineral, particularly at zones of bone remodeling, when they are provided systemically in a mammal. Accordingly, these molecules may be included as useful agents for targeting therapeutics to bone tissue. Alternatively, an antibody or other binding protein that interacts specifically with a surface molecule on the desired target tissue cells also can be used. Such targeting molecules further can be covalently associated to the therapeutic molecule e.g., by chemical crosslinking, or by using standard genetic engineering means to create, for example, an acid labile bond such as an Asp-Pro linkage. Useful targeting molecules can be designed, for example, using the single chain binding site technology disclosed, for example, in U.S. Pat. No. 5,091,513.

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Finally, therapeutic molecules can be administered alone or in combination with other molecules known to have a beneficial effect on tissue morphogenesis, including molecules capable of tissue repair and regeneration and/or inhibiting inflammation. Examples of useful cofactors for stimulating bone tissue growth in osteoporotic individuals, for example, include but are not limited to, vitamin D<sub>3</sub>, calcitonin, prostaglandins, parathyroid hormone, dexamethasone, estrogen and IGF-I or IGF-II. Useful cofactors for nerve tissue repair and regeneration can include nerve growth factors. Other useful cofactors include symptom-alleviating cofactors, including antiseptics, antibiotics, antiviral and antifungal agents and analgesics and anesthetics.

Therapeutic molecules further can be formulated into pharmaceutical compositions by admixture with pharmaceutically acceptable nontoxic excipients and carriers. As noted above, such compositions can be prepared for parenteral administration, particularly in the form of liquid solutions or suspensions; for oral administration, particularly in the form of tablets or capsules; or intranasally, particularly in the form of powders, nasal drops or aerosols. Where adhesion to a tissue surface is desired the composition may include the morphogen dispersed in a fibrinogen-thrombin composition or other bioadhesive such as is disclosed, for example in PCT US91/09275, the

disclosure of which is incorporated herein by reference. The composition then can be painted, sprayed or otherwise applied to the desired tissue surface.

The compositions can be formulated for parenteral or oral administration to humans or other mammals in therapeutically effective amounts, e.g., amounts which provide appropriate concentrations of the analog to target tissue for a time sufficient to induce the desired effect.

Where the analog is to be used as part of a transplant procedure, it can be provided to the living tissue or organ to be transplanted prior to removal of tissue or organ from the donor. The analog may be provided to the donor host directly, as by injection of a formulation comprising the analog into the tissue, or indirectly, e.g., by oral or parenteral administration, using any of the means described above.

Alternatively or, in addition, once removed from the donor, the organ or living tissue can be placed in a preservation solution containing the therapeutic molecule. In addition, the recipient also preferably is provided with the analog just prior to, or concommitant with, transplantation. In all cases, the analog can be administered directly to the tissue at risk, as by injection to the tissue, or it may be provided systemically, either by oral or parenteral administration, using any of the methods and formulations described herein and/or known in the art.

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Where the therapeutic molecule comprises part of a tissue or organ preservation solution, any commercially available preservation solution can be used to advantage. For example, useful solutions known in the art include Collins solution, Wisconsin solution, Belzer solution, Eurocollins solution and lactated Ringer's solution. Generally, an organ preservation solution usually possesses one or more of the following properties: (a) an osmotic pressure substantially equal to that of the inside of a mammalian cell, (solutions typically are hyperosmolar and have K+ and/or Mg++ ions present in an amount sufficient to produce an osmotic pressure slightly higher than the inside of a mammalian cell); (b) the solution typically is capable of maintaining substantially normal ATP levels in the cells; and (c) the solution usually allows optimum maintenance of glucose metabolism in the cells. Organ

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preservation solutions also may contain anticoagulants, energy sources such as glucose, fructose and other sugars, metabolites, heavy metal chelators, glycerol and other materials of high viscosity to enhance survival at low temperatures, free oxygen radical inhibiting and/or scavenging agents and a pH indicator. A detailed description of preservation solutions and useful components can be found, for example, in US Patent No. 5,002,965, the disclosure of which is incorporated herein by reference.

As will be appreciated by those skilled in the art, the concentration of the compounds described in a therapeutic composition will vary depending upon a number of factors, including the dosage of the drug to be administered, the chemical characteristics (e.g., hydrophobicity) of the compounds employed, and the route of administration. The preferred dosage of drug to be administered also is likely to depend on such variables as the type and extent of tissue loss or defect, the overall health status of the particular patient, the relative biological efficacy of the compound selected, the formulation of the compound, the presence and types of excipients in the formulation, and the route of administration. In general terms, the therapeutic molecules of this invention may be provided to and individual where typical dose ranges are from about 10 ng/kg to about 1 g/kg of body weight per day; a preferred dose range being from about 0.1 mg/kg to 100 mg/kg of body weight. No obvious morphogeninduced pathological lesions are induced when mature morphogen (e.g., OP-1, 20 mg) is administered daily to normal growing rats for 21 consecutive days. Moreover, 10 mg systemic injections of morphogen (e.g., OP-1) injected daily for 10 days into normal newborn mice does not produce any gross abnormalities.

### Other Embodiments

The invention may be embodied in other specific forms without departing from the spirit or essential characteristics thereof. The present embodiments are therefore to be considered in all respects as illustrative and not restrictive, the scope of the invention being indicated by the appended claims rather than by the foregoing description, and all changes which come within the meaning and range of equivalency of the claims are therefore intended to be embraced therein.

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#### SEQUENCE LISTING

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(1) GENERAL INFORMATION:
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            (i) APPLICANT:
                (B) NAME: CREATIVE BIOMOLCULES, INC
                (C) STREET: 45 SOUTH STREET (D) STATE: HOPKINTON
10
                (E) COUNTRY: USA
                (F) POSTAL CODE (ZIP): 01748
(G) TELEPHONE: (508)-435-9001
                (H) TELEFAX: (508)-435-0992
                (I) TELEX:
15
            (i) APPLICANT:
                (B) NAME: LUDWIG INSTITUTE FOR CANCER RESEARCH
                (C) STREET: 1345 AVENUE OF THE AMERICAS
                (D) STATE: NY
20
                (E) COUNTRY: USA
                (F) POSTAL CODE (ZIP): 10105
(G) TELEPHONE: (212)-765-3000
                (H) TELEFAX:
                (I) TELEX:
25
          (ii) TITLE OF INVENTION: MORPHOGENIC PROTEIN-SPECIFIC CELL
                   SURFACE RECEPTORS AND USES THEREFOR
         (iii) NUMBER OF SEQUENCES: 15
30
          (iv) CORRESPONDENCE ADDRESS:
                 (A) ADDRESSEE: PATENT ADMINISTRATOR, TESTA, HURWITZ &
                        THIBEAULT
                 (B) STREET: 53 STATE STREET
35
                 (C) CITY: BOSTON
                 (D) STATE: MA
               (E) COUNTRY: USA
                 (F) ZIP: 02109
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           (v) COMPUTER READABLE FORM:
                 (A) MEDIUM TYPE: Floppy disk
                 (B) COMPUTER: IBM PC compatible (C) OPERATING SYSTEM: PC-DOS/MS-DOS
                 (D) SOFTWARE: PatentIn Release #1.0, Version #1.25
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          (vi) CURRENT APPLICATION DATA:
                 (A) APPLICATION NUMBER:
                 (B) FILING DATE:
                 (C) CLASSIFICATION:
50
        (viii) ATTORNEY/AGENT INFORMATION:
                 (A) NAME: PITCHER, EDMUND R.
                 (B) REGISTRATION NUMBER: 27,829
                (C) REFERENCE/DOCKET NUMBER: CRP-097PC
55
          (ix) TELECOMMUNICATION INFORMATION:
                (A) TELEPHONE: (617) 248-7000
                 (B) TELEFAX: (617) 248-7100
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(2) INFORMATION FOR SEQ ID NO:1:

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10				AGG Arg													624
15				CGC Arg													672
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25				ATC Ile 260													816
30				ATC Ile													864
35	CTG Leu	CAG Gln 290	AGA Arg	CAG Gln	ACG Thr	CTG Leu	GAG Glu 295	CCC Pro	CAT His	CTG Leu	GCT Ala	CTG Leu. 300	AGG Arg	CTA Leu	GCT Ala	GTG Val	912
				TGC <b>C</b> ys													960
40				CCA Pro													1008
45				AGC Ser 340						_							1056
50				TCA Ser													1104
55				ACC Thr													1152
33				GAC Asp													1200
60	TTT	GGC	CTG	GTG	CTG	TGG	GAG	TTA	GCC	CGC	CGG	ACC	ATC	GTG	<b>AA</b> T	GGC	1248

	Phe	Gly	Leu	Val	Leu 405	Trp	Glu	Ile	Ala	Arg 410	Arg	Thr	Ile	Val	Asn 415	Gly	
5	ATC Ile	GTG Val	GAG Glu	GAC Asp 420	TAT Tyr	AGA Arg	CCA Pro	CCC Pro	TTC Phe 425	TAT Tyr	GAT Asp	GTG Val	GTG Val	CCC Pro 430	AAT Asn	GAC Asp	1296
10	CCC Pro	AGC Ser	TTT Phe 435	GAG Glu	GAC Asp	ATG Met	AAG Lys	AAG Lys 440	GTG Val	GTG Val	TGT Cys	GTG Val	GAT Asp 445	CAG Gln	CAG Gln	ACC Thr	1344
15	CCC Pro	ACC Thr 450	ATC Ile	CCT Pro	AAC Asn	CGG Arg	CTG Leu 455	GCT Ala	GCA Ala	GAC Asp	CCG Pro	GTC Val 460	CTC Leu	TCA Ser	GGC Gly	CTA Leu	1392
15	GCT Ala 465	CAG Gln	ATG Met	ATG Met	CGG Arg	GAG Glu 470	TGC Cys	TGG Trp	TAC Tyr	CCA Pro	AAC Asn 475	CCC Pro	TCT Ser	GCC Ala	CGA Arg	CTC Leu 480	1440
20				CGG Arg													1488
25				AAA Lys 500													1509
30	(2)			rion Seque	ENCE	CHAF	RACTI	ERIST	rics:								
35				(B)	TY	NGTH: PE: & POLOG	mino	aci	id	cids	5						
				OLEC SEQUE			_			) ID	NO: 2	2:					
40	Met 1			Gly						-			Leu	Leu	Met 15	Ala	
	Leu	Val	Thr	Gln 20	Gly	Asp	Pro	Val	Lys 25	Pro	Ser	Arg	Gly	Pro 30	Leu	Val	
45	Thṛ	Cys	Thr 35	Суѕ	Glu	Ser	Pro	His 40	Cys	Lys	Gly	Pro	Thr 45	Cys	Àrg	Gly	
50	Ala	Trp 50	Cys	Thr	Val	Val	Leu 55	Val	Arg	Glu	Glu	Gly 60	Arg	His	Pro	Gln	
	Glu 65	His	Arg	Gly	Cys	Gly 70	Asn	Leu	His	Arg	Glu 75	Leu	Cys	Arg	Gly	Arg 80	
55	Pro	Thr	Glu	Phe	Val 85	Asn	His	Tyr	Cys	Cys 90	Asp	Ser	His	Leu	Cys 95	Asn	
	His	Asn	Val	Ser 100	Leu	Val	Leu	Glu	Ala 105	Thr	Gln	Pro	Pro	Ser 110	Glu	Gln	

	Pro	Gly	Thr 115	Asp	Gly	Gln	Leu	Ala 120	Leu	Ile	Leu	Gly	Pro 125	Val	Leu	Ala
5	Leu	Leu 130	Ala	Leu	Val	Ala	Leu 135	Gly	Val	Leu	Gly	Leu 140	Trp	His	Val	Arg
	Arg 145	Arg	Gln	Glu	Lys	Gln 150	Arg	Gly	Leu	His	Ser 155	Glu	Leu	Gly	Glu	Se:
10	Ser	Leu	Ile	Leu	Lys 165	Ala	Ser	Glu	Gln	Gly 170	Asp	Thr	Met	Leu	Gly 175	Ası
15	Leu	Leu	Asp	Ser 180	Asp	Cys	Thr	Thr	Gly 185	Ser	Gly	Ser	Gly	Leu 190	Pro	Phe
	Leu	Val	Gln 195	Arg	Thr	Val	Ala	Arg 200	Gln	Val	Ala	Leu	Val 205	Glu	Cys	Va]
20	Gly	Lys 210	Gly	Arg	Tyr	Gly	Glu 215	Val	Trp	Arg	Gly	Leu 220	Trp	His	Gly	Glı
	Ser 225	Val	Ala	Val	Lys	Ile 230	Phe	Ser	Ser	Arg	Asp 235	Glu	Gln	Ser	Trp	Phe 240
25	Arg	Glu	Thr	Glu	11e 245	Tyr	Asn	Thr	Val	Leu 250	Leu	Arg	His	Asp	Asn 255	Ile
30	Leu	Gly	Phe	11e 260	Ala	Ser	Asp	Met	Thr 265	Ser	Arg	Asn	Ser	Ser 270	Thr	Glr
	Leu	Trp	Leu 275	Ile	Thr	His	Tyr	His 280	Glu	His	Gly	Ser	Leu 285	Tyr	Asp	Phe
<b>3</b> 5	Leu	Gln 290	Arg	Gln	Thr	Leu	Glu 295	Pro	His	Leu	Ala	Leu 300	Arg	Leu	Ala	Val
	Ser 305	Ala	Ala	Cys	Gly	Leu 310	Ala	His	Leu	His	Val 315	Glu	Ile	Phe	Gly	Thr 320
40	Gln	Gly	Lys '	Pro	Ala 325	Ile	Ala	His	Arg	Asp 330	Phe	Lys	Ser	Arg	Asn 335	Vai
45	Leu	Val	Lys	Ser 340	Asn	Leu	Gln.	Cys	Cys 345	Ile	Ala	Asp	Leu	Gly 350	Leu	Ala
	Val	Met	His 355	Ser	Gln	Gly	Ser	Asp 360	Tyr	Leu	Asp	Ile	Gly 365	Asn	Asn	Pro
50	Arg	Val 370	Gly	Thr	Lys	Arg	Туr 375	Met	Ala	Pro	Glu	Val 380	Leu	Asp	Glu	Glr
	Ile 385	Arg	Thr	Asp	Суѕ	Phe 390	Glu	Ser	Ţyr	Lys	Trp 395	Thr	qsA	Ile	Trp	Ala 400
55	Phe	Gly	Leu	Val	Leu 405	Trp	Glu	Ile	Ala	Arg 410	Arg	Thr	Ile	Val	Asn 415	Gly
<b>.</b> n	Ile	Val	Glu	Asp 420	Tyr	Arg	Pro	Pro	Phe 425	Tyr	Asp	Val	Val	Pro 430	Asn	Asp

	Pro	Ser	Phe 435	Glu	Asp	Met	Lys	Lys 440	Val	Val	Cys	Val	Asp 445	Gln	Gln	Thr	
5	Pro	Thr 450	Ile	Pro	Asn	Arg	Leu 455	Ala	Ala	Asp	Pro	Val 460	Leu	Ser	Gly	Leu	
	Ala 465	Gln	Met	Met	Arg	Glu 470	Cys	Trp	Tyr	Pro	Asn 475	Pro	Ser	Ala	Arg	Leu 480	
10	Thr	Ala	Leu	Arg	Ile 485	Lys	Lys	Thr	Leu	Gln 490	Lys	Ile	Ser	Asn	Ser 495	Pro	
15	Glu	Lys	Pro	Lys 500	Val	Ile	Gln										
	(2)	INFO	ORMA	rion	FOR	SEQ	ID N	10 : 3 :	:			•					
20		(i)	() () ()	QUENC A) LI B) TY C) SY O) TO	ENGTI (PE : [RANI	i: 27	724 k Leic ESS:	ase acio sino	pai:	rs							
25		(ii)	MOI	LECUI	LE TY	PE:	cDN2	A									
30		(ix)	() ()	ATURI A) NZ B) LO D) O'	AME/I	ION:	104			rodu	ct= '	"Huma	an Al	∟K2"			
35	CTC	•		QUEN(								CGA	GGC	ACG (	cggc'	PTGAAG	60
40	GAC	TGTG	GGC 2	AGAT	GTGA(	CC A	AGAG	CTG	C AT	TAAG'	rtgt	ACA			GAT Asp		115
40	GTG Val	ATG Met	ATT Ile	CTT Leu	CCT Pro	GTG Val 10	CTT Leu	ATC Ile	ATG Met	ATT Ile	GCT Ala 15	CTC Leu	CCC	TCC Ser	CCT Pro	AGT Ser 20	163
45	ATG	GAA Glu	GAT Asp	GAG Glu	AAG Lys 25	CCC	AAG Lys	GTC Val	AAC Asn	CCC Pro 30	AAA	CTC Leu	TAC Tyr	ATG Met	TGT Cys 35	GTG	211
50	TGT Cys	GAA Glu	GGT Gly	CTC Leu 40	TCC Ser	TGC Cys	GGT Gly	AAT Asn	GAG Glu 45	GAC Asp	CAC His	TGT Cys	GAA Glu	GGC Gly 50	Gln	CAG Gln	259
55	TGC Cys	TTT Phe	TCC Ser 55	Ser	CTG Leu	AGC Ser	ATC Ile	AAC Asn 60	Asp	GGC Gly	TTC Phe	CAC His	GTC Val 65	TAC Tyr	CAG Gln	AAA Lys	307
60	GGC Gly	TGC Cys 70	Phe	CAG Gln	GTT Val	TAT Tyr	GAG Glu 75	Gln	GGA Gly	AAG Lys	ATG Met	ACC Thr 80	TGT Cys	AAG Lys	ACC Thr	CCG Pro	355

-	TCC Ser								403
5	AAC Asn								451
.10	CAG Gln								499
.15	GCA Ala								547
20	AAA Lys 150								595
25	ACT Thr								643
23	GAT Asp								691
30	TTT Phe								739
35	GTC Val								787
40	GAA Glu 230								835
45	TTC Phe								883
. <b></b>	ATC Ile								931
50	CAG Gln							Leu	979
55	TAT Tyr								1027
60	CTG Leu 310								1075

5			CAA Gln														1123
J			CTG Leu														1171
10			GTC Val														1219
15			CGT Arg 375														1267
20			ATC Ile														1315
25			TTT Phe														1363
23		_	ATA Ile	_													1411
30			CCA Pro														1459
35			CCA Pro 455														1507
40			GCC Ala														1555
45			ACA Thr														1603
45			GAC Asp							TGA0	CATT	TTC 2	ATAG	rgtc2	AA.		1650
50	GAAC	GAAG	AT 1	rtgac	CGTTC	T TO	TCAT	rtgto	CAC	CTG	GAC	CTA	ATGC	rgg (	CTG	CTGGT	1710
	TGT	CAGAZ	ATG (	CAAT	CATO	T G	CTC	CTC	CCZ	YTAAA	GCT	GCT	TGAC	CAA (	GCAC	GACGTC	1770
55	GTAC	CCCAC	GCC [2	ATGTO	TTG	GG GZ	AGACA	ATCAZ	A AAC	CAC	CTA	ACC	rcgc	rcg 2	ATGAC	TGTGA	1830
	ACTO	3GGC2	ATT .	CAC	SAAC	rg Ti	CAC	ACTGO	AGA	AGAC	TAAT	GTT	GAC	AGA (	CACTO	STTGCA	1890
	AAGO	GTAG(	GA (	CTGG	AGGAZ	AC AC	CAGAC	SAAA:	r cc:	<b>LAAA</b> 1	AGAG	ATC	rggg	CAT 7	raagt	CAGTG	1950
60	GCT7	rtgcz	ATA: C	GCTT:	CAC	AA G	CTC	CTAG	A CAC	CTCC	CAC	GGGZ	AAAC:	rca A	AGGAC	GTGGT	2010

	GAA	TTTT	AAT	TCAG	CAAT	AT T	GCCT	GTGC	T TC	TCTT	CTTT	ATT	GCAC	TAG	GAAT	TCTTTG
5	CAT	TCCT	TAC	TTGC	ACTG	TT A	CTCT	TAAT	т тт	AAAG	ACCC	AAC	TTGC	CAA	AATG	TTGGCT
J	GCG	TACT	CCA	CTGG	TCTG	тс т	TTGG	ATAA	T AG	GAAT	TCAA	TTT	GGCA	AAA	CAAA	ATGTAA
	TGT	CAGA	CTT	TGCT	GCAT	тт т	ACAC	ATGT	G CT	GATG	TTTA	CAA	TGAT	GCC	GAAC	ATTAGG
10	AAT	TGTT	TAT	ACAC	AACT	TT G	CAAA	TTAT'	т та	TTAC	TTGT	GCA	CTTA	GTA	GTTT	TTACAA
	AAC	TGCT	TTG	TGCA	TATG	TT A	AAGC	TTAT'	т тт	TATG	TGGT	CTT	ATGA	TTT	TATT	ACAGAA
15	ATG	TTTT	TAA	CACT	ATAC	тс т	AAAA	TGGA	C AT	TTTC	TTTT	ATT.	ATCA	GTT .	AAAA	TCACAT
13	TTT	AAGT	GCT	TCAC	ATTT	GT A	TGTG	TGTA	g ac	TGTA	ACTT	TTT	TTCA	GTT	CATA	TGCAGA
	ACG	TATT	TAG	CCAT	TACC	CA C	GTGA	CACC	A CC	GAAT	ATAT	TAT	CGAT	TŢA	GAAG	CAAAGA
20	TTT	CAGT.	AGA .	ATTT	TAGT	CCŢ	GAAC	GCTA	c <b>G</b> G	GGAA	AATG	CAT	TTTC	TTC .	AGAA	TTATCC
	ATT.	ACGT	GCA	TTTA	AACT	CT G	CCAG	AAAA	A AA	TAAC	TATT	TTG	TTTT.	AAT	CTAC	TTTTTG
25	TAT	TTAG	TAG	TTAT	TTGT.	AT A	AATT	AAAT	AA A	CTGT	TTTC	AAG	TCAA	AAA .	AAAA	
	(2)	INF	ORMA	TION	FOR	SEO	ו מז	NO - 4								
	(-,			SEQU:												
30				(A	) LE	NGTH	: 50	9 am:	ino a		s					
								linea								
35		(:	ii)	MOLE	CULE	TYP	E: p:	rote:	in							
				SEQU						•						
40	Met 1	Val	Asp	Gly	Val 5	Met	Ile	Leu	Pro	Val 10	Leu	Ile	Met	Ile	Ala 15	Leu
	Pro	Ser	Pro	Ser 20	Met	Glu	Asp	Glu	Lys 25	Pro	Lys	Val	Asn	Pro 30	Lys	Leu
45	Tyr	Met	Cys 35	Val	Cys	Glu	Gly	Leu 40	Ser	Cys	Gly	Asn	Glu 45	Asp	His	Cys
	Glu	Gly 50	Gln	Gln	Cys	Phe	Ser 55	Ser	Leu	Ser	Ile	Asn 60	Asp	Gly	Phe	His
50		Tyr	Gln	Lys	Gly		Phe	Gln	Val	Tyr		Gln	Gly	Lys	Met	
	65					70					75					80
	Cys	Lys	Thr	Pro	Pro 85	Ser	Pro	Gly	Gln	Ala 90	Val	Glu	Cys	Cys	Gln 95	Gly
55	Asp	Trp	Cys	Asn 100	Arg	Asn	Ile	Thr	Ala 105	Gln	Leu	Pro	Thr	Lys 110	Gly	Lys
60	Ser	Phe	Pro 115	Gly	Thr	Gln	Asn	Phe 120	His	Leu	Glu	Val	Gly 125	Leu	Ile	Ile

	Leu	Ser 130	Val	Val	Phe	Ala	Val 135	Cys	Leu	Leu	Ala	Cys 140	Leu	Leu	Gly	Val
5	Ala 145	Leu	Arg	Lys	Phe	Lys 150	Arg	Arg	Asn	Gln	Glu 155	Arg	Leu	Asn	Pro	Arg 160
	Asp	Val	Glu	Tyr	Gly 165	Thr	Ile	Glu	Gly	Leu 170	Ile	Thr	Thr	Asn	Val 175	Gly
10	qaA	Ser	Thr	Leu 180	Ala	Asp	Leu	Leu	Asp 185	His	Ser	Cys	Thr	Ser 190	Gly	Ser
15	Gly	Ser	Gly 195	Leu	Pro	Phe	Leų	Val 200	Gln	Arg	Thr	Val	Ala 205	Arg	Gln	Ile
	Thr	Leu 210	Leu	Glu	Cys	Val	Gly 215		Gly	Arg	Tyr	Gly 220	Glu	Val	Trp	Arg
20	Gly 225	Ser	Trp	Gln	Gly	Glu 230	Asn	Val	Ala	Val	Lys 235	Ile	Phe	Ser	Ser	Arg 240
25	Asp	Glu	Lys	Ser	Trp 245	Phe	Arg	Glu	Thr	Glu 250	Leu	Tyr	Asn	Thr	Val 255	Met
23	Leu	Arg	His	Glu 260	Asn	Ile	Leu	Gly	Phe 265	Ile	Ala	Ser	Asp	Met 270	Thr	Ser
30	Arg	His	Ser 275	Ser	Thr	Gln	Leu	Trp 280	Leu	Ile	Thr	His	Tyr 285	His	Glu	Met
	Gly	Ser 290	Leu	Tyr	Asp	Tyr	Leu 295	Gln	Leu	Thr	Thr	Leu 300	Asp	Thr	Val	Ser
35	Cys 305	Leu	Arg	Ile	Val	Leu 310	Ser	Ile	Ala	Ser	Gly 315	Leu	Ala	His	Leu	His 320
40	Ile	Glu	Ile	Phe	Gly 325	Thr	Gln	Gly	Lys	Pro 330	Ala	Ile	Ala	His	Arg 335	Asp
40	Leu	Lys	Ser	Lys 340	Asn	Ile	Leu	Val	Lys 345	Lys	Asn	Gly	Gln	Суs 350	Cys	Ile
45	Ala	Asp	Leu 355	Gly	Leu	Ala	Val	Met 360	His	Ser	Gln	Ser	Thr 365	Asn	Gln	Leu
	Asp	Val 370	Gly	Asn	Asn	Pro	Arg 375	Val	Gly	Thr	Lys	Arg 380	Tyr	Met	Ala	Pro
50	Glu 385	Val	Leu	Asp	Glu	Thr 390	Ile	Gln	Val	Asp	Cys 395	Phe	Asp	Ser	Tyr	Lys 400
	Arg	Val	Asp	Ile	Trp 405		Phe	Gly	Leu	Val 410	Leu	Trp	Glu	Val	Ala 415	Arg
55	Arg	Met	<b>V</b> al	Ser 420	Asn	Gly	Ile	Val	Glu 425	Asp	Tyr	Lys	Pro	Pro 430		Tyr
60	Asp	Val	Val 435	Pro	Asn	Asp	Pro	Ser 440	Phe	Glu	Asp	Met	Arg 445	Lys	Val	Val

	450 455 460	
5	Pro Thr Leu Thr Ser Leu Ala Lys Leu Met Lys Glu Cys Trp Tyr Gln 465 470 480	
	Asn Pro Ser Ala Arg Leu Thr Ala Leu Arg Ile Lys Lys Thr Leu Thr 485 490 495	
10	Lys Ile Asp Asn Ser Leu Asp Lys Leu Lys Thr Asp Cys 500 505	
15	(2) INFORMATION FOR SEQ ID NO:5:	
20	(i) SEQUENCE CHARACTERISTICS:  (A) LENGTH: 2932 base pairs  (B) TYPE: nucleic acid  (C) STRANDEDNESS: single  (D) TOPOLOGY: linear	
	(ii) MOLECULE TYPE: cDNA	
25 30	<pre>(ix) FEATURE:     (A) NAME/KEY: CDS     (B) LOCATION: 3101908     (D) OTHER INFORMATION: /product= "Human ALK3"</pre>	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:	
35	GCTCCGCGCC GAGGGCTGGA GGATGCGTTC CCTGGGGTCC GGACTTATGA AAATATGCAT	60
,,	CAGTTTAATA CTGTCTTGGA ATTCATGAGA TGGAAGCATA GGTCAAAGCT GTTTGGAGAA	120
	AATCAGAAGT ACAGTTTTAT CTAGCCACAT CTTGGAGGAG TCGTAAGAAA GCAGTGGGAG	180
10	TTGAAGTCAT TGTCAAGTGC TTGCGATCTT TTACAAGAAA ATCTCACTGA ATGATAGTCA	240
	TTTAAATTGG TGAAGTAGCA AGACCAATTA TTAAAGGTGA CAGTACACAG GAAACATTAC	300
15	AATTGAACA ATG ACT CAG CTA TAC ATT TAC ATC AGA TTA TTG GGA GCC Met Thr Gln Leu Tyr Ile Tyr Ile Arg Leu Leu Gly Ala 1 5 10	348
50	TAT TTG TTC ATC ATT TCT CGT GTT CAA GGA CAG AAT CTG GAT AGT ATG Tyr Leu Phe Ile Ile Ser Arg Val Gln Gly Gln Asn Leu Asp Ser Met 15 20 25	396
55	CTT CAT GGC ACT GGG ATG AAA TCA GAC TCC GAC CAG AAA AAG TCA GAA Leu His Gly Thr Gly Met Lys Ser Asp Ser Asp Gln Lys Lys Ser Glu 30 35 40 45	444
	AAT GGA GTA ACC TTA GCA CCA GAG GAT ACC TTG CCT TTT TTA AAG TGC Asn Gly Val Thr Leu Ala Pro Glu Asp Thr Leu Pro Phe Leu Lys Cys 50 55 60	492
in.	יים ארך ארם האר האר מוכר לא האר האר ארם האר האר ארם ארם האר ארם ארם ארם ארם ארם ארם ארם ארם ארם א	540

	Tyr	Cys	Ser	Gly 65	His	Cys	Pro	Asp	Asp 70	Ala	Ile	Asn	Asn	Thr 75	Суѕ	Ile		
5		AAT Asn															588	3
10		ACA Thr 95															636	5
15		AAA Lys															684	į
		ACC Thr					_										732	!
20		ATA Ile															780	)
25	_	TCT Ser		_													828	}
30	_	TGT Cys 175			_					_						TAC Tyr	876	j
35		CGT Arg															924	Ĺ
33		AAA Lys															972	<u> </u>
40		CCT Pro														GTC ' Val	1020	)
45		CAA Gln															1068	}
50		GGC Gly 255	Glu	Lys	Val		Val	Lys	Val	Phe	Phe	Thr	Thr				1116	;
55		TGG Trp															1164	Ī
		AAC Asn															1212	!
60	TGG	ACT	CAG	CTC	TAT	TTG	ATT	ACT	GAT	TAC	CAT	GAA	AAT	GGA	TCT	CTC ·	1260	)

	Trp	Thr	Gln	Leu 305	Tyr	Leu	Ile	Thr	Asp 310	Tyr	His	Glu	Asn	Gly 315	Ser	Leu	
5	TAT Tyr	GAC Asp	TTC Phe 320	CTG Leu	AAA Lys	TGT Cys	GCT Ala	ACA Thr 325	CTG Leu	GAC Asp	ACC Thr	AGA Arg	GCC Ala 330	CTG Leu	CTT Leu	AAA Lys	1308
10			TAT Tyr														1356
15	TAT Tyr 350	GGC Gly	ACC Thr	CAA Gln	GGA Gly	AAG Lys 355	CCC Pro	GCA Ala	ATT Ile	GCT Ala	CAT His 360	CGA Arg	GAC Asp	CTA Leu	AAG Lys	AGC Ser 365	1404
15	AAA Lys	AAC Asn	ATC Ile	CTC Leu	ATC Ile 370	AAG Lys	AAA Lys	AAT Asn	GGG Gly	AGT Ser 375	TGC Cys	TGC Cys	ATT Ile	GCT Ala	GAC Asp 380	CTG Leu	1452
20			GCT Ala														1500
25	TTG Leu	AAT Asn	ACC Thr 400	AGG Arg	GTG Val	GGC Gly	ACC Thr	AAA Lys 405	CGC Arg	TAC Tyr	ATG Met	GCT Ala	CCC Pro 410	GAA Glu	GTG Val	CTG Leu	1548
30			AGC Ser														1596
35	ATC Ile 430	TAC Tyr	AGC Ser	TTC Phe	GGC	CTA Leu 435	ATC Ile	ATT Ile	TGG Trp	GAG Glu	ATG Met 440	GCT Ala	CGT Arg	CGT Arg	TGT Cys	ATC Ile 445	1644
			GGG Gly														1692
40			GAT Asp														1740
45			CGG Arg 480														1788
50	Arg	GCA Ala 495	GTT Val	TTG Leu	Lys	CTA Leu	Met	Ser	GAA Glu	TGC <b>C</b> ys	Trp	GCC Ala 505	CAC His	AAT Asn	CCA Pro	GCC Ala	1836
55			CTC Leu														1884
			CAA Gln					TGAT	'GGT'I	'AA A	CCAT	CGGA	G GA	GAAA	CTCT	•	1935
60	AGAC	TGC	AG A	ACTO	TTTT	T AC	CCAT	GGCA	TGG	GTGG	TAA	TAGA	GTGG	AA I	'AAGG	ATGTT	1995

60

	AACTTGGTTC TCAGACTCTT TCTTCACTAC GTGTTCACAG GCTGCTAATA TTAAACCTTT	2055
5	CAGTACTCTT ATTAGGATAC AAGCTGGGAA CTTCTAAACA CTTCATTCTT TATATATGGA	2115
J	CAGCTTTATT TTAAATGTGG TTTTTGATGC CTTTTTTTAA GTGGGTTTTT ATGAACTGCA	2175
	TCAAGACTTC AATCCTGATT AGTGTCTCCA GTCAAGCTCT GGGTACTGAA TTGCCTGTTC	2235
10	ATAAAACGGT GCTTTCTGTG AAAGCCTTAA GAAGATAAAT GAGCGCAGCA GAGATGGAGA	2295
	AATAGACTTT GCCTTTTACC TGAGACATTC AGTTCGTTTG TATTCTACCT TTGTAAAACA	2355
15	GCCTATAGAT GATGATGTGT TTGGGATACT GCTTATTTTA TGATAGTTTG TCCTGTGTCC	2415
	TTAGTGATGT GTGTGTCT CCATGCACAT GCACGCCGGG ATTCCTCTGC TGCCATTTGA	2475
	ATTAGAAGAA AATAATTTAT ATGCATGCAC AGGAAGATAT TGGTGGCCGG TGGTTTTGTG	2535
20	CTTTAAAAAT GCAATATCTG ACCAAGATTC GCCAATCTCA TACAAGCCAT TTACTTTGCA	2595
	AGTGAGATAG CTTCCCCACC AGCTTTATTT TTTAACATGA AAGCTGATGC CAAGGCCAAA	2655
25	AGAAGTTTAA AGCATCTGTA AATTTGGACT GTTTTCCTTC AACCACCATT TTTTTTGTGG	2715
	TTATTATTTT TGTCACGGAA AGCATCCTCT CCAAAGTTGG AGCTTCTATT GCCATGAACC	2775
	ATGCTTACAA AGAAAGCACT TCTTATTGAA GTGAATTCCT GCATTTGATA GCAATGTAAG	2835
. 30	TGCCTATAAC CATGTTCTAT ATTCTTTATT CTCAGTAACT TTTAAAAGGG AAGTTATTTA	2895
	TATTTTGTGT ATAATGTGCT TTATTTGCAA ATCACCC	2932
35	(2) INFORMATION FOR SEQ ID NO:6:	
	(i) SEQUENCE CHARACTERISTICS:	
	(A) LENGTH: 532 amino acids (B) TYPE: amino acid	
40	(D) TOPOLOGY: linear	
	(ii) MOLECULE TYPE: protein	
45	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:	
	Met Thr Gln Leu Tyr Ile Tyr Ile Arg Leu Leu Gly Ala Tyr Leu Phe 1 5 10 15	
50	Ile Ile Ser Arg Val Gln Gly Gln Asn Leu Asp Ser Met Leu His Gly 20 25 30	
	Thr Gly Met Lys Ser Asp Ser Asp Gln Lys Lys Ser Glu Asn Gly Val	
55	Thr Leu Ala Pro Glu Asp Thr Leu Pro Phe Leu Lys Cys Tyr Cys Ser 50 55 60	
	Gly His Cys Pro Asp Asp Ala Ile Asn Asn Thr Cys Ile Thr Asn Gly 65 70 75 80	

His Cys Phe Ala Ile Ile Glu Glu Asp Asp Gln Gly Glu Thr Thr Leu Ala Ser Gly Cys Met Lys Tyr Glu Gly Ser Asp Phe Gln Cys Lys Asp 100 105 1105 Ser Pro Lys Ala Gln Leu Arg Arg Thr Ile Glu Cys Cys Arg Thr Asn 115 120 125Leu Cys Asn Gln Tyr Leu Gln Pro Thr Leu Pro Pro Val Val Ile Gly 10 135 Pro Phe Phe Asp Gly Ser Ile Arg Trp Leu Val Leu Leu Ile Ser Met 145 150 155 160 15 Ala Val Cys Ile Ile Ala Met Ile Ile Phe Ser Ser Cys Phe Cys Tyr Lys His Tyr Cys Lys Ser Ile Ser Ser Arg Arg Arg Tyr Asn Arg Asp 20 Leu Glu Gln Asp Glu Ala Phe Ile Pro Val Gly Glu Ser Leu Lys Asp 200 Leu Ile Asp Gln Ser Gln Ser Ser Gly Ser Gly Ser Gly Leu Pro Leu 25 Leu Val Gln Arg Thr Ile Ala Lys Gln Ile Gln Met Val Arg Gln Val 30 Gly Lys Gly Arg Tyr Gly Glu Val Trp Met Gly Lys Trp Arg Gly Glu 245 250 255 Lys Val Ala Val Lys Val Phe Phe Thr Thr Glu Glu Ala Ser Trp Phe 35 Arg Glu Thr Glu Ile Tyr Gln Thr Val Leu Met Arg His Glu Asn Ile Leu Gly Phe Ile Ala Ala Asp Ile Lys Gly Thr Gly Ser Trp Thr Gln 40 Leu Tyr Leu Ile Thr Asp Tyr His Glu Asn Gly Ser Leu Tyr Asp Phe 45 Leu Lys Cys Ala Thr Leu Asp Thr Arg Ala Leu Leu Lys Leu Ala Tyr Ser Ala Ala Cys Gly Leu Cys His Leu His Thr Glu Ile Tyr Gly Thr 340 345 350 50 Gln Gly Lys Pro Ala Ile Ala His Arg Asp Leu Lys Ser Lys Asn Ile 355 360 365 Leu Ile Lys Lys Asn Gly Ser Cys Cys Ile Ala Asp Leu Gly Leu Ala Val Lys Phe Asn Ser Asp Thr Asn Glu Val Asp Val Pro Leu Asn Thr 60

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		Arg	Val	Gly	Thr	Lys 405	Arg	Tyr	Met	Ala	Pro 410	Glu	Val	Leu	Asp	Glu 415	Ser	
	5	Leu	Asn	Lys	Asn 420	His	Phe	Gln	Pro	туг 425	Ile	Met	Ala	Asp	Ile 430	туг	Ser	
		Phe	Gly	Leu 435	Ile	Ile	Trp	Glu	Met 440	Ala	Arg	Arg	Cys	Ile 445	Thr	Gly	Gly	
	10	Ile	Val 450	Glu	Glu	Tyr	Gln	Leu 455	Pro	Tyr	Tyr	Asn	Met 460	Val	Pro	Ser	Asp	
.:	15	Pro 465	Ser	Tyr	Glu	Asp	Met 470	Arg	Glu	Val	Val	Cys 475	Val	Lys	Arg	Leu	Arg 480	
		Pro	Ile	Val	Ser	Asn 485	Arg	Trp	Asn	Ser	Asp 490	Glu	Cys	Leu	Arg	Ala 495	Val	
	20	Leu	Lys	Leu	Met 500	Ser	Glu	Cys	Trp	Ala 505	His	Asn	Pro	Ala	Ser 510	Arg	Leu	
		Thr	Ala	Leu 515	Arg	Ile	Lys	Lys	Thr 520	Leu	Ala	Lys	Met	Val 525	Glu	Ser	Gln	
	25	Asp	<b>V</b> al 530	Lys	Ile												•	
•	30	(2)			QUENC	CE CH	IARAC	TERI	STIC	S: pair	rs ်							
	35		(ii)	(0	B) TY C) SI C) TO LECUI	POLO	DEDNE DGY :	ESS: line	sing ear									
•	40		(ix)	( E	ATURE A) NA B) LO	ME/F	ON:	187.			roduc	:t= '	'Muri	.ne F	ALK6'			
	45		(xi)	SEC	QUENC	E DE	ESCRI	PTIC	ON: S	SEQ 1	D NO	):7:						
		AAG	GGC	GGC 1	GAAC	TTGC	CC GC	CGT	GTG	TCC	TAG	GAG	GGCC	CGGA	AGG A	ACCCG	GGAC	C 60
	50	TGG	BAAGO	CGG (	GGCG	GGTT	'A A	CTTCC	GCTC	CAA S	CAC	AACC	ATTI	GGCG	CT C	AGCI	TATGAC	120
		AAGA	AGAGO	CAA A	CAA	AAGI	A T	AAGG?	AGCA	ccc	CGGCC	ATA	AGTO	AAGA	AGA C	AAGI	TTAT	r 180
-	55	GATZ		ATG ( Met I 1														228
.:	60	AAG Lys 15	GAG Glu	GAT Asp	GGA Gly	GAG Glu	AGT Ser 20	ACA Thr	GCC Ala	CCC Pro	ACC Thr	CCT Pro 25	CGG Arg	CCC Pro	AAG Lys	ATC Ile	CTA Leu 30	276

5	CGT Arg	TGT Cys	AAA Lys	TGC Cys	CAC His 35	CAC His	CAC His	TGT Cys	CCG Pro	GAA Glu 40	GAC Asp	TCA Ser	GTC Val	AAC Asn	AAT Asn 45	ATC Ile	324
J	TGC Cys	AGC Ser	ACA Thr	GAT Asp 50	GGG Gly	TAC Tyr	TGC Cys	TTC Phe	ACG Thr 55	ATG Met	ATA Ile	GAA Glu	GAA Glu	GAT Asp 60	GAC Asp	TCT Ser	372
10	GGA Gly	ATG Met	CCT Pro 65	GTT Val	GTC Val	ACC Thr	TCT Ser	GGA Gly 70	TGT Cys	CTA Leu	GGA Gly	CTA Leu	GAA Glu 75	GGG Gly	TCA Ser	GAT Asp	420
15	TTT Phe	CAA Gln 80	TGT Cys	CGT Arg	GAC Asp	ACT Thr	CCC Pro 85	ATT Ile	CCT Pro	CAT His	CAA Gln	AGA Arg 90	AGA Arg	TCA Ser	ATT Ile	GAA Glu	468
20	TGC Cys 95	TGC Cys	ACA Thr	GAA Glu	AGG Arg	AAT Asn 100	GAG Glu	TGT Cys	AAT Asn	AAA Lys	GAC Asp 105	CTC Leu	CAC His	CCC Pro	ACT Thr	CTG Leu 110	516
25		CCT Pro															564
	GCC Ala	TTG Leu	CTT Leu	ATC Ile 130	TCT Ser	GTG Val	ACT Thr	GTC Val	TGT Cys 135	AGT Ser	TTA Leu	CTC Leu	TTG Leu	GTC Val 140	CTC Leu	ATT Ile	612
30	ATT Ile	TTA Leu	TTC Phe 145	TGT Cys	TAC Tyr	TTC Phe	AGG Arg	ТАТ Туг 150	AAA Lys	AGA Arg	CAA Gln	GAA Glu	GCC Ala 155	CGA Arg	CCT Pro	CGG Arg	660
35	TAC Tyr	AGC Ser 160	ATT Ile	GGG Gly	CTG Leu	GAG Glu	CAG Gln 165	GAC Asp	GAG Glu	ACA Thr	TAC Tyr	ATT Ile 170	CCT Pro	CCT Pro	GGA Gly	GAG Glu	708
40		CTG Leu															756
45	GGC Gly	CTC Leu	CCT Pro	CTG Leu	CTG Leu 195	GTC Val	CAA Gln	AGG Arg	ACA Thr	ATA Ile 200	GCT Ala	AAG Lys	CAA Gln	ATT Ile	CAG Gln 205	ATG Met	804
		AAG Lys															852
50	TGG Trp	CGT Arg	GGA Gly 225	GAA Glu	AAG Lys	GTG Val	GCT Ala	GTG Val 230	AAA Lys	GTG Val	TTC Phe	TTC Phe	ACC Thr 235	ACG Thr	GAG Glu	GAA Glu	900
55		AGC Ser 240															948
60		GAG Glu															996

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5	TCC Ser	TGG Trp	ACT Thr	CAG Gln	TTG Leu 275	TAC Tyr	CTC Leu	ATC Ile	ACA Thr	GAC Asp 280	TAT Tyr	CAT His	GAA Glu	AAC Asn	GGC Gly 285	TCC Ser	1044
<b>.</b>	CTT Leu	TAT Tyr	GAC Asp	ТАТ Туг 290	CTG Leu	AAA Lys	TCC Ser	ACC Thr	ACC Thr 295	TTA Leu	GAC Asp	GCA Ala	AAG Lys	TCC Ser 300	ATG Met	CTG Leu	1092
10	AAG Lys	CTA Leu	GCC Ala 305	TAC Tyr	TCC Ser	TCT Ser	GTC Val	AGC Ser 310	GGC Gly	CTA Leu	TGC Cys	CAT His	TTA Leu 315	CAC His	ACG Thr	GAA Glu	1140
15	ATC Ile	TTT Phe 320	AGC Ser	ACT Thr	CAA Gln	GGC Gly	AAG Lys 325	CCA Pro	GCA Ala	ATC Ile	GCC Ala	CAT His 330	CGA Arg	GAC Asp	TTG Leu	AAA Lys	1188
20	AGT Ser 335	AAA Lys	AAC Asn	ATC Ile	CTG Leu	GTG Val 340	AAG Lys	AAA Lys	AAT Asn	GGA Gly	ACT Thr 345	TGC Cys	TGC Cys	ATA Ile	GCA Ala	GAC Asp 350	1236
25	CTG Leu	GGC Gly	TTG Leu	GCT Ala	GTC Val 355	AAG Lys	TTC Phe	ATT Ile	AGT Ser	GAC Asp 360	ACA Thr	AAT Asn	GAG Glu	GTT Val	GAC Asp 365	ATC Ile	1284
	CCA Pro	CCC Pro	AAC Asn	ACC Thr 370	CGG Arg	GTT Val	GGC Gly	ACC Thr	AAG Lys 375	CGC Arg	TAT Tyr	ATG Met	CCT Pro	CCA Pro 380	GAA Glu	GTG Val	1332
30	CTG Leu	GAC Asp	GAG Glu 385	AGC Ser	TTG Leu	AAT Asn	AGA Arg	AAC Asn 390	CAT His	TTC Phe	CAG Gln	TCC Ser	TAC Tyr 395	ATT Ile	ATG Met	GCT Ala	1380
35	GAC Asp	ATG Met 400	TAC Tyr	AGC Ser	TTT Phe	GGA Gly	CTC Leu 405	ATC Ile	CTC Leu	TGG Trp	GAG Glu	ATT Ile 410	GCA Ala	AGG Arg	AGA Arg	TGT Cys	1428
40	GTT Val 415	TCT Ser	GGA Gly	GGT Gly	ATA Ile	GTG Val 420	GAA Glu	GAA Glu	TAC Tyr	CAG Gln	CTT Leu 425	CCC Pro	TAT Tyr	CAC His	GAC Asp	CTG Leu 430	1476
45	GTG Val	CCC Pro	AGT Ser	GAC Asp	CCT Pro 435	TCT Ser	TAT Tyr	GAG Glu	GAC Asp	ATG Met 440	AGA Arg	GAA Glu	ATT Ile	GTG Val	TGC Cys 445	ATG Met	1524
						TCA Ser								-			1572
50	CTC Leu	AGG Arg	CAG Gln 465	ATG Met	GGG Gly	AAG Lys	CTT Leu	ATG Met 470	ACA Thr	GAG Glu	TGC Cys	TGG Trp	GCG Ala 475	CAG Gln	AAT Asn	CCT Pro	1620
55						GCC Ala											1668
60						ATT Ile 500			TGAC	GTCA	GA T	PACTI	GTGG	SA CA	GAGC	CAAGA	1722

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	TTA	TCAC	AGA	AGCA	TCGT	TA G	CCCA	AGCC	T TG	AACG	TTAG	CCT	ACTG	CCC	AGTG	AGTT	CA
5	GAC	TTTC	CTG	GAAG	AGAG	CA C	GGTG	GGCA	G AC	ACAG	AGGA	ACC	CAGA	AAC	ACGG	ATTC	T
,	CAT	GGCT	TTC	TGAG	GAGG	AG A	AACT	GTTT	G GG	TAAC	TTGT	TCA	AGAT	ATG	ATGC	ATGT'	rg
	CTT	TCTA	AGA	AAGC	CCTG	та т	TTTG	AATT.	A CC	ATTT	TTTT	ATA	AAAA	AAA			
10	(2)	INF	orma	TION	FOR	SEQ	ID :	NO : 8	:								
15			(i)	(B	) LE	NGTH PE:	RACT : 50 amin GY:	2 am	ino . id		s						<i>:</i>
		(	ii)	MOLE	CULE	TYP	E: p	rote	in								
20		(:	xi)	SEQU	ENCE	DES	CRIP	TION	: SE	Q ID	NO:	8:					
	Met 1	Leu	Leu	Arg	Ser 5	Ser	Gly	Lys	Leu	Asn 10	Val	Gly	Thr	Lys	Lys 15	Glu	
25	Asp	Gly	Glu	Ser 20	Thr	Ala	Pro	Thr	Pro 25	Arg	Pro	Lys	Ile	Leu 30	Arg	Cys	
30	Lys	Cys	His 35	His	His	Сув	Pro	Glu 40	Asp	Ser	Val	Asn	Asn 45	Ile	Суѕ	Ser	
	Thr	Asp 50	Gly	Tyr	Cys	Phe	Thr 55	Met	Ile	Glu	Glu	Asp 60	Asp	Ser	Gly	Met	
35	Pro 65	Val	Val	Thr	Ser	Gly 70	Суѕ	Leu	Gly	Leu	Glu 75	Gly	Ser	Asp	Phe	Gln 80	
	Сув	Arg	Asp	Thr	Pro 85	Ile	Pro	His	Gln	Arg 90	Arg	Ser	Ile	Glu	Суs 95	Cys	
40	Thr	Glu	Arg	Asn 100	Glu	Cys	Asn	Lys	Asp 105	Leu	His	Pro	Thr	Leu 110	Pro	Pro	•
45	Leu	Lys	Asp 115	Arg	Asp	Phe	Val	Asp 120	Gly	Pro	Ile	His	His 125	Lys	Ala	Leu	
••	Leu	Ile 130	Ser	Val	Thr	Val	Cys 135	Ser	Leu	Leu	Leu	Val 140	Leu	Ile	Ile	Leu	
50	Phe 145	Cys	Tyr	Phe	Arg	Tyr 150	Lys	Arg	Gln	Glu	Ala 155	Arg	Pro	Arg	Tyr	Ser 160	
	Ile	Gly	Leu	Glu	Gln 165	Asp	Glu	Thr	Tyr	Ile 170	Pro	Pro	Gly	Glu	Ser 175	Leu	
55	Arg	Asp	Leu	Ile 180	Glu	Gln	Ser	Gln	Ser 185	Ser	Gly	Ser	Gly	Ser 190	Gly	Leu	
50	Pro	Leu	Leu 195	Val	Gln	Arg	Thr	Ile 200	Ala	Lys	Gln	Ile	Gln 205	Met	Val	Lys	.:

- Gln Ile Gly Lys Gly Arg Tyr Gly Glu Val Trp Met Gly Lys Trp Arg 210 215 220 Gly Glu Lys Val Ala Val Lys Val Phe Phe Thr Thr Glu Glu Ala Ser 5 230 Trp Phe Arg Glu Thr Glu Ile Tyr Gln Thr Val Leu Met Arg His Glu Asn Ile Leu Gly Phe Ile Ala Ala Asp Ile Lys Gly Thr Gly Ser Trp 260 265 270 10 Thr Gln Leu Tyr Leu Ile Thr Asp Tyr His Glu Asn Gly Ser Leu Tyr 15 Asp Tyr Leu Lys Ser Thr Thr Leu Asp Ala Lys Ser Met Leu Lys Leu Ala Tyr Ser Ser Val Ser Gly Leu Cys His Leu His Thr Glu Ile Phe 20 Ser Thr Gln Gly Lys Pro Ala Ile Ala His Arg Asp Leu Lys Ser Lys Asn Ile Leu Val Lys Lys Asn Gly Thr Cys Cys Ile Ala Asp Leu Gly 25 Leu Ala Val Lys Phe Ile Ser Asp Thr Asn Glu Val Asp Ile Pro Pro 30 Asn Thr Arg Val Gly Thr Lys Arg Tyr Met Pro Pro Glu Val Leu Asp 370 380 Glu Ser Leu Asn Arg Asn His Phe Gln Ser Tyr Ile Met Ala Asp Met 35 390 Tyr Ser Phe Gly Leu Ile Leu Trp Glu Ile Ala Arg Arg Cys Val Ser Gly Gly Ile Val Glu Glu Tyr Gln Leu Pro Tyr His Asp Leu Val Pro 40 425 Ser Asp Pro Ser Tyr Glu Asp Met Arg Glu Ile Val Cys Met Lys Lys 440 Leu Arg Pro Ser Phe Pro Asn Arg Trp Ser Ser Asp Glu Cys Leu Arg 455 Gln Met Gly Lys Leu Met Thr Glu Cys Trp Ala Gln Asn Pro Ala Ser 50 Arg Leu Thr Ala Leu Arg Val Lys Lys Thr Leu Ala Lys Met Ser Glu 490 55 Ser Gln Asp Ile Lys Leu 500
  - (2) INFORMATION FOR SEQ ID NO:9:

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5		(i	(	A) L B) T C) S	CE C ENGT YPE: TRAN OPOL	H: 1 nuc DEDN	822 leic ESS:	base aci sin	pai d	rs							
		(ii	) MO	LECU	LE T	YPE:	cDN.	A									
10		(ix	(	B) L	AME/: OCAT: THER	ION:	49. ORMA		: /f	unct	ion=	"OS	TEOG	ENIC	PRO	TEIN"	
15								EXP. name			L						
		(xi	) SE	QUEN	CE D	ESCR:	IPTI	ON:	SEQ	ID N	0:9:						
20	GGT	GCGG	GCC (	CGGA	GCCC	GG A	GCCC	GGGT.	A GC	GCGT.	AGAG	CCG	GCGC	Me		C GTG s Val	57
25	CGC Arg	TCA Ser 5	CTG Leu	CGA Arg	GCT Ala	GCG Ala	GCG Ala 10	CCG Pro	CAC His	AGC Ser	TTC Phe	GTG Val 15	GCG Ala	CTC Leu	TGG Trp	GCA Ala	105
30	CCC Pro 20	CTG Leu	TTC Phe	CTG Leu	CTG Leu	CGC Arg 25	TCC Ser	GCC Ala	CTG Leu	GCC Ala	GAC Asp 30	TTC Phe	AGC Ser	CTG Leu	GAC Asp	AAC Asn 35	153
	GAG Glu	GTG Val	CAC His	TCG Ser	AGC Ser 40	TTC Phe	ATC Ile	CAC His	CGG Arg	CGC Arg 45	CTC Leu	CGC Arg	AGC Ser	CAG Gln	GAG Glu 50	CGG Arg	201
35	CGG Arg	GAG Glu	ATG Met	CAG Gln 55	CGC Arg	GAG Glu	ATC Ile	CTC Leu	TCC Ser 60	ATT Ile	TTG Leu	GGC Gly	TTG Leu	CCC Pro 65	CAC His	CGC Arg	249
40					CTC Leu												297
45	CTG Leu	GAC Asp 85	Le <i>u</i> CTG	TAC Tyr	AAC Asn	GCC Ala	ATG Met 90	GCG Ala	GTG Val	GAG Glu	GAG Glu	GGC Gly 95	GGC Gly	GGG Gly	CCC Pro	GGC Gly	345
50					TCC Ser												393
55 <sup>°</sup>					AGC Ser 120												441
<i></i>					TTC Phe												489
60	CAC	CCA	CGC	TAC	CAC	CAT	CGA	GAG	TTC	CGG	ттт	GAT	CTT	TCC	AAG	ATC	537

																	_
	His	Pro	Arg 150	Tyr	His	His	Arg	Glu 155	Phe	Arg	Phe	Asp	Leu 160	Ser	Lys	Ile	·
5			GGG Gly														585
10			CGG Arg														633
15			CTC Leu														681
15			CGT Arg														729
20			GCC Ala 230														777
25			CAG Gln								Gly						825
30			GCG Ala		Leu												873
25			GTG Val														921
35	CGG Arg	TCC Ser	ACG Thr	GGG Gly 295	AGC Ser	AAA Lys	CAG Gln	CGC Arg	AGC Ser 300	CAG Gln	AAC Asn	CGC Arg	TCC Ser	AAG Lys 305	ACG Thr	CCC Pro	.969
40			CAG Gln 310														1017
45	AGC Ser	GAC Asp 325	CAG Gln	AGG Arg	CAG Gln	Ala	TGT Cys 330	AAG Lys	AAG Lys	CAC His	GAG Glu	CTG Leu 335	TAT Tyr	GTC Val	AGC Ser	TTC Phe	1065
50	CGA Arg 340	GAC Asp	CTG Leu	GGC Gly	TGG Trp	CAG Gln 345	GAC Asp	TGG Trp	ATC Ile	ATC Ile	GCG Ala 350	CCT Pro	GAA Glu	GGC Gly	TAC Tyr	GCC Ala 355	1113
			TAC Tyr	Cys													1161
55			ACC Thr														1209
60	CCG	GAA	ACG	GTG	ccc	AAG	ccc	TGC	TGT	GCG	CCC	ACG	CAG	CTC	AAT	GCC	1257

	Pro Giu Thr Val Pro Lys Pro Cys Cys Ala Pro Thr Gln Leu Asn Ala 390 395 400	
5	ATC TCC GTC CTC TAC TTC GAT GAC AGC TCC AAC GTC ATC CTG AAG AAA Ile Ser Val Leu Tyr Phe Asp Asp Ser Ser Asn Val Ile Leu Lys Lys 405	1305
. 10	TAC AGA AAC ATG GTG GTC CGG GCC TGT GGC TGC CAC TAGCTCCTCC Tyr Arg Asn Met Val Val Arg Ala Cys Gly Cys His 420 425 430	1351
	GAGAATTCAG ACCCTTTGGG GCCAAGTTTT TCTGGATCCT CCATTGCTCG CCTTGGCCAG	1411
· 15	GAACCAGCAG ACCAACTGCC TTTTGTGAGA CCTTCCCCTC CCTATCCCCA ACTTTAAAGG	1471
13	TGTGAGAGTA TTAGGAAACA TGAGCAGCAT ATGGCTTTTG ATCAGTTTTT CAGTGGCAGC	1531.
	ATCCAATGAA CAAGATCCTA CAAGCTGTGC AGGCAAAACC TAGCAGGAAA AAAAAAACAAC	1591
20	GCATAAAGAA AAATGGCCGG GCCAGGTCAT TGGCTGGGAA GTCTCAGCCA TGCACGGACT	1651
•	CGTTTCCAGA GGTAATTATG AGCGCCTACC AGCCAGGCCA CCCAGCCGTG GGAGGAAGGG	1711
25	GGCGTGGCAA GGGGTGGGCA CATTGGTGTC TGTGCGAAAG GAAAATTGAC CCGGAAGTTC	1771
	CTGTAATAAA TGTCACAATA AAACGAATGA ATGAAAAAAA AAAAAAAAA A	1822
30	(2) INFORMATION FOR SEQ ID NO:10:  (i) SEQUENCE CHARACTERISTICS:  (A) LENGTH: 431 amino acids  (B) TYPE: amino acid  (D) TOPOLOGY: linear	
35	(ii) MOLECULE TYPE: protein	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:	
•40	Met His Val Arg Ser Leu Arg Ala Ala Ala Pro His Ser Phe Val Ala 1 5 10 15	
	Leu Trp Ala Pro Leu Phe Leu Leu Arg Ser Ala Leu Ala Asp Phe Ser	
45	Leu Asp Asn Glu Val His Ser Ser Phe Ile His Arg Arg Leu Arg Ser 35 40 45	
50	Gln Glu Arg Arg Glu Met Gln Arg Glu Ile Leu Ser Ile Leu Gly Leu 50 55 60	
	Pro His Arg Pro Arg Pro His Leu Gln Gly Lys His Asn Ser Ala Pro 65 70 75 80	
55	Met Phe Met Leu Asp Leu Tyr Asn Ala Met Ala Val Glu Glu Gly Gly 85 90 95	
60	Gly Pro Gly Gln Gly Phe Ser Tyr Pro Tyr Lys Ala Val Phe Ser 100 105 110	

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	Thr	Gln	Gly 115	Pro	Pro	Leu	Ala	Ser 120	Leu	Gln	Asp	Ser	His 125	Phe	Leu	Thr
5	Asp	Ala 130	Asp	Met	Val	Met	Ser 135	Phe	Val	Asn	Leu	Val 140	Glu	His	Asp	Lys
	Glu 145	Phe	Phe	His	Pro	Arg 150	Tyr	His	His	Arg	Glu 155	Phe	Arg	Phe	Asp	Leu 160
10	Ser	Lys	Ile	Pro	Glu 165	Gly	Glu	Ala	Val	Thr 170	Ala	Ala	Glu	Phe	Arg 175	Ile
15	Tyr	Lys	Asp	Tyr 180	Ile :	Arg	Glu	Arg	Phe 185	Asp	Asn	Glu	Thr	Phe 190	Arg	Ile
**	Ser	Val	Tyr 195	Gln	Val	Leu	Gln	Glu 200	His	Leu	Gly	Arg	Glu 205	Ser	qaA	Leu
20	Phe	Leu 210	Leu	Asp	Ser	Arg	Thr 215	Leu	Trp	Ala	Ser	Glu 220	Glu	Gly	Trp	Leu
	Val 225	Phe	Asp	Ile	Thr	Ala 230	Thr	Ser	Asn	His	Trp 235	Val	Val.	Asn	Pro	Arg 240
25	His	Asn	Leu	Gly	Leu 245	Gln	Leu	Ser	Val	Glu 250	Thr	Leu	qaA	Gly	Gln 255	Ser
30 -	Ile	Asn	Pro	Lys 260	Leu	Ala	Gly	Leu	Ile 265	Gly	Arg	His	Gly	Pro 270	Gln	Asn
JU .	Lys	Gln	Pro 275	Phe	Met	Val	Ala	Phe 280	Phe	Lys	Ala	Thr.	Glu 285	Val	His	Phe
35	Arg	Ser 290	Ile	Arg	Ser	Thr	Gly 295	Ser	Lys	Gln		Ser 300	Gln	Asn	Arg	Ser
	Lys 305	Thr	Pro	Lys	Asn	Gln 310	Glu	Ala	Leu	Arg	Met 315	Ala	Asn	Val	Ala	Glu 320
40	Asn	Ser	Ser	Ser	Asp 325	Gln	Arg	Gln	Ala	Cys 330	Lys	Lys	His	Glu	Leu 335	Tyr
45	Val	Ser	Phe	Arg 340	Asp	Leu	Gly	Trp	Gln 345	Asp	Trp	Ile	Ile	Ala 350	Pro	Glu
13	Gly	Tyr	Ala 355	Ala	Tyr	Tyr	Cys	Glu 360	Gly	Gĺu	Cys	Ala	Phe 365	Pro	Leu	Asn
50	Ser	Tyr 370	Met	Asn	Ala	Thr	Asn 375	His	Ala	Ile	Val	Gln 380	Thr	Leu	Val	His
	Phe 385	Ile	Asn	Pro	Glu	Thr 390	Val	Pro	Lys	Pro	Cys 395	Суs	Ala	Pro	Thr	Gln 400
<b>5</b> 5	Leu	Asn	Ala	Ile	Ser 405	Val	Leu	Tyr	Phe	Asp 410	Asp	Ser	Ser	Asn	Val 415	Ilė :
60	Leu	Lys	Lys	Tyr 420	Arg	Asn	Met	Val	Val 425	Arg	Ala	Cys	Gly	Cys 430	His	:

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	(2)	INFO	RMAT	ION	FOR	SEQ	ID N	0:11	:								
5		(i)	(A (B	) LE	E CH NGTH PE: POLO	: 10 amin	2 am o ac	ino id		s							
		(ii)	MOL	ECUL	E TY	PE:	prot	ein									
10		(ix)	(B	) NA ) LO	: ME/K CATI HER	ON:	11	02	/la	bel=	OPX						
15					a g	roup	of	one	a is or m he s	ore	spec	ifie	d am			from s	
20		(xi)	SEQ	UENC	E DE	SCRI	PTIO	N: S	EQ I	D NO	:11:						
		Cys 1	Xaa	Xaa	His	Glu 5	Leu	Tyr	Val	Xaa	Phe 10	Xaa	Asp	Leu	Gly	Trp 15	Xaa
25		Asp	Trp	Xaa	Ile 20	Ala	Pro	Xaa	Gly	Tyr 25	Xaa	Ala	Tyr	Tyr	Cys 30	Glu	Gly
30		Glu	Cys	Xaa 35	Phe	Pro	Leu	Xaa	Ser 40	Xaa	Met	Asn	Ala	Thr 45	Asn	His	Ala
		Ile	Xaa 50	Gln	Xaa	Leu	Val	His 55	Xaa	Xaa	Xaa	Pro	Xaa 60	Xaa	Val	Pro	Lys
35		Xaa 65	Cys	Cys	Ala	Pro	Thr 70	Xaa	Leu	Xaa	Ala	Xaa 75	Ser	Val	Leu	Tyr	Xaa 80
		Asp	Xaa	Ser	Xaa	Asn 85	Val	Xaa	Leu	Xaa	Lys 90	Xaa	Arg	Asn	Met	Val 95	Val
10		Xaa	Ala	Cys	Gly 100	Cys	His				•						
	(2)	INFO	RMAT	ON I	FOR S	SEQ :	ID NO	0:12	:								
15		(i)	(A) (B) (C)	LEN TYI	E CHA NGTH: PE: r RANDE	28 nucle EDNES	base ic a SS: s	e pa: acid sing!	irs		•						
50		(44)			POLOC			ır									
	,	(ii) iii)															
55	`	(iv)					,										
:0	•	(xi)	SEQU	JENCE	E DES	CRIE	OIT	l: SE	EQ II	NO:	12:						

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	GCGGATCCTG TTGTGAAGGN AATATGTG		28
	(2) INFORMATION FOR SEQ ID NO:13:		
5	<ul> <li>(i) SEQUENCE CHARACTERISTICS:</li> <li>(A) LENGTH: 24 base pairs</li> <li>(B) TYPE: nucleic acid</li> <li>(C) STRANDEDNESS: single</li> <li>(D) TOPOLOGY: linear</li> </ul>		
10	(ii) MOLECULE TYPE: cDNA		
	(iii) HYPOTHETICAL: NO		
15	(iv) ANTI-SENSE: NO		
20	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:		
20	GCGATCCGTC GCAGTCAAAA TTTT		24
	(2) INFORMATION FOR SEQ ID NO:14:		
25	<ul> <li>(i) SEQUENCE CHARACTERISTICS:</li> <li>(A) LENGTH: 26 base pairs</li> <li>(B) TYPE: nucleic acid</li> <li>(C) STRANDEDNESS: single</li> <li>(D) TOPOLOGY: linear</li> </ul>		
30	(ii) MOLECULE TYPE: cDNA		
	(iii) HYPOTHETICAL: NO		
35	(iv) ANTI-SENSE: NO		
•	(10, 11,11 51,155) 110		
40	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:	•	
	GCGGATCCGC GATATATTAA AAGCAA		26
	(2) INFORMATION FOR SEQ ID NO:15:		
45	<ul> <li>(i) SEQUENCE CHARACTERISTICS:</li> <li>(A) LENGTH: 20 base pairs</li> <li>(B) TYPE: nucleic acid</li> <li>(C) STRANDEDNESS: single</li> <li>(D) TOPOLOGY: linear</li> </ul>		
50			
	(ii) MOLECULE TYPE: cDNA		
	(iii) HYPOTHETICAL: NO		
55	(iv) ANTI-SENSE: NO		
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:		
60	CGGAATTCTG GTGCCATATA		20

PCT/US95/05467

## What is claimed is:

1 2	1.	analo	for identifying an OP-1 receptor-binding analog, said g being characterized as having substantially the same
3 4			ng affinity for a cell surface receptor as OP-1, the method ising the steps of:
5 6		(a)	providing a sample containing a protein selected from the group consisting of:
7 8 9	÷		<ul><li>(i) a polypeptide chain comprising an amino acid sequence defined by residues 16-123 of Seq. ID No. 3 (ALK-2), or an OP1-binding analog thereof;</li></ul>
10 11 12			<ul><li>(ii) a polypeptide chain comprising an amino acid sequence defined by residues 24-152 of Seq. ID No. 5 (ALK-3),, or a OP1-binding analog thereof;</li></ul>
13 14 15			(iii) a polypeptide chain comprising an amino acid sequence defined by residues 23-122 of Seq. ID No. 7 (ALK-6),, or a OP1 binding analog thereof;
16 17 18	·		(iv) a polypeptide chain having binding affinity for OP-1 and sharing at least 40% amino acid identity with residues 23- 122 of Seq. ID No. 7 (ALK-6),;
19 20 21 22			(v) a polypeptide chain having binding affinity for OP-1 and encoded by a nucleic acid obtainable by amplification with one or more primer sequences defined by Seq. ID Nos. 12-15 or
23 24 25 26			(vi) a polypeptide chain having binding affinity for OP-1 and encoded by a nucleic acid that hybridizes under stringent conditions with a nucleic acid comprising the sequence defined by nucleotides 256-552 of Seq. ID No. 7 (ALK-6),;
27 28	·	(b)	contacting said sample with a candidate OP1 receptor- binding analog; and
29 30		(c)	detecting specific binding between said candidate OP1 receptor- binding analog and said protein.
1 2 3 4	2.	analog bindir	nod for identifying an OP-1 receptor-binding analog, said being characterized as having substantially the same ag affinity for a cell surface receptor as OP1, the method sing the steps of:

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5		<ul><li>(a) providing a c</li></ul>	ell that expresses a surface receptor protein having
6		binding speci	ficity for OP-1 selected from the group consisting
7		of:	
8		(i) a polype	ptide chain comprising an amino acid sequence
9			by residues 16-123 of Seq. ID No. 3 (ALK-2), or an
10			ing analog thereof;
••		(44)	
11			peptide chain comprising an amino acid sequence
12			by residues 24-152 of Seq. ID No. 5 (ALK-3),, or an
13	•	OPI-BING	ing analog thereof;
14		(iii) a polyp	peptide chain comprising an amino acid sequence
15		defined 1	by residues 23-122 of Seq. ID No. 7 (ALK-6),, or an
16		OP1 bind	ing analog thereof;
17		(iv) a polyp	peptide chain having binding affinity for OP-1 and
18		sharing a	at least 40% amino acid identity with residues 23-
19		122 of S	eg. ID No. 7 (ALK-6),;
20		(v) a polyr	peptide chain having binding affinity for OP-1 and
21		encoded 1	by a nucleic acid obtainable by amplification with
22		one or m	ore primer sequences defined by Seq. ID Nos. 12-15;
23 -		or	·
24		(vi) a polyr	peptide chain having binding affinity for OP-1 and
25		encoded !	by a nucleic acid that hybridizes under stringent
26		condition	ns with a nucleic acid comprising the sequence
27		defined	by nucleotides 256-552 of Seq. ID No. 7 (ALK-6),;
28		(b) contacting sa	id cell with a candidate OP1 receptor-binding
29		analog; and	
30		(c) detecting ind	uction of an OP-1-mediated cellular response.
1	3.	The method of claim	2 wherein said OP-1 mediated cellular response
2		detected in step (c)	is induction of a kinase activity, inhibition of
3		epithelial cell grow	th, or induction of a cell differentiation
4		marker.	
1	4.	The method of claim	2 or 3 wherein said cell comprises a transfected
2		nucleic acid compris	sing a reporter gene in operative association with a
3		control element deri	ved from an OP-1 inducible protein.
1	5.	The method of any of	claims 1-4 wherein said sample further comprises
2		part or all of a Typ	e II serine/threonine kinase receptor protein
3		having binding affir	nity for OP-1, activin or BMP-4.

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1 2	6.	A method for producing an OP-1 receptor binding analog, the method comprising the steps of:
3 4		<ul><li>(a) obtaining, by the method of any of claims 1-5, a candidate OP-1 binding analog, and</li></ul>
5 6 7		(b) producing either said candidate analog or a second OP-1 binding analog derived from said candidate and having substantially the same OP-1 receptor-binding domain as said candidate.
1 2 3	7.	The method of producing an OP-1 receptor-binding analog of claim 6 wherein said analog produced in step (b) is by recombinant DNA techniques, or by nonbiological peptide synthesis.
1 2	8.	A kit for identifying OP-1 or a candidate OP-1 receptor binding analog in a sample, the kit comprising:
3 4		(a) a receptacle adapted to receive a sample and containing a protein selected from the group consisting of:
5 6 7		<ul><li>(i) a polypeptide chain comprising an amino acid sequence defined by residues 16-123 cf Seq. ID No. 3 (ALK-2), or an OP1-binding analog thereof;</li></ul>
9 10	•	(ii) a polypeptide chain comprising an amino acid sequence defined by residues 24-152 of Seq. ID No. 5 (ALK-3),, or an OP1-binding analog thereof;
11 12 13		(iii) a polypeptide chain comprising an amino acid sequence defined by residues 23-122 of Seq. ID No. 7 (ALK-6),, or an OP1 binding analog thereof;
14 15 16		(iv) a polypeptide chain having binding affinity for OP-1 and sharing at least 40% amino acid identity with residues 23- 122 of Seq. ID No. 7 (ALK-6),;
17 18 19 20		<ul> <li>(v) a polypeptide chain having binding affinity for OP-1 and encoded by a nucleic acid obtainable by amplification with one or more primer sequences defined by Seq. ID Nos. 12-15; or</li> </ul>
21 22 23 24 25		(vi) a polypeptide chain having binding affinity for OP-1 and encoded by a nucleic acid that hybridizes under stringent conditions with a nucleic acid comprising the sequence defined by nucleotides 256-552 of Seq. ID No. 7 (ALK-6),; and
26 27		(b) means for detecting interaction of OP-1 or a candidate OP-1 receptor-binding analog with said protein of part (a), said OP-1

28 29		or candidate analog comprising part of said sample provided to said receptacle.
1	9.	The kit of claim 8 wherein said means in part (b) comprises either
2 3		<ul><li>(i) means for detecting specific binding interaction of OP-1 or said candidate analog with said protein; or</li></ul>
· 4		(ii) means for detecting induction of an OP-1 mediated cellular response.
1 . 2	10.	The kit of claim 8 or 9 further comprising a serine/threonine Type II receptor having binding specificity for OP-1, activin or BMP-4.
1 2	11.	An OP-1 receptor-binding analog produced by the method of any of claims 1-7 or use of the kit of claims 8-10.
1	12.	The analog produced by the method of any of claims 1-8, said analog
3		(i) comprising an amino acid sequence sharing greater than 60% identity with the C-terminal 96 amino acids of the sequence represented by Seq. ID No. 9 (OP-1, residues 335-431), and
6		(ii) being substantially incapable of inducing an OP-1 mediated cellular response.
1 2	13.	The analog of claim 11 or 12 further having binding affinity for a Type II serine/threonine kinase cell surface receptor.
1 2	14.	The analog of claim 13 wherein said Type II receptor also has binding affinity for activin or BMP-4.
1 2 3 4 5	15.	An isolated ligand-receptor complex comprising two molecules interacting as specific binding partners, the first said molecule defining said ligand and comprising at least the C-terminal 96 amino acids of OP1 (residues 335-431 of Seq ID No. 9) or a receptor-binding analog thereof, and the second said molecule defining said receptor and being selected from the group consisting of:
7 8 9		<ul><li>(i) a polypeptide chain comprising an amino acid sequence defined by residues 16-123 of Seq. ID No. 3 (ALK-2), or an OP1-binding analog thereof;</li></ul>
10 11 12		<ul><li>(ii) a polypeptide chain comprising an amino acid sequence defined by residues 24-152 of Seq. ID No. 5 (ALK-3),, or an OP1-binding analog thereof;</li></ul>
13		(iii) a polypeptide chain comprising an amino acid sequence
14 15		defined by residues 23-122 of Seq. ID No. 7 (ALK-6),, or an OP1 binding analog thereof;

16		(iv) a polypeptide chain having binding affinity for OP-1 and
17		sharing at least 40% amino acid identity with residues 23-
18		122 of Seq. ID No. 7 (ALK-6),;
10		The of bod. In No. 1 (AMA 0777
19		(v) a polypeptide chain having binding affinity for OP-1 and
20		encoded by a nucleic acid obtainable by amplification with
21		one or more primer sequences defined by Seq. ID Nos. 12-15;
22		or
23		(vi) a polypeptide chain having binding affinity for OP-1 and
24		encoded by a nucleic acid that hybridizes under stringent
25		conditions with a nucleic acid comprising the sequence
26		defined by nucleotides 256-552 of Seq. ID No. 7 (ALK-6),.
1	16.	The complex of claim 15 further comprising part or all of a Type II
2		serine/threonine kinase receptor .
1	17.	The complex of claim 16 wherein said Type II receptor also has binding
2		affinity for activin or BMP-4.
	1.0	mb complement only of allows 15 17 changin and fineh malesman defining
1	18.	The complex of any of claims 15-17 wherein said first molecule defining
2		said ligand is an OP-1 receptor-binding analog comprises part or all
3		of the proteins selected from the group consisting of 60A, BMP-5, BMP-
4		6, Vgr-1, OP2, OP3 and receptor-binding amino acid sequence variants of
5		xenogenic homologs thereof.
1	19.	An isolated binding partner having specific binding affinity for an
2		epitope on a ligand-receptor complex, said complex being characterized
3		as comprising an OP-1 protein or an analog thereof in specific binding
4		interaction with the ligand binding domain of a cell surface receptor
5		defined by Seq. ID No. 3 (ALK-2), 5, or 7, or an OP1-binding analog
6		thereof; said binding partner having substantially no binding affinity
7		for the uncomplexed form of said OP-1 protein or OP-1 protein analog.
1	20.	The isolated binding partner of claim 19 wherein said binding partner
2		is further characterized as having substantially no binding affinity
3		for the uncomplexed form of said cell surface receptor protein or said
4		analog thereof.
1	21.	The binding partner of claim 19 wherein said binding partner is a
2	21.	monoclonal or polyclonal antibody.
1	22.	Use of the OP-1 receptor-binding analog of any claims 11-14 in a method
2	for	
3		(i) antagonizing OP-1 binding to a cell surface receptor; or
<b>A</b>		(ii) antagonizing induction of an OP-1 mediated cellular
4		
5		response.

comprises an antibody having binding specificity for  (i) the ligand binding domain of a cell surface receptor defi- by Seq. ID Nos. 3, 5, or 7 or an OP-1 binding analog thereof; or  (ii) the receptor binding domain of OP-1, represented by Seq. No. 9, or a receptor-binding analog thereof.  (i) a polypeptide chain comprising an amino acid sequence defined by residues 16-123 of Seq. ID No. 3 (ALK-2), or a OP1-binding analog thereof;  (ii) a polypeptide chain comprising an amino acid sequence defined by residues 24-152 of Seq. ID No. 5 (ALK-3), or OP1-binding analog thereof;  (iii) a polypeptide chain comprising an amino acid sequence defined by residues 23-122 of Seq. ID No. 7 (ALK-6), or OP1-binding analog thereof;  (iv) a polypeptide chain having binding affinity for OP-1 and sharing at least 40% amino acid identity with residues 23- 122 of Seq. ID No. 7 (ALK-6),;  (v) a polypeptide chain having binding affinity for OP-1 and encoded by a nucleic acid obtainable by amplification wite one or more primer sequences defined by Seq. ID Nos. 12-1 or  (vi) a polypeptide chain having binding affinity for OP-1 and encoded by a nucleic acid that hybridizes under stringen conditions with a nucleic acid comprising the sequence defined by nucleotides 256-552 of Seq. ID No. 7 (ALK-6),;  in a method for antagonizing  (i) OP-1 binding to a cell surface receptor; or (ii) induction of an OP-1 mediated cellular response.  25. A method comprising the step of:			
by Seq. ID Nos. 3, 5, or 7 or an OP-1 binding analog thereof; or  (ii) the receptor binding domain of OP-1, represented by Seq. No. 9, or a receptor-binding analog thereof.  24. Use of a protein selected from the group consisting of:  (i) a polypeptide chain comprising an amino acid sequence defined by residues 16-123 of Seq. ID No. 3 (ALK-2), or a OP1-binding analog thereof;  (ii) a polypeptide chain comprising an amino acid sequence defined by residues 24-152 of Seq. ID No. 5 (ALK-3), or OP1-binding analog thereof;  (iii) a polypeptide chain comprising an amino acid sequence defined by residues 23-122 of Seq. ID No. 7 (ALK-6), or OP1 binding analog thereof;  (iv) a polypeptide chain comprising an amino acid sequence defined by residues 23-122 of Seq. ID No. 7 (ALK-6), or OP1 binding analog thereof;  (iv) a polypeptide chain having binding affinity for OP-1 and sharing at least 40% amino acid identity with residues 23 122 of Seq. ID No. 7 (ALK-6),;  (v) a polypeptide chain having binding affinity for OP-1 and encoded by a nucleic acid obtainable by amplification wit one or more primer sequences defined by Seq. ID Nos. 12-1 or  (vi) a polypeptide chain having binding affinity for OP-1 and encoded by a nucleic acid that hybridizes under stringen conditions with a nucleic acid comprising the sequence defined by nucleotides 256-552 of Seq. ID No. 7 (ALK-6),;  in a method for antagonizing  (i) OP-1 binding to a cell surface receptor; or  (ii) induction of an OP-1 mediated cellular response.  A method for antagonizing activin binding to a cell surface receptor, the method comprising the step of:  providing a cell expressing a said receptor with a protein hav binding specificity for the amino acid sequence defined by residues 16-123 of Seq ID No. 3 or an OP-1 binding sequence	2		comprises an antibody having binding specificity for
No. 9, or a receptor-binding analog thereof.  24. Use of a protein selected from the group consisting of:  (i) a polypeptide chain comprising an amino acid sequence defined by residues 16-123 of Seq. ID No. 3 (ALK-2), or a OP1-binding analog thereof;  (ii) a polypeptide chain comprising an amino acid sequence defined by residues 24-152 of Seq. ID No. 5 (ALK-3),, or OP1-binding analog thereof;  (iii) a polypeptide chain comprising an amino acid sequence defined by residues 23-122 of Seq. ID No. 7 (ALK-6),, or OP1-binding analog thereof;  (iv) a polypeptide chain having binding affinity for OP-1 and sharing at least 40% amino acid identity with residues 23-122 of Seq. ID No. 7 (ALK-6),;  (v) a polypeptide chain having binding affinity for OP-1 and encoded by a nucleic acid obtainable by amplification with one or more primer sequences defined by Seq. ID Nos. 12-1 or  (vi) a polypeptide chain having binding affinity for OP-1 and encoded by a nucleic acid comprising the sequence defined by nucleotides 256-552 of Seq. ID No. 7 (ALK-6),;  in a method for antagonizing  (i) OP-1 binding to a cell surface receptor; or  (ii) induction of an OP-1 mediated cellular response.  A method for antagonizing activin binding to a cell surface receptor, the method comprising the step of:  providing a cell expressing a said receptor with a protein hav binding specificity for the amino acid sequence defined by residues 16-123 of Seq ID No. 3 or an OP-1 binding sequence	4		<del>-</del>
(i) a polypeptide chain comprising an amino acid sequence defined by residues 16-123 of Seq. ID No. 3 (ALK-2), or a OP1-binding analog thereof;  (ii) a polypeptide chain comprising an amino acid sequence defined by residues 24-152 of Seq. ID No. 5 (ALK-3), or OP1-binding analog thereof;  (iii) a polypeptide chain comprising an amino acid sequence defined by residues 23-122 of Seq. ID No. 7 (ALK-6), or OP1 binding analog thereof;  (iv) a polypeptide chain having binding affinity for OP-1 and sharing at least 40% amino acid identity with residues 23-122 of Seq. ID No. 7 (ALK-6),  (v) a polypeptide chain having binding affinity for OP-1 and encoded by a nucleic acid obtainable by amplification with one or more primer sequences defined by Seq. ID Nos. 12-1 or  (vi) a polypeptide chain having binding affinity for OP-1 and encoded by a nucleic acid that hybridizes under stringen conditions with a nucleic acid comprising the sequence defined by nucleotides 256-552 of Seq. ID No. 7 (ALK-6),;  in a method for antagonizing  (i) OP-1 binding to a cell surface receptor; or  (ii) induction of an OP-1 mediated cellular response.  25. A method for antagonizing activin binding to a cell surface receptor, the method comprising the step of:  providing a cell expressing a said receptor with a protein hav binding specificity for the amino acid sequence defined by residues 16-123 of Seq ID No. 3 or an OP-1 binding sequence			(ii) the receptor binding domain of OP-1, represented by Seq. ID No. 9, or a receptor-binding analog thereof.
defined by residues 16-123 of Seq. ID No. 3 (ALK-2), or a OP1-binding analog thereof;  (ii) a polypeptide chain comprising an amino acid sequence defined by residues 24-152 of Seq. ID No. 5 (ALK-3),, or OP1-binding analog thereof;  (iii) a polypeptide chain comprising an amino acid sequence defined by residues 23-122 of Seq. ID No. 7 (ALK-6),, or OP1 binding analog thereof;  (iv) a polypeptide chain having binding affinity for OP-1 and sharing at least 40% amino acid identity with residues 23 122 of Seq. ID No. 7 (ALK-6),;  (v) a polypeptide chain having binding affinity for OP-1 and encoded by a nucleic acid obtainable by amplification with one or more primer sequences defined by Seq. ID Nos. 12-1 or  (vi) a polypeptide chain having binding affinity for OP-1 and encoded by a nucleic acid that hybridizes under stringen conditions with a nucleic acid comprising the sequence defined by nucleotides 256-552 of Seq. ID No. 7 (ALK-6),;  in a method for antagonizing  (i) OP-1 binding to a cell surface receptor; or  (ii) induction of an OP-1 mediated cellular response.  25. A method for antagonizing activin binding to a cell surface receptor, the method comprising the step of:  providing a cell expressing a said receptor with a protein hav binding specificity for the amino acid sequence defined by residues 16-123 of Seq ID No. 3 or an OP-1 binding sequence	1	24.	Use of a protein selected from the group consisting of:
defined by residues 24-152 of Seq. ID No. 5 (ALK-3),, or OP1-binding analog thereof;  (iii) a polypeptide chain comprising an amino acid sequence defined by residues 23-122 of Seq. ID No. 7 (ALK-6),, or OP1 binding analog thereof;  (iv) a polypeptide chain having binding affinity for OP-1 and sharing at least 40% amino acid identity with residues 23 122 of Seq. ID No. 7 (ALK-6),;  (v) a polypeptide chain having binding affinity for OP-1 and encoded by a nucleic acid obtainable by amplification with one or more primer sequences defined by Seq. ID Nos. 12-1 or  (vi) a polypeptide chain having binding affinity for OP-1 and encoded by a nucleic acid that hybridizes under stringen conditions with a nucleic acid comprising the sequence defined by nucleotides 256-552 of Seq. ID No. 7 (ALK-6),;  in a method for antagonizing  (i) OP-1 binding to a cell surface receptor; or  (ii) induction of an OP-1 mediated cellular response.  25. A method for antagonizing activin binding to a cell surface receptor, the method comprising the step of:  providing a cell expressing a said receptor with a protein hav binding specificity for the amino acid sequence defined by residues 16-123 of Seq ID No. 3 or an OP-1 binding sequence	3		defined by residues 16-123 of Seq. ID No. 3 (ALK-2), or an
defined by residues 23-122 of Seq. ID No. 7 (ALK-6),, or OP1 binding analog thereof;  (iv) a polypeptide chain having binding affinity for OP-1 and sharing at least 40% amino acid identity with residues 23 122 of Seq. ID No. 7 (ALK-6),;  (v) a polypeptide chain having binding affinity for OP-1 and encoded by a nucleic acid obtainable by amplification with one or more primer sequences defined by Seq. ID Nos. 12-1 or  (vi) a polypeptide chain having binding affinity for OP-1 and encoded by a nucleic acid that hybridizes under stringen conditions with a nucleic acid comprising the sequence defined by nucleotides 256-552 of Seq. ID No. 7 (ALK-6),;  in a method for antagonizing  (i) OP-1 binding to a cell surface receptor; or  (ii) induction of an OP-1 mediated cellular response.  A method for antagonizing activin binding to a cell surface receptor, the method comprising the step of:  providing a cell expressing a said receptor with a protein hav binding specificity for the amino acid sequence defined by residues 16-123 of Seq ID No. 3 or an OP-1 binding sequence	6		defined by residues 24-152 of Seq. ID No. 5 (ALK-3),, or an
sharing at least 40% amino acid identity with residues 23  122 of Seq. ID No. 7 (ALK-6);  (v) a polypeptide chain having binding affinity for OP-1 and encoded by a nucleic acid obtainable by amplification with one or more primer sequences defined by Seq. ID Nos. 12-1  or  (vi) a polypeptide chain having binding affinity for OP-1 and encoded by a nucleic acid that hybridizes under stringen conditions with a nucleic acid comprising the sequence defined by nucleotides 256-552 of Seq. ID No. 7 (ALK-6);  in a method for antagonizing  (i) OP-1 binding to a cell surface receptor; or  (ii) induction of an OP-1 mediated cellular response.  25. A method for antagonizing activin binding to a cell surface receptor, the method comprising the step of:  providing a cell expressing a said receptor with a protein hav binding specificity for the amino acid sequence defined by residues 16-123 of Seq ID No. 3 or an OP-1 binding sequence	9		defined by residues 23-122 of Seq. ID No. 7 (ALK-6),, or an
encoded by a nucleic acid obtainable by amplification with one or more primer sequences defined by Seq. ID Nos. 12-1 or  (vi) a polypeptide chain having binding affinity for OP-1 and encoded by a nucleic acid that hybridizes under stringen conditions with a nucleic acid comprising the sequence defined by nucleotides 256-552 of Seq. ID No. 7 (ALK-6),;  in a method for antagonizing  (i) OP-1 binding to a cell surface receptor; or  (ii) induction of an OP-1 mediated cellular response.  A method for antagonizing activin binding to a cell surface receptor, the method comprising the step of:  providing a cell expressing a said receptor with a protein hav binding specificity for the amino acid sequence defined by residues 16-123 of Seq ID No. 3 or an OP-1 binding sequence	12		sharing at least 40% amino acid identity with residues 23-
encoded by a nucleic acid that hybridizes under stringen conditions with a nucleic acid comprising the sequence defined by nucleotides 256-552 of Seq. ID No. 7 (ALK-6),;  in a method for antagonizing  (i) OP-1 binding to a cell surface receptor; or  (ii) induction of an OP-1 mediated cellular response.  A method for antagonizing activin binding to a cell surface receptor, the method comprising the step of:  providing a cell expressing a said receptor with a protein hav binding specificity for the amino acid sequence defined by residues 16-123 of Seq ID No. 3 or an OP-1 binding sequence	L5 L6		encoded by a nucleic acid obtainable by amplification with one or more primer sequences defined by Seq. ID Nos. 12-15;
(i) OP-1 binding to a cell surface receptor; or  (ii) induction of an OP-1 mediated cellular response.  25. A method for antagonizing activin binding to a cell surface receptor, the method comprising the step of:  providing a cell expressing a said receptor with a protein hav binding specificity for the amino acid sequence defined by residues 16-123 of Seq ID No. 3 or an OP-1 binding sequence	(9 20		encoded by a nucleic acid that hybridizes under stringent conditions with a nucleic acid comprising the sequence
(ii) induction of an OP-1 mediated cellular response.  25. A method for antagonizing activin binding to a cell surface receptor, the method comprising the step of:  3 providing a cell expressing a said receptor with a protein hav binding specificity for the amino acid sequence defined by residues 16-123 of Seq ID No. 3 or an OP-1 binding sequence	22		in a method for antagonizing
25. A method for antagonizing activin binding to a cell surface receptor, the method comprising the step of:  3 providing a cell expressing a said receptor with a protein hav binding specificity for the amino acid sequence defined by residues 16-123 of Seq ID No. 3 or an OP-1 binding sequence	!3		(i) OP-1 binding to a cell surface receptor; or
the method comprising the step of:  providing a cell expressing a said receptor with a protein hav binding specificity for the amino acid sequence defined by residues 16-123 of Seq ID No. 3 or an OP-1 binding sequence	4		(ii) induction of an OP-1 mediated cellular response.
binding specificity for the amino acid sequence defined by residues 16-123 of Seq ID No. 3 or an OP-1 binding sequence		25.	A method for antagonizing activin binding to a cell surface receptor, the method comprising the step of:
	4 5		residues 16-123 of Seq ID No. 3 or an OP-1 binding sequence

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7		sequence identity with residue 335-431 of the sequence defined by
8		Seq ID No. 9,
9		such that said protein, when provided to said cell, is competent
10		to interact specifically with said receptor, thereby
11		substantially inhibiting activin binding to said receptor.
1	26.	A method for antagonizing BMP-4 binding to a cell surface receptor, the
2		method comprising the step of:
3		providing a cell expressing a said receptor with a protein having
4		binding specificity for the ligand binding domain defined by
5		residues 24-152 of Seq ID No. 5 (ALK-3), or residues 23-122 of
6		Seq ID No. 7 (ALK-6), or an OP-1 binding sequence variant
7		thereof, said protein sharing at least 60% amino acid sequence
8		identity with residues 335-431 of the sequence defined by Seq ID
9		No. 9,
10		such that said protein, when provided to said cell, is competent
11		to interact specifically with said receptor, thereby
12		substantially inhibiting BMP-4 binding to said receptor.
1	27.	Use of the OP-1 receptor binding analog of claim 12-14 in the method of
2 .		claim 25 or 26

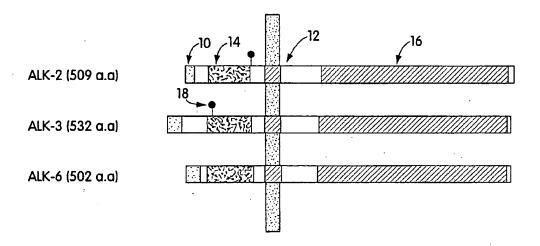


Fig. 1

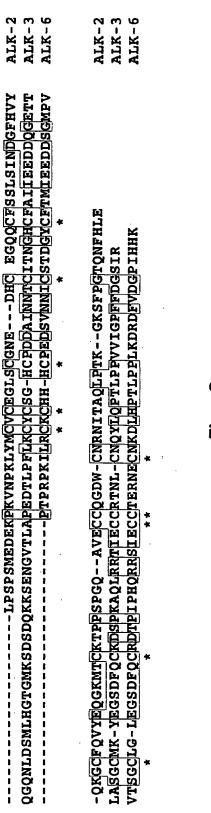


Fig. 2

**SUBSTITUTE SHEET (RULE 26)** 

	-RKFKRRNQERLNPPDVEYGTIEGLITTNVGDSHLADLLDHSCTSGSGSGLPFLVQRHVARQITLLECVGKGRYG	ALK-2
:	KSISSRRRYNRDLEQDEAFIPVGES-LKDLIDQSQSSGSGSGLPLLVQRTIAKQIQMVRQVGKGRYG	ALK-3
	FRYKROEARPRYSIGLEODETYIPPGES-LRDLIEQSOSSGSGSGLPLLVORTIAKQIQMVKQIGKGRYG	ALK-6
	ness to contract assessment of the first first of the fir	:
S	EVWKGDWKJGENVAVKLFSSK	ALK-2
U	EVWMGKWRGEKVAVKVFFTT	ALK-3
BS	EVWMGKWRGEKVAVKVFFTTEEASWFRETEIYQTVLMRHENILGFIAADIKGTGSWTQLYLITDYHENGSLYDYL	ALK-4
Π	QLTFLDTVSCERIVLSIASGLAHLHETFGTQGKPAIAHRDLKSKNILVKKNGQCCIADLGLAVMHSQSTNQLDV	ALK-2
U	KCATLDTRALLKLAYSAACGLCHLHTEIYGTQGKPAIAHRDLKSKNILIKKNGSCCIADLGLAVKFNSDTNEVDV	ALK-3
ΪE	KSTTLDAKSMUKLAYSSVSGLCHLHTEIFSTQGKPAIAHRDLKSKNILVKKNGTCCIADLGLAVKFISDTNEVDI	ALK-6
SH	GNNPRVGTKRYMA DEVI.DEPTOVDCEDSYKRVDIWARGINTMENARRWYSNGIVEDYK PPFYDVVBNDPSFEDMR	AI.K-2
EE		ALK-3
:T (		ALK-6
R		
UŁ		ALK-2
E	I EVVCVKRLRPIVSNRWMSDECLPAVLKLMSECWAHNPASRLTALRIKKTLAKMVESQDVKI	ALK-3
26	EIVCMKKLRPSFPNRWSSDECLROMGKLMTECWAONPASRLTALRVKKTLAKMSESODIKL	ALK-6
)		

Fig. 3